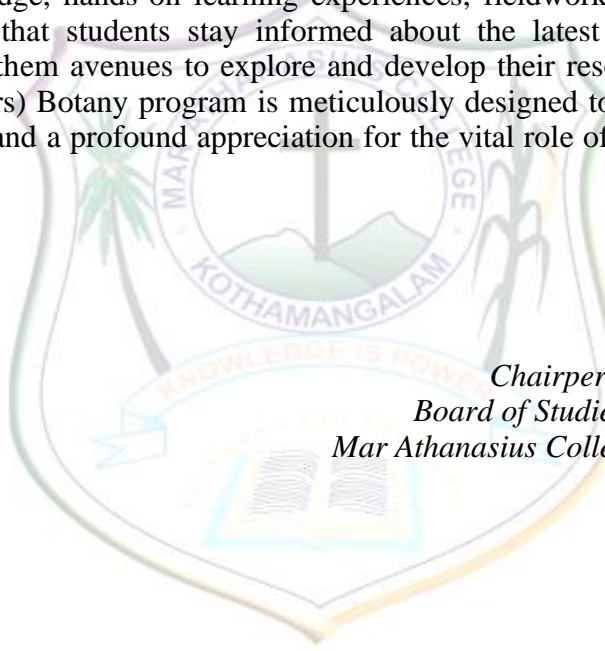


## Preface

The four-year undergraduate program in Botany is structured in accordance with the guidelines set by the Department of Higher Education, Government of Kerala, and the National Education Policy. Its primary objective is to impart students with a comprehensive understanding of plant science.

The curriculum and syllabus of the BSc (Honours) Botany program is crafted to equip students with essential knowledge and skills to navigate the complexities of the plant world. With a specialization in Plant Biotechnology, the syllabus encompasses various facets of plant life, spanning from sub-microscopic structures to vast ecosystems that influence our environment. The holistic curriculum includes a balanced mix of theoretical knowledge, hands-on learning experiences, fieldwork, and case studies. This approach ensures that students stay informed about the latest advancements in plant sciences, offering them avenues to explore and develop their research interests. In short, the B. Sc. (Honours) Botany program is meticulously designed to foster critical thinking, scientific inquiry, and a profound appreciation for the vital role of plants in sustaining life on Earth.



*Chairperson and Members  
Board of Studies in Botany (UG)  
Mar Athanasius College (Autonomous)  
Kothamangalam*

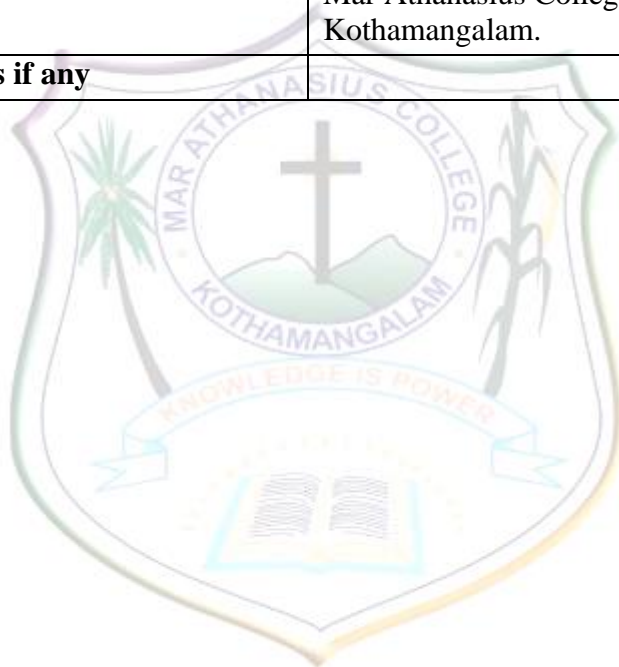
**MAR ATHANASIVS COLLEGE (AUTONOMOUS), KOTHAMANGALAM**

**Members of the Board of Studies - UG Programme**

**Subject: BOTANY**

<b>Chairperson</b>	<b>Dr. Siju Thomas T</b> Assistant Professor and Head Department of Botany, Mar Athanasius College (Autonomous), Kothamangalam.
<b>Experts (2) (Outside University)</b>	<b>1. Dr. Dennis Thomas T</b> Professor, Department of Plant Science, Central University of Kerala, Kasargod.
	<b>2. Dr. Santhosh Nampy</b> Professor, Department of Botany, University of Calicut, Thenhipalam, Malappuram.
<b>One Expert - nominated by VC (M. G. University)</b>	<b>Dr. E. A. Siril</b> Professor and Head, Department of Botany, University of Kerala, Kariavattom, Thiruvananthapuram
<b>Member from Industry</b>	<b>Dr. Safeer P. M</b> Director, CIRIST Ecosystem Pvt. Ltd., Kinfra Hitech Park, Kalamassery, Ernakulam.
<b>Meritorious Alumnus</b>	<b>Dr. Giby Kuriakose</b> Assistant Professor, Department of Botany, Sacred Heart College (Autonomous), Thevara, Kochi.
<b>Other Members of the Department</b>	<b>1. Dr. Aji Abraham</b> Associate Professor, Mar Athanasius College (Autonomous), Kothamangalam.
	<b>2. Ms. Meril Sara Kurian</b> Assistant Professor, Mar Athanasius College (Autonomous), Kothamangalam.
	<b>3. Mr. Sarath G Nair</b>

	Assistant Professor, Mar Athanasius College (Autonomous), Kothamangalam.
	<b>4. Dr. Jayalakshmi P. S.</b> Assistant Professor, Mar Athanasius College (Autonomous), Kothamangalam
	<b>5. Dr. Akhila Sen</b> Assistant Professor, Mar Athanasius College (Autonomous), Kothamangalam
	<b>6. Dr. Dhannia P Narayanan</b> Assistant Professor on contract, Mar Athanasius College (Autonomous), Kothamangalam.
<b>Special Invitees if any</b>	



## **Programme Outcomes (PO)**

### **PO 1: Critical thinking and Analytical reasoning**

Capability to analyse and evaluate evidence, arguments, claims, beliefs on the basis of empirical evidence; identify relevant assumptions or implications; formulate coherent arguments; critically evaluate practices, policies and theories to develop knowledge and understanding; critical sensibility to lived experiences, with self awareness and reflexivity of both self and society.

### **PO 2: Scientific reasoning and Problem solving**

Ability to analyse, interpret and draw conclusions from quantitative/qualitative data; and critically evaluate ideas, evidence and experiences from an open-minded and reasoned perspective; capacity to extrapolate from what one has learned and apply their competencies to solve different kinds of non-familiar problems, rather than replicate curriculum content knowledge; and apply one's learning to real life situations.

### **PO 3: Multidisciplinary/interdisciplinary/transdisciplinary Approach**

Acquire interdisciplinary /multidisciplinary/transdisciplinary knowledge base as a consequence of the learning they engage with their programme of study; develop a collaborative-multidisciplinary/interdisciplinary/transdisciplinary- approach for formulate constructive arguments and rational analysis for achieving common goals and objectives.

### **PO 4: Communication Skills**

Ability to express thoughts and ideas effectively in writing and orally; Communicate with others using appropriate media; confidently share one's views and express herself/himself; demonstrate the ability to listen carefully, read and write analytically, and present complex information in a clear and concise manner to different groups.

### **PO 5: Leadership and Entrepreneurship Skills**

Ability to work effectively and lead respectfully with diverse teams; setting direction, formulating an inspiring vision, building a team who can help achieve the vision, motivating and inspiring team members to engage with that vision, and using management skills to guide people to the right destination, in a smooth and efficient way. After inculcating all the necessary graduate qualities, a graduate can become an entrepreneur.

### **PO 6: Social Consciousness and Responsibility**

Ability to contemplate the impact of research findings on conventional practices, and a clear understanding of responsibility towards societal needs and reaching the targets for attaining inclusive and sustainable development.

**PO 7: Equity, Inclusiveness and Sustainability**

Appreciate equity, inclusiveness and sustainability and diversity; acquire ethical and moral reasoning and values of unity, secularism and national integration to enable to act as dignified citizens; able to understand and appreciate diversity (caste, ethnicity, gender and marginalization), managing diversity and use of an inclusive approach to the extent possible.

**PO 8: Moral and Ethical Reasoning**

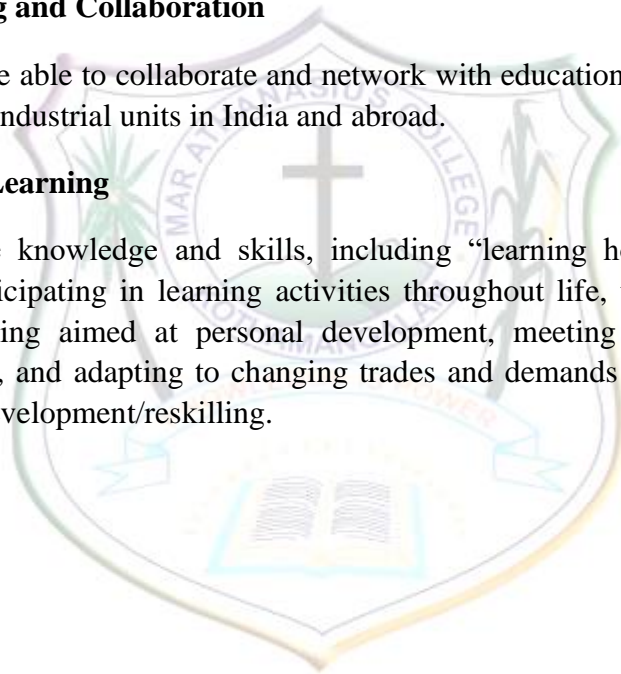
Ability to embrace moral/ethical values in conducting one's life, formulate a position/argument about an ethical issue from multiple perspectives, and use ethical practices in all work. Capable of demonstrating the ability to identify ethical issues related to one's work, avoid unethical behavior.

**PO 9: Networking and Collaboration**

Acquire skills to be able to collaborate and network with educational institutions, research organisations and industrial units in India and abroad.

**PO 10: Lifelong Learning**

Ability to acquire knowledge and skills, including "learning how to learn", that are necessary for participating in learning activities throughout life, through self-paced and self-directed learning aimed at personal development, meeting economic, social and cultural objectives, and adapting to changing trades and demands of work place through knowledge/skill development/reskilling.



### Programme Specific Outcome

Upon completion of the B.Sc. (Honors) Botany Programme, the graduates will be able to:

<b>Sl. No.</b>	<b>Programme Specific Outcomes</b>	<b>PO</b>
1	Identify and differentiate major plant groups by analysing morphological and anatomical features.	1,2,7,10
2	Understand the ethical considerations, social consciousness and responsibilities associated with conservation of nature and sustainable practices.	6,7,8,10
3	Analyze the metabolic processes related to the growth, development, physiology, and reproduction of plants, and their applications in designing and interpreting scientific experiments.	1,2,3,9
4	Acquire computational skills, practical proficiency in laboratory techniques, fieldwork, and experimental methodologies in Botany, fostering employability.	2,5,4,9
5	Identify the interdisciplinary scope and applications of plant science and explore entrepreneurial skills.	1,3,5,9
6	Communicate scientific ideas and research findings through oral presentations, written reports, and scientific publications.	4,9,6,10

## SCHEME OF INSTRUCTIONAL CREDITS AND HOURS

No	Semester	Course Title		Course Type	Credit	Hrs/Week	Total Hours
1	1	Introduction to Plant Science and Applied Botany		DSC A	4	5	90
2	1	Ecotourism		MDC	3	4	72
3	2	Plant Resources and Avenues in Botany		DSC A	4	5	90
4	2	Gardening and landscaping		MDC	3	4	72
5	3	Microbiology and Phycology		DSC A	4	5	90
6	3	Mycology, Lichenology and Crop Pathology		DSC A	4	5	90
7	3	Analytical Techniques in Plant Sciences (S)	Any One	DSE	4	4	72
8	3	Horticulture and Post Harvest Technology					
9	3	Angiosperm Taxonomy and Economic Botany		DSC B	4	5	90
10	3	Agri based Micro Enterprises		MDC	3	3	54
11	3	Bioethics and IPR		VAC	3	3	54
12	4	Archegoniates		DSC A	4	5	90
13	4	Plant Cell and Molecular Biology		DSC A	4	5	90
14	4	Introduction to Biotechnology (S)	Any One	DSE	4	4	72
15	4	Phytotechnology					
16	4	Plant Anatomy and Microtechnique		DSC B	4	5	90
17	4	Biofertilizers and Biocontrol Agents		SEC	3	3	54
18	4	Conservation biology and Sustainable Development		VAC	3	3	54
19	4	Internship		INT	2		
20	5	Angiosperm Systematics and Economic Botany		DSC	4	5	90
21	5	Plant Physiology and Biochemistry		DSC	4	5	90
22	5	Plant Biotechnology and Introduction to Bioinformatics (S)	Any Three	DSE	4	4	72
23	5	Plant Tissue Culture					
24	5	Bioactive Phytochemicals					
25	5	Food Science and Quality Control					

26	5	Mushroom Cultivation and Value Addition		SEC	3	3	54
27	6	Plant Anatomy and Developmental Biology		DSC	4	5	90
28	6	Research Methodology, Biostatistics and Computer Application		DSC	4	5	90
29	6	Microbial Biotechnology (S)		DSE	4	5	90
30	6	Genetics and Evolution	<i>Any One</i>	DSE	4	4	72
31	6	Phytogeography, Forestry and Ecotourism					
32	6	Entrepreneurial Botany		SEC	3	3	54
33	6	Environmental Science and Human Rights		VAC	3	3	54
34	7	Modern Trends in Thallophytes		DCC	4	5	90
35	7	Plant Growth and Metabolism		DCC	4	4	72
36	7	Advances in Molecular Cell Biology and Immunology		DCC	4	4	72
37	7	Ecology and Environmental conservation	<i>Any Three</i>	DCE	4	4	72
38	7	Conventional and Modern approaches in Plant Breeding					
39	7	Seed Technology					
40	7	Agroecology					
41	8	Modern Trends in Archegoniates		DCC	4	5	90
42	8	Modern Trends in Plant Systematics		DCC	4	5	90
43	8	Computational Biology and Omics in Plant Sciences		DCE	4	5	90
44	8	Methods in Research, Biophysics and Biological Techniques		DCE	4	5	90
45	8	Phytochemistry and Pharmacognosy		DCE	4	5	90
46	8	Aquatic Botany		DCE	4	5	90
47	8	Project		PRJ	12/8		



## Syllabus Index: Botany Major

### Name of the Major Subject: Botany (Specialisation in Plant Biotechnology)

#### Semester 1

Course Code	Title of the Course	Type of the Course	Credit	Hours/Week	Hour distribution/week			
					L	T	P	O
M24BO1DSC100	Introduction to Plant Science and Applied Botany	DSC A	4	5	3	-	2	-
M24BO1MDC100	Ecotourism	MDC	3	4	2	-	2	-

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others

#### Semester 2

Course Code	Title of the Course	Type of the Course	Credit	Hours/Week	Hour distribution/week			
					L	T	P	O
M24BO2DSC100	Plant Resources and Avenues in Botany	DSC A	4	5	3	-	2	-
M24BO2MDC100	Gardening and landscaping	MDC	3	4	2	-	2	-

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others

#### Semester 3

Course Code	Title of the Course	Type of the Course	Credit	Hours/Week	Hour distribution/week			
					L	T	P	O
M24BO3DSC200	Microbiology and Phycology	DSC A	4	5	3	-	2	-
M24BO3DSC201	Mycology, Lichenology and Crop Pathology	DSC A	4	5	3	-	2	-
M24BO3DSE200	Analytical Techniques in Plant Sciences (S)	Any One DSE	4	4	4	-	-	-
M24BO3DSE201	Horticulture and Post Harvest Technology							
M24BO3DSC202	Angiosperm Taxonomy and Economic Botany	DSC B	4	5	3	-	2	-
M24BO3MDC200	Agri based Micro Enterprises	MDC	3	3	3	-	-	-
M24BO3VAC200	Bioethics and IPR	VAC	3	3	3	-	-	-

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others, S- Specialisation

### Semester 4

Course Code	Title of the Course		Type of the Course	Credit	Hours/Week	Hour distribution/week			
						L	T	P	O
M24BO4DSC200	Archegoniates		DSC A	4	5	3	-	2	-
M24BO4DSC201	Plant Cell and Molecular Biology		DSC A	4	5	3	-	2	-
M24BO4DSE200	Introduction to Biotechnology (S)	Any One	DSE	4	4	4	-	-	-
M24BO4DSE201	Phytotechnology								
M24BO4DSC202	Plant Anatomy and Microtechnique	DSC B	4	5	3	-	2	-	
M24BO4SEC200	Biofertilizers and Biocontrol Agents		SEC	3	3	3	-	-	-
M24BO4VAC200	Conservation biology and Sustainable Development		VAC	3	3	3	-	-	-
M24BO4INT200	Internship			2					

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others, S- Specialisation

### Semester 5

Course Code	Title of the Course		Type of the Course	Credit	Hours/Week	Hour distribution/week			
						L	T	P	O
M24BO5DSC300	Angiosperm Systematics and Economic Botany		DSC	4	5	3	-	2	-
M24BO5DSC301	Plant Physiology and Biochemistry		DSC	4	5	3	-	2	-
M24BO5DSE300	Plant Biotechnology and Introduction to Bioinformatics (S)	Any Three	DSE	4	4	4	-	-	-
M24BO5DSE301	Plant Tissue Culture								
M24BO5DSE302	Bioactive Phytochemicals								
M24BO5DSE303	Food Science and Quality Control								
M24BO5SEC300	Mushroom Cultivation and Value Addition		SEC	3	3	3	-	-	-

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others, S- Specialisation

### Semester 6

Course Code	Title of the Course	Type of the Course	Credit	Hours/Week	Hour distribution/week			
					L	T	P	O
M24BO6DSC300	Plant Anatomy and Developmental Biology	DSC	4	5	3	-	2	-
M24BO6DSC301	Research Methodology, Biostatistics and Computer Application	DSC	4	5	3	-	2	-
M24BO6DSE300	Microbial Biotechnology (S)	DSE	4	5	3	-	2	-
M24BO6DSE301	Genetics and Evolution	Any One	DSE	4	4	-	-	-
M24BO6DSE302	Phytogeography, Forestry and Ecotourism							
M24BO6SEC300	Entrepreneurial Botany	SEC	3	3	3	-	-	-
M24BO6VAC300	Environmental Science and Human Rights	VAC	3	3	3	-	-	-

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others, S- Specialisation

### Semester 7

Course Code	Title of the Course	Type of the Course	Credit	Hours/Week	Hour distribution/week			
					L	T	P	O
M24BO7DCC400	Modern Trends in Thallophtyes	DCC	4	5	3	-	2	-
M24BO7DCC401	Plant Growth and Metabolism	DCC	4	4	-	-	-	-
M24BO7DCC402	Advances in Molecular Cell Biology and Immunology	DCC	4	4	-	-	-	-
M24BO7DCE400	Ecology and Environmental conservation	Any Three	DCE	4	4	-	-	-
M24BO7DCE401	Conventional and Modern approaches in Plant Breeding							
M24BO7DCE402	Seed Technology							
M24BO7DCE403	Agroecology							

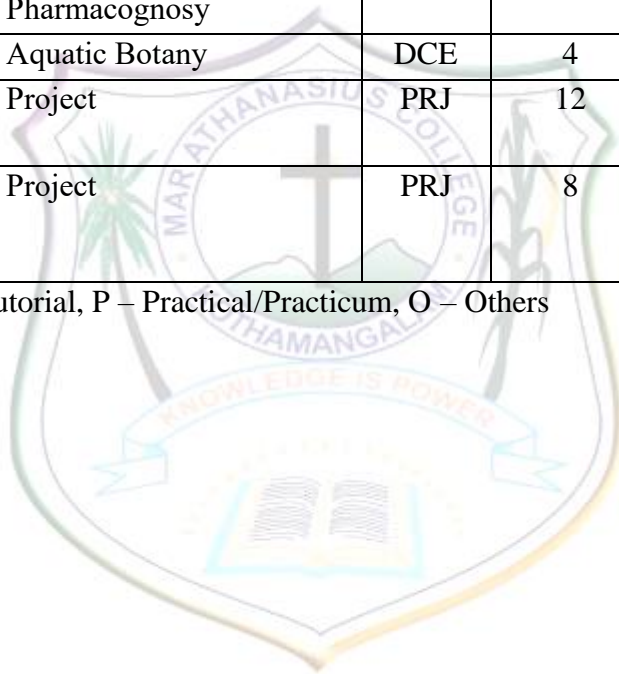
L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others

### Semester 8

Course Code	Title of the Course	Type of	Credit	Hours/	Hour
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		<i>the Course</i>		<i>Week</i>	<i>distribution/ week</i>			
					L	T	P	O
M24BO8DCC400	Modern Trends in Archegoniates	DCC	4	5	3	-	2	-
M24BO8DCC401	Modern Trends in Plant Systematics	DCC	4	5	3	-	2	-
M24BO8DCE400	Computational Biology and Omics in Plant Sciences	DCE	4	5	3	-	2	-
M24BO8DCE401	Methods in Research, Biophysics and Biological Techniques	DCE	4	5	3	-	2	-
M24BO8DCE402	Phytochemistry and Pharmacognosy	DCE	4	5	3	-	2	-
M24BO8DCE403	Aquatic Botany	DCE	4	5	3	-	2	-
M24BO8PRJ400	Project	PRJ	12	Honours with Research - 2 DCC + Project				
M24BO8PRJ401	Project	PRJ	8	Honours – 2 DCC + 1 DSC/DCE + Project OR 2 DCE				

L – Lecture, T – Tutorial, P – Practical/Practicum, O – Others



**Courses which have Study tour/ Field visit / Institution visit**


<i>No</i>	<i>Semester</i>	<i>Course Type</i>	<i>Course Title</i>	<i>Type of Activity</i>
1	1	DSC A	Introduction to Plant Science and Applied Botany	Field visit (1 day)
2	1	MDC	Ecotourism	Field visit (1 day)
3	2	DSC A	Plant Resources and Avenues in Botany	Industry visit (1 day)
4	2	MDC	Gardening and landscaping	Field visit (1 day)
5	3	DSC A	Microbiology and Phycology	Field visit (1 day)
6	3	DSC A	Mycology, Lichenology and Crop Pathology	Field visit (1 day)
7	3	DSE	Analytical Techniques in Plant Sciences	Institution visit (One day)
8	3	DSE	Horticulture and Post Harvest Technology	Visit to garden (1 day)
9	3	MDC	Agri based Micro Enterprises	Visit to lab/ Garden/ Agri- entrepreneur (1 day)
10	3	DSC A	Archegoniatas	Field visit (1 day)
11	4	DSE	Phytotechnology	Visit to Botanical garden (1 day)
12	4	SEC	Biofertilizers and Biocontrol Agents	Field visit (1 day)
13	5	DSC	Angiosperm Systematics and Economic Botany	Field visit (3 days)
14	5	DSE	Plant Tissue Culture	Visit to Lab (1 day)
15	5	DSE	Food Science and Quality Control	Visit to food industry (1 day)
16	6	DSE	Microbial Biotechnology	Visit to Lab (1 day)
17	6	DSE	Phytogeography, Forestry and Ecotourism	Visit to ecotourism centre (1 day)
18	7	DCC	Modern Trends in Thallophytes	Field visit (1 day)
19	7	DCE	Ecology and Environmental conservation	Field visit (1 day)
20	8	DCC	Modern Trends in Archegoniatas	Field visit (1 day)
21	8	DCE	Computational Biology and Omics in Plant Sciences	Visit to lab (1 day)
22	8	DCE	Phytochemistry and Pharmacognosy	Lab/Industry visit (1 day)
23	8	DCE	Aquatic Botany	Field visit (1 day)

### Courses with Practical and Record

	<i>Semester</i>	<i>Course Type</i>	<i>Course Title</i>
1	1	DSC A	Introduction to Plant Science and Applied Botany
2	1	MDC	Ecotourism
3	2	DSC A	Plant Resources and Avenues in Botany
4	2	MDC	Gardening and landscaping
5	3	DSC A	Microbiology and Phycology
6	3	DSC A	Mycology, Lichenology and Crop Pathology
7	3	DSC B	Angiosperm Taxonomy and Economic Botany
8	4	DSC A	Archegoniates
9	4	DSC A	Plant Cell and Molecular Biology
10	4	DSC B	Plant Anatomy and Microtechnique
11	5	DSC	Angiosperm Systematics and Economic Botany
12	5	DSC	Plant Physiology and Biochemistry
13	6	DSC	Plant Anatomy and Developmental Biology
14	6	DSC	Research Methodology, Biostatistics and Computer Application
15	6	DSE	Microbial Biotechnology
16	7	DCC	Modern Trends in Thallophytes
17	8	DCC	Modern Trends in Archegoniates
18	8	DCC	Modern Trends in Plant Systematics
19	8	DCE	Computational Biology and Omics in Plant Sciences
20	8	DCE	Methods in Research, Biophysics and Biological Techniques
21	8	DCE	Phytochemistry and Pharmacognosy
22	8	DCE	Aquatic Botany



**SEMESTER I**

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Introduction to Plant Science and Applied Botany</b>				
<b>Type of Course</b>	<b>DSC A</b>				
<b>Course Code</b>	<b>M24BO1DSC100</b>				
<b>Course Level</b>	<b>100</b>				
<b>Course Summary</b>	The course "Introduction to Plant Science and Applied Botany" aims to nurture an appreciation for the importance of plants among future generations. It will introduce students to notable botanists and their contributions, the distinct characteristics of key plant groups, traditional and contemporary methodologies in plant sciences, and various branches related to the field. Through this course, students are encouraged to foster a curiosity about the plant kingdom and to actively engage in efforts to conserve plant species.				
<b>Semester</b>	1	Credits			Total Hours
<b>Course Details</b>	Learning approach	Lecture	Tutorial	Practical	Others
		3	-	1	-
<b>Prerequisite, if any</b>	Basic knowledge of biology				

#### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No.</b>
01	Comprehend the significance of plants, significant advancements in botany and human initiatives to sustain life on Earth.	U	2,5
02	Illustrate the diversity and evolutionary trends across the plant kingdom.	A	1
03	Develop skills in using instruments and techniques employed in basic studies of plant science.	A	4
04	Analyse the traditional and modern approaches in biological studies.	An	5
05	Compare the major branches of botany and evaluate	An	3,5



interdisciplinary research potential.

**\*Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course Description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Insights into the History of Botany: Contributions of eminent botanists: (a) Theophrastus, (b) Carl Linnaeus, (c) Janaki Ammal (d) M S Swaminathan.	2	1
	1.2	Plants and Human life: Medicine, food and fibre, timber (Natural and Processed), aesthetic value, maintaining ecosystem services. <b>Learning Activity 1:</b> Group Discussion on <ul style="list-style-type: none"> <li>● Uses of plants</li> <li>● Plants as Purifiers of our planet.</li> </ul>	3	1
	1.3	Morphological characters, habit and habitat of major plant groups: Algae, Bryophytes, Pteridophytes, Gymnosperms and Angiosperms. <b>Learning Activity 2:</b> An explorative nature walk to understand biodiversity of a selected locality: Paddy Field / Wetland ecosystem / Sacred Groves / Any other locality which harbors biodiversity and represents most of the major plant groups.	10	2
2	<b>Module 2 (15 hours)</b>			
	2.1	Brief overview of Botany, citing events that changed the course of world history: Quinine tree, Coconut, Rice, Wheat, Sugarcane and <i>Penicillium notatum</i> Distinct members of the plant world: Fungi, Mycorrhiza, Lichens (Salient features only) Psychoactive plants and zoopharmacognosy: Marula plant ( <i>Sclerocarya birrea</i> ); Lemurs eating tamarind and fig leaves.	4	2
	2.2	Special adaptations in plants: Insectivorous plants, Heliotropism in sunflowers, Pseudocopulation strategy in orchids. Gigantic plants: e.g. <i>Sequoiadendron giganteum</i> .	5	2

		Plants that live in extreme environments: hot water springs, deserts, marshes, Arctic regions. Biomimicry: Nature as model: Lotus effect® technology in paint industry; <i>Citrus maxima</i> fruit wall inspired design of crash absorbing structures.		
	2.3	Traditional approach and methods in plant science: (A) Exploration: Field Visit. (B) Collection of plant material: significance & tools used. (C) Preservation: Killing Agent: (Formalin), Fixing Agent: (FAA). Wet Preservation: Museum jar preservation. dry preservation: herbarium. (D) Free-hand sectioning: Transverse section (TS), Longitudinal section (LS). (E) Characters documented during field study. (F) Classification: Artificial, Natural and Phylogenetic (Definition and One Example Each). (G) Documentation: Significance of scientific diagrams and field books.	6	3,4
	<b>Module 3 (15 Hours)</b>			
	3.1	Modern Approaches in plant science (A) Sectioning: Microtomy (B) Visualization techniques: Simple, compound and electron microscope (SEM & TEM) (Brief study only). (C) Separation techniques (Principle and Application): (i) Chromatography: TLC and Paper chromatography. (ii) Centrifugation: tabletop centrifuge and ultracentrifuge. (iii) Electrophoresis: agarose gel electrophoresis (AGE).	5	3,4
3	3.2	Recent trends in Plant science: (A) Molecular techniques (General Account and Applications): PCR, DNA barcoding (B) Remote Sensing (Brief Account): Application of Remote sensing and GIS for mapping of natural resources. (C) Use of Artificial intelligence (AI) in plant science. <b>Learning Activity 3:</b> Visit to a laboratory to familiarize with instruments mentioned above.	5	3,4
	3.3	Major Branches and Scope of Plant Science: Brief account and research potential in: Plant systematics, Ecology, Phytochemistry, Plant physiology, Genetics, Ethnobotany, Crop improvement & Plant genetic engineering	5	5

	<b>Practical (30 hours)</b>			
	4.1	Field Activities (Mandatory) Conduct a field trip and survey, to appreciate the diversity of plant kingdom and to identify plants belonging to all the major groups discussed in theory. Prepare a set of 12 geo-tagged photographs containing at least one representative from each Major group.	15	2
4	4.2	Laboratory Activities (Conduct Any Three) <ul style="list-style-type: none"> <li>❖ Prepare a report and presentation on Botanists who made significant contributions to science.</li> <li>❖ Familiarize students with a compound microscope and dissecting / simple microscope. Assess the magnifications of the microscope you are examining. Compare the real image (Naked eye) with the magnified virtual image of an appropriate plant specimen and make illustrations of magnified specimens.</li> <li>❖ Prepare temporary, single stained hand sections (TS and LS; one each) of appropriate plant specimens for light microscopic studies.</li> <li>❖ Design a foldscope</li> <li>❖ Prepare an extract of leaves of appropriate plant material and perform centrifugation using a table top centrifuge.</li> <li>❖ Separation of plant pigments using paper/thin layer chromatography</li> </ul>	15	1, 3
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
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<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b></p> <p><b>A. Continuous Comprehensive Assessment (CCA)</b>  Theory Total = 25 marks  Quiz, Test Papers, seminar  Practical Total = 15 marks  Lab performance, record, field report etc.</p>
	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>

## REFERENCES

1. Ames, O., & Ames, B. (1937). *Pollination of Orchids through Pseudocopulation*. Botanical Museum Leaflets, Harvard University, 5(1), 1–xix.
2. Bajpai, P. K. (2010). *Biological instrumentation and methodology*. S Chand & co Ltd.
3. Barthlott, W., & Neinhuis, C., (1997). *Purity of the sacred lotus, or escape from contamination in biological surfaces*. *Planta* 202, 1–8.
4. Beerling D. (2007). *The Emerald Planet: How plants changed Earth's history*. Oxford University Press, New York.
5. Berg L. R. (2008). *Introductory Botany: Plants, People, and the Environment, Second Edition*. Thomson Brooks/Cole., Thomson Higher Education, 10 Davis Drive, Belmont, CA 94002-3098, USA.
6. Cotteril, R. (2002). *Biophysics- an Introduction*. John Wiley and Sons.
7. Krishnamurthy, K. V. (2004). *An Advanced Text Book on Biodiversity Principles and practice*. Oxford and IBH Publishing Co. Pvt. Ltd.
8. Kumar, N., Yamaç, S.S., & Velmurugan, A. (2015). *Applications of Remote Sensing and GIS in Natural Resource Management*. Journal of the Andaman Science Association Vol. 20(1):1-6.
9. Kochhar, S. L. (2016). *Economic botany*. Cambridge University Press.
10. Misra, A., & Agrawal, P. R., (1978). *Lichens*. Oxford and IBH, NewDelhi.
11. National Research Council (U.S.). Committee on an Examination of Plant Science Research Programs in the United States. (1992) *Plant biology research and training for the 21st century*. Washington, D.C. : National Academy Press.
12. Nita, B. (2002). *Hand book on Mushrooms*. Oxford & IBH Publishing C. Pvt.
13. Pandey, B. P. (2001). *Taxonomy of angiosperms*. S. Chand Publishing.
14. Pandey, B. P. (2022). *College Botany Volume-I, II, III*. S. Chand Publishing.
15. Rodriguez, E., & Wrangham, R.W. (1993). *Zoopharmacognosy: the use of medicinal plants by animals*. Recent Advances in Phytochemistry. Vol. 27
16. Reece, J.B., Urry, L.A. and Cain, M.L., 2017. *Campbell biology*. Pearson.


17. Sambamurthy A. V. S. S. (2006). *A Textbook of Bryophytes, Pteridophytes, Gymnosperms and Paleobotany*. I.K. International publication, New Delhi.
18. Saranya, T., Deisy, C., Sridevi, S., & Sonai M. A. (2023). *A comparative study of deep learning and Internet of Things for precision agriculture*. Engineering Applications of Artificial Intelligence. 122 (C).
19. Sharma, A., Ashutosh, S., Alexey, T., Alexander, B., Tanupriya, C., Madani, A. A., & Manuel S. (2023). *Artificial intelligence and internet of things oriented sustainable precision farming: Towards modern agriculture*. Open Life Sciences. 18 (1)
20. Sharma, V.K. (1991). *Techniques in microscopy and cell biology*. Tata McGraw-Hill, New Delhi.
21. Starr, C. (2007). *Biology: concepts and applications. VI edn*. Thomson Press.

### SUGGESTED READINGS

1. Akkem, Y., Saroj K. B., & Aruna V, (2023). *Smart farming using artificial intelligence: A review*. Engineering Applications of Artificial Intelligence. Vol. 120, 105899.
2. Arber, Agnes (1986) [1912; 2nd ed. 1938]. Stearn, William T. (ed.). *Herbals: their origin and evolution. A chapter in the history of botany, 1470-1670* (3rd ed.). Cambridge: Cambridge University Press. ISBN 9780521338790.
3. Arya Vaidya Sala Kottakkal (1994- 1997). *Indian Medicinal Plants Vol I-V*. Orient Longmann
4. Bendra, A., & Ashok, K. (1980). *Economic botany*. Rastogi publications, Meerut.
5. Chandel, N., Chakraborty, S., Rajwade, Y., Dubey, K., Tiwari, M. K., & Jat, D. (2021). *Identifying crop water stress using deep learning models*. Neural Computing and Applications. 33. 10.1007/s00521-020-05325-4.
6. Dayna, B., Benyus, J.M., Dwyer, J., Ritter, S., & Tocke, R. (2011). *Biomimicry - resource handbook*. A seed bank of best practices-CreateSpace
7. Dicks, H. (2023) - *The Biomimicry Revolution-Learning from Nature How to Inhabit the Earth*. Columbia University Press.
8. Durai, S. K. S., & Mary, D. S. (2022). *Smart farming using Machine Learning and Deep Learning techniques*. Decision Analytics Journal. Vol, 3, 100041.
9. Gifford, E. M., & Foster, A.S. (1988). *Morphology and Evolution of Vascular Plants*, W.H. Freeman & Company, New York.
10. Gifford, E.M., & Foster, A.S. (1988). *Morphology and Evolution of Vascular Plants*. W.H. Freeman & Company, New York.
11. <https://biomimicry.net/what-is-biomimicry/>
12. <https://biomimicry.org/>
13. <https://biomimicry.org/what-is-biomimicry/>
14. Janine M. B. (2009). *Biomimicry - innovation inspired by nature*. HarperCollins e-books
15. Jeffery, C. (1968). *An Introduction to Plant Taxonomy*. J and A Churchill, London.
16. Majumdar, G. P. (1982). "*Studies in History of Science in India*". In Chattopadhyaya, Debiprasad (ed.). *The history of botany and allied sciences in India* (c. 2000 B.C. to 100 A.D.). Asha Jyoti, New Delhi: Editorial Enterprise.

17. Pandey, B.P. (2005). Collage Botany : Vol I, 5th edn. S.Chand &Company LTD. New Delhi.
18. Pawlyn, M. (2019). *Biomimicry in Architecture*. RIBA Publishing.
19. Raven, P.H., Evert, R.F., & Eichhorn, S.E. (2013). *Biology of plants*. VIIIth Ed. W.H. Freeman Publishers.
20. Sambamurthy, A.V.S.S., & Subrahmanyam, N.S. (1989). *A Text Book of Economic Botany*. Wiley Eastern Ltd.
21. Sharma, V.K. (1991). *Techniques in microscopy and cell biology*. Tata McGraw-Hill, New Delhi.
22. Singh, V., Pandey, P.C., & Jain, D.K. (2017). *Anatomy of Angiosperms*. Rastogi Publication, Meerut.
23. Sivarajan, V. V. (1991). *Introduction to the Principles of Plant taxonomy*. Oxford IBH Publishing Co. Pvt. Ltd., New Delhi.
24. Varantha, P., & Gautham, N, (2005). *Biophysics*. Norosa Publishing House New Delhi.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Ecotourism</b>					
<b>Type of Course</b>	<b>MDC</b>					
<b>Course Code</b>	<b>M24BO1MDC100</b>					
<b>Course Level</b>	<b>100</b>					
<b>Course Summary</b>	The course titled “Ecotourism” provides a comprehensive exploration of sustainable tourism practices and their impact on the environment. The course describes the principle, scope, and role of ecotourism in achieving conservation goals, community engagement and benefits, ecotourism resources, planning steps of ecotourism and the role of international non-governmental organizations in ecotourism.					
<b>Semester</b>	1	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		2	-	1		-
<b>Pre-requisites, if any</b>	There are no specific prerequisites for this course.					

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Describe the fundamental principles and concepts of ecotourism	U	2
2	Summarize the components of ecotourism and the role of NGOs in ecotourism	U	2
3	Examine the characteristics and functioning of various centers of ecotourism in India	An	2
4	Explain the role of ecotourism in livelihood security	E	5

5	Design an ecotourism plan and management of ecotourism initiatives from case studies from successful ecotourism projects.	C	5
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*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Definition, concept, principles, relevance and scope, do's and don'ts of tourists in ecotourism, ecotourism impact on the environment. Eco-friendly practices, responsible tourism, sustainable tourism.	3	1
	1.2	Components of ecotourism-biodiversity conservation, education, local people, environmental awareness, cultural diversity and respect, responsible marketing, economic and social benefits.	3	1,2
	1.3	Ecotourism Resources – Natural, Geographical, cultural, festivals, events and Natural heritage sites. Terms associated with ecotourism - Adventure tourism, geotourism, wildlife tourism, canopy walkway, ecolabelling, greenwashing, hydel tourism, Eco-lodges.	3	2
	1.4	Ecotourism activities -Adventure sports, cultural activities, educational workshops, Photography, community development.	2	2
	1.5	Biodiversity and its conservation – significance of in situ conservation, Protected areas – national parks, wildlife and bird sanctuaries, forest reserves, marine national park (Gulf of Mannar). Endemism and biodiversity Hotspots - Western Ghats as a source of Ecotourism	4	2
2	<b>Module 2 (15 hours)</b>			
	2.1	Ecotourism prospects and potential of India, Ecotourism resources in India -Scope and destinations -Sundarbans, Kaziranga National Park.	3	3
	2.2	Ecotourism in Kerala, Ecotourism centres in Kerala,	3	3



		Wildlife tourism,		
	2.3	Ecotourism Planning: Steps of Ecotourism Planning- Preliminary assessment, stakeholder engagement, ecotourism Goals and Objectives, carrying capacity, Infrastructure, visitors management, conservation of ecosystem in the area, community involvement and benefits.	4	3
	2.4	Ecotourism and livelihood security- Community-based ecotourism(CBET) a tool for conservation, challenges in CBET, Joint Forest Management	2	3,4
	2.5	Role of NGOs: Role of international agencies in ecotourism – The International Ecotourism Society (TIES), World Wide Fund for Nature (WWF) and United Nations World Tourism Organization (UNWTO).	3	2,3
	<b>Practical/ Field visits (30 hours)</b>			
3	3.1	Case study on Thenmala Ecotourism and Periyar Wildlife Sanctuary, Salim Ali Bird sanctuary Thattekkadu, Kodanadu ecotourism .	6	5
	3.2	Field visit to an ecotourism site, observe and analyse the sustainable practices and submit a detailed report.	15	5
	3.3	Identify and prepare a checklist of some plant species, birds and animals having economic, ecological and cultural significance as an ecotourist attraction	4	3
	3.4	Examine the current state of natural resources and develop suitable messages and appropriate media for educating different target groups	5	5
4	<b>Teacher-specific course components</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based studies and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
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<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b></p> <p><b>A. Continuous Comprehensive Assessment (CCA)</b></p> <p>Theory Total = 15 marks  Quiz, Test Papers, seminar  Practical Total = 15 marks  Lab performance, record, field report etc.</p>
	<p><b>B. End Semester examination (ESE)</b></p> <p>Theory Total = 35 marks, Duration 1 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10 marks  Part B (Short essay) – 3 out of 6 x 5 = 15 marks  Part C (Long essay) – 1 out of 2 x 10 = 10 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>

### References

1. A K Bhattacharya, 2005. *Ecotourism and Livelihoods*. Concept Publ. company, New Delhi.
2. Kreg Lindberg, Deonal E. Hawkins, 1999. *Ecotourism: A guide for planners and managers*. Natraj Publishers, Dehradun.
3. Batta A., 2000. *Tourism and environment*. Indus Publishing Co., New Delhi.
4. Cater E, 1994. *Ecotourism in the third world: Problems and prospects for sustainability*. In: E. Cater and G. Lowman (Ed.) *Ecotourism: a sustainable option*, Wiley, Chichester.
5. Croall J, 1995. *Preserve or Destroy: Tourism and Environment*. Calouste Gulbenkian Foundation, London.
6. Lindberg, K. and D.E. Hawkins. (eds). (1993). *Ecotourism: a guide for planners and managers*. North Bennington: The Ecotourism Society.
7. Vinod Kumar, Sunil Kumar, Nitin Kamboj: *Biological diversity: Current Status and Conservation policies* (2021).
8. Stephen Wearing and John Neil (1999). *Ecotourism: Impacts, Potentials and Possibilities*, Reed Educational and Professional Publishing Limited.
9. David A Fennell and Ross K Dowling (2003). *Ecotourism Policy and Planning*. CABI Publishing, Cambridge, USA.
10. David Fennell. *Ecotourism, Third edition (2008)*. Published by Taylor and Francis e-Library.
11. India Eco-Development Project, <http://www.periyartigerreserve.org/html>
12. Community-based ecotourism, <http://www.periyartigerreserve.org/html>



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Plant Resources and Avenues in Botany</b>					
<b>Type of Course</b>	<b>DSC</b>					
<b>Course Code</b>	<b>M24BO2DSC100</b>					
<b>Course Level</b>	<b>100</b>					
<b>Course Summary</b>	The course aims to impart knowledge on the importance of plants and plant based products in everyday life. Plethora of opportunities and innovations in plant science research are also discussed. The course is designed to provide the students an idea about the career prospects in botany. The course also equips them with technical knowhow on business prospects and skills needed to successfully convert them into entrepreneurial ventures.					
<b>Semester</b>	2	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	1	-	
<b>Pre-requisite, if any</b>	Basic knowledge on plants resources and its importance in everyday life					

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
01	Identify and assess plant resources in various contexts.	U	1
02	Understand the scope and opportunities of a botany student.	U	5
03	Summarize the foundational knowledge about sustainable agriculture, horticultural activities, organic farming, nursery management and mushroom cultivation to human welfare.	An	2,4

04	Develop an understanding of entrepreneurial opportunities in plant science and fostering an entrepreneurial mindset	An	5
05	Reframe the significance of the plant world, gain insights into the potentials of personal prosperity and career opportunities in plant science.	E	5
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>Module 1 (15 Hours)</b>				
1	1.1	Plants as resource: with special significance to useful part and plant products A. Drug yielding plants: (General account with special reference to the following): Danthappala ( <i>Wrightia tinctoria</i> ), Sarpagandhi ( <i>Rauwolfia serpentina</i> ) and Periwinkle ( <i>Catharanthus roseus</i> ) Interesting plants: Rooibos, Cordyceps, Yerba mate, Mandrake. B. Plant as staple food: Special reference to Rice ( <i>Oryza sativa</i> ) and Cassava ( <i>Manihot esculenta</i> ) C. Plant as source of fiber: Cotton and Coir. D. Rubber yielding plants: Pará rubber tree. E. Plants yielding essential oils: Eucalyptus and lemongrass F. Plants in herbal and cosmetic formulations: 'Bhringaraj ( <i>Eclipta alba</i> )', <i>Hibiscus rosa-sinensis</i> , <i>Indigofera tinctoria</i> G. Vegan Cosmetics: Cleanser: Neem, Rose. H. Hair and Skin care products: Henna, Turmeric, Aloe vera. I. Plant based Milk alternatives: Green Milk	12	1
	1.2	Plant-based industries: Fruit production and processing: Dry Fruits and Canning. Fruit and Vegetable-based products: Pickle, Pulp,	3	1

		Soup, Jam and Jellies. Bamboo and Cane-based products. Plywood industry. Nutraceuticals.		
2	<b>Module 2 (15 Hours)</b>			
	2.1	Introduction to Organic Farming, gardening and landscaping, Horticulture, Mushroom cultivation	2	3
	2.2	Budding, Grafting, Layering, Floriculture and Flower arrangement, Bonsai, Terrarium.	3	3
	2.3	Brief account of Plant Research Institutes in India. Botanical Survey of India (BSI), National Bureau of Plant Genetic Resources (NBPGR), National Rice Research Institute (NRRI), Indian Cardamom Research Institute (ICRI), Jawaharlal Nehru Tropical Botanic Garden and Research Institute (JNTBGRI), Kerala Forest Research Institute (KFRI), Central Plantation Crops Research Institute (CPCRI), Rubber Research Institute of India (RRII).	3	2
	2.4	Research and Innovations in Plant Science: Research: Significance in addressing Climate change, Food Security, Biodiversity conservation Innovations in plant Science (Mention only): Crop improvement - Flood resistant rice, Green Revolution (Norman Borlaug- high Yielding Wheat), Genetic engineering - Bt. Cotton, Gene editing for disease resistance, Synthetic biology	3	2
	2.5	Hands-on Training (Any Two): <ul style="list-style-type: none"> <li>● Mushroom cultivation</li> <li>● Budding, Grafting, Layering</li> <li>● Terrarium/Bonsai</li> <li>● Development of an artificially propagated plant and submit for valuation.</li> <li>● Algal culture.</li> <li>● Tissue Culture.</li> <li>● Flower arrangement</li> </ul> <b>Activity 1 (Optional):</b> Industrial Visit / Flower Show / Agricultural Fest / Farm Visit / Food or a Center that utilizes Post Harvest Processing.	4	3
3	<b>Module 3 (15 Hours)</b>			
	3.1	Introduction to entrepreneurship: Definition and significance in the context of plant science. Basic traits and skills for entrepreneurs. Brief exploration of successful plant-based startups	3	4

		and their impact- grow the Funguy, Vgrow, Jackfruit 360, Synthite.		
	3.2	Identifying problems or opportunities within the plant science domain. Steps in the entrepreneurial journey: Overview of market assessment, enterprise selection, and resource mobilization. Schemes for Financial Assistance. Brief introduction to IPR, copyrights and GI tags.	4	4
	3.3	Opportunities in Green World: General – Scientific assistant, Plant geneticist, Computational biologist, Field botanist, Naturalist, Biotechnologist, Molecular Biologist, Nursery Manager, Plant Researcher, Teacher/Professor, Plant Pathologist, Ecologist, Plant Biochemist, Environmental Conservationist, Plant Microbiologist, Environment Consultant, Horticulturist, Plant explorer, Taxonomist, Cytologist, Biological Technician, Park Ranger, Nursery or Green House manager, Farming consultant, Paleobotanist, Plant certification, Quality assurance, Herbarium curator, Gardener, Landscape designer, Biophilic designer	4	2,5
	3.4	Career paths in Botany: Areas where a botanist can work: Research Lab/Institutions, Chemical Industry, Food Companies, Arboretum, Forest Services, Biotechnology Firms, Oil Industry, Land Management Agencies, Seed and Nursery Companies, Plant Health Inspection Services, National Parks, Biological Supply Houses, Plant Resources Laboratory and Educational Institutions <b>Activity 2:</b> Conduct a one-day workshop for students to confer awareness on academic progression, research, career and entrepreneurial prospects and opportunities in Botany.	4	2,5
4	<b>Practical (30 hours)</b>			
		Field Activities (Mandatory)		
	4.1	<ul style="list-style-type: none"> <li>Conduct one day industrial visit: To a plant-based industry in your vicinity. Prepare a report on functioning, products and marketing with Geo-tagged photographs</li> </ul>	10	2,3
	4.2	Laboratory Activities (Conduct any five)		

		<ol style="list-style-type: none"> <li>1. Make collections of plant products specified in the syllabus and submit</li> <li>2. Polybag cultivation of mushroom</li> <li>3. Demonstrate Air layering, T-budding and patch budding</li> <li>4. Select any plant based start-up initiative and prepare a report</li> <li>5. Present a mock up idea for a plant based entrepreneurship</li> <li>6. Flower arrangement – fresh and dry</li> <li>7. Jam, Jelly preparation</li> </ol>	20	1, 3, 4, 5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b>  Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b>  <b>A. Continuous Comprehensive Assessment (CCA)</b>  Theory Total = 25 marks  Quiz, Test Papers, seminar  Practical Total = 15 marks  Lab performance, record, field report etc</p>
	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x1 =10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>

## REFERENCES

1. Acquaah, G., (2019). *Horticulture: Principles and Practices (4th edition)*. India: Pearson India Education Services Pvt. Ltd.
2. Arya Vaidya Sala Kottakkal (1994- 1997). *Indian Medicinal Plants., Vol I-V*. Orient Longmann



3. Arya, H., & Bhatt, T. K., (2021). *Introduction of intellectual property rights. In The Design & Development of Novel Drugs and Vaccines*. Academic Press. (pp. 275-281).
4. Aydara, E. F., Sena, T., & Beraat, O. (2020). *Plant-based milk substitutes: Bioactive compounds, conventional and novel processes, bioavailability studies, and health effects*. *Journal of Functional Foods*. 103975. 1-15.
5. Chrispeels, M. J., & Sadava, D. E., (1994). *Plants, Genes and Agriculture*. Jones & Bartlett Publishers.
6. Cruses, W.V., & Fellows, P. J., (2000). *Commercial fruits and vegetable processing*. CRC press, United
7. Kalian Kumar De, 1996. *Plant Tissue Culture*. New Central Book Agency (P) Ltd.
8. Mohanty, S. K. (2005). *Fundamentals of entrepreneurship*. PHI Learning Private Limited, Rimjhim House, 111, Patparganj Industrial Estate, Delhi-110092. ISBN-978-81-203-2867-9
9. Narayana, P. S., Varalakshmi, D., & Pullaiah, T., (2021). *Research Methodology in Plant Science 2nd Edition*. Scientific(R) Publishers. Jodhpur , Rajasthan, India.
10. Sambamurthy, A. V. S. S., & Subrahmanyam, N. S., (1989). *A Text Book of Economic Botany*. Wiley Eastern, New Delhi. ISBN: 9780852268803, 0852268807
11. Sandhu, M. K., (1989). *Plant Propagation*. Wiley Eastern Ltd., Bangalore.
12. Simpson, B. B., & Conner-Ogorzaly, M. (1986). *Economic Botany - Plants in Our World*. McGraw Hill, New York.
13. Smith, V. A., (1903). *The Indian Civil Service as a profession (1 ed.)*. Dublin: Hodges, Figgis, & Co., Ltd.
14. Stagg B. C., & Justin, D., (2023). *Plants, education and sustainability: rethinking the teaching of botany in school science*. *Journal of Biological Education*. 57 (5), 941-943
15. Zheng, B., Hualu, Z., & David, J. M., (2021). *Nutraceutical-fortified plant-based milk analogs: Bioaccessibility of curcumin-loaded almond, cashew, coconut, and oat milks*. *LWT - Food Science and Technology*. Vol, 147, 111517

## SUGGESTED READINGS

1. Amprazis, A., & Papadopoulou, P., (2018). “*Primary School Curriculum Contributing to Plant Blindness: Assessment Through the Biodiversity Perspective*”. *Advances in Ecological and Environmental Research* 3 (11): 238–256.
2. Armstrong, E. M., Larson, E. R., Harper, H. C., Webb, R., Dohleman, F., Araya, Y., & Grierson, C. S., (2023). *One Hundred Important Questions Facing Plant Science: An International Perspective*. *New Phytologist* 238 (2): 470–481.
3. Bailey, L. H., (2009). *Manual of Gardening*. Srishti Book Distributors, New Delhi.
4. Bassett, M. J., (1986). *Breeding Vegetable Crops*. Westport, Conn.: AVI Publishing.
5. Beasley, K., Lee-Hammond, L., & Hesterman, S., (2021). “*A Framework for Supporting the Development of Botanical Literacies in Early Childhood Education*”. *International Journal of Early Childhood* 53 (2): 119–137.


6. Biles, R. E., (2003). *The Complete Book of Gardening*. Biotech Books, Delhi.
7. Bonney, R., Cooper, C.B., Dickinson, J., & Steve, K., (2009). *Citizen science: a developing tool for expanding science knowledge and scientific literacy*. *Bioscience* 59, 977e984.
8. Borlaug, N., (1970). *The Green Revolution, Peace, and Humanity*. Nobel Lecture. Available at <http://www.nobel.se>.
9. Bose, T. K., & Mukherjee, D., (1972). *Gardening in India*. Oxford & IBH Publishing Co., New Delhi.
10. Buckley, C., (2020). *Plant Magic: Herbalism in Real Life*. Roost Books Publishers, New York.
11. Chen, G., & Weibang, S., (2018). *The role of botanical gardens in scientific research, conservation, and citizen science*. *Plant Diversity*. 40 (4) P. 181-188
12. Chen, X., Lu, X., Shu, N., Wang, S., Wang, J., Wang, D., Guo, L., & Ye, W., (2017). *Targeted mutagenesis in cotton (*Gossypium hirsutum* L.) using the CRISPR/Cas9 system*. *Scientific Reports*. 7, 44304.
13. Clough, D. R., Fang, T. P., Vissa, B., & Wu, A., (2019). *Turning lead into gold: How do entrepreneurs mobilize resources to exploit opportunities?* *Academy of Management Annals*, 13(1), 240-271.
14. Cohn, J.P., 2008. *Citizen science: can volunteers do real research?* *Bioscience* 58, 192e197.
15. Conrad, C.C., & Hilchey, K.G., (2011). *A review of citizen science and community-based environmental monitoring: issues and opportunities*. *Environ. Monit. Assess.* 176, 273e291.
16. Courtier, J., & Clarke, G., (1997). *Indoor plants: The Essential Guide to Choosing and Caring for Houseplants*. Reader's Digest, New York.
17. Fuller, K. W., & Gallon, J. A., (1985). *Plant Products and New Technology*. Clarendon Press, Oxford, New York.
18. Gopichandran, V., & Satish, K., Ch., (2012). *Mainstreaming AYUSH: an ethical analysis*. *Indian Journal of Medical Ethics*. 9 (4): 272–277.
19. Hershey, D. R., (1996). *"A Historical Perspective on Problems in Botany Teaching."* *The American Biology Teacher* 58 (6): 340–347.
20. Kletečki, N., Hruševar, D., Mitić, B., & Šorgo, A., (2023). *"Plants are Not Boring, School Botany is."* *Education Sciences* 13 (5): 489.

### Websites

1. <http://www.rubberboard.org.in>
2. <https://work.chron.com/normal-job-duties-botanist-career-25069.html>
3. <https://www.indeed.com/career-advice/finding-a-job/jobs-in-plant-science>
4. <https://cpcri.icar.gov.in/>
5. <https://csirnet.nta.ac.in/>
6. <https://fgtb.icfre.gov.in>
7. <https://icar-nrri.in>
8. <https://iisr.icar.gov.in>
9. <https://jntbgri.res.in/>
10. <https://leverageedu.com/blog/career-in-botany/>

11. <https://nbpgr.ernet.in>
12. <https://ssc.nic.in/>
13. <https://upsc.gov.in/>
14. <https://www.ctcri.org/>
15. <https://www.kfri.res.in/>
16. <https://botany.org/home/careers-jobs/careers-in-botany/requirements-for-a-career-in-botany.html>
17. [www.bsi.gov.in](http://www.bsi.gov.in)



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Gardening and landscaping</b>					
<b>Type of Course</b>	<b>MDC</b>					
<b>Course Code</b>	<b>M24BO2MDC100</b>					
<b>Course Level</b>	<b>100</b>					
<b>Course Summary</b>	This course provides a comprehensive exploration of gardening and landscaping principles, equipping students with the knowledge and skill to create and maintain beautiful sustainable outdoor spaces. Students will earn foundational knowledge in nursery management techniques, including propagation and soil preparation. The course will familiarise students with essential tools, components and structures used in garden designing. Exploring eco-friendly practices in garden design can contribute to environmental conservation.					
<b>Semester</b>	2	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		2	-	1		-
<b>Pre-requisites, if any</b>	Basic understanding of Biology					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Understand the basics of ornamental and landscape gardening	U	1,4
2	Recollect the basic knowledge of plant growth structures used in gardening	K	4
3	Review the principles of gardening and nursery management	U	4
4	Explain various propagation techniques used in a nursery	U	4

5	Apply the knowledge of gardening and landscaping to design a garden	C	5
<i>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</i>			

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Introduction to landscaping, gardening and commercial floriculture – importance and prospects	2	1
	1.2	Types of plants in landscaping– Trees, shrubs, climbers, annuals, herbaceous perennials, bulbous crops, palms, ferns, cacti & succulents, aquatic ornamentals.	2	1
	1.3	Types of gardens- fruit garden, ornamental garden, herbal garden, kitchen garden, Kids Garden Indoor plants (Money plant, Snake plant, Monstera, ZZ plant, Aglaonema)	4	2
	1.4	Horticultural practices related to gardening – training, pruning and mulching, its benefits. Nursery bed preparation	2	2
	1.5	Vegetative propagation methods – natural and artificial cuttings – leaf, stem and root, layering–air layering, simple layering, grafting- approach grafting, Tongue grafting, budding- T budding, patch budding	5	4
2	<b>Module 2 (15 hours)</b>			
	2.1	Nursery layout & structures: Polyhouse, mist chamber, rain shelter, potting shed, composting shed. Sprinkler irrigation.	3	3
	2.2	Gardening tools & implements Garden spade, rake, fork, garden shears, secateurs, grafting & budding knife, pruning saw, mowers, brush cutter, garden tillers	2	2

	<b>2.3</b>	Garden components and adornments (brief account only)	2	2
	<b>2.4</b>	Rockery, Terrarium, Kokedema, Bonsai (brief account only)	2	2, 3
	<b>2.5</b>	Elements of art-colour, line, form, scale. Principles of Landscape design- Unity, Balance, transition, proportion, rhythm, focalisation, repetition, simplicity.	3	2, 5
	<b>2.6</b>	Steps in developing a Landscape Design Brief Account Only a) Site analysis- b) Identification of functional requirements; c) site development by exploiting natural forms; d) Elements in landscape design- form, water, garden furniture, lights, paving etc. e) study of plant trees, shrubs and ground cover, indoor plants etc.	3	2, 5
	<b>Practicals (30 hours)</b>			
<b>3</b>	<b>3.1</b>	Visit to a well-established nursery/ Garden and submit a detailed report	8	6
	<b>3.2</b>	TTC test for assessing seed viability	2	3
	<b>3.3</b>	Preparation of potting mixture	2	4
	<b>3.4</b>	On-hand training for air-layering, approach grafting and T-budding techniques	6	4
	<b>3.5</b>	Identification of Garden tools and implements.	4	3
	<b>3.6</b>	Designing of Terrarium	4	5
	<b>3.7</b>	Designing of Kokedama balls/ bottle gardens	4	5
<b>4</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based studies and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 15 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. End Semester examination (ESE)</b> Theory Total = 35 marks, Duration 1 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 3 out of 6 x 5 = 15 marks Part C (Long essay) – 1 out of 2 x 10 = 10 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks


### References

1. Laurie, A. & Ries, V.H. 2012. *Floriculture- Fundamentals and Practices*, Agrobios
2. Hartmann, HT. and Kester, D.E.1986. *Plant Propagation - Principles and practices*. Prentice Hall, New Delhi.
3. Peter, K. V. *Basics of Horticulture*. New India Publishing Agency, New Delhi.
4. Randhawa GS & Mukhopadhyay A. 1986. *Floriculture in India*. Allied Publishers.
5. Rajmohan,K., Soni,K.B., Gomathi, KS & Prakah,R. 2004. *Essentials of Plant Tissue Culture*. Kerala Agricultural University
6. Larson, R.A., 1980. *Introduction to Floriculture*. Academic Press, London
7. Sheela, V.L. 2011. *Horticulture*. MJP Publishers, Chennai.
8. Singh, J. 2002. *Basic Horticulture*, Kalyani Publishers.
9. Bose TK, Maiti RG, Dhua RS & Das P. 1999. *Floriculture and Landscaping*. Naya Prokash.
10. De, L.C. 2012. *Handbook of Gardening*, Aavishkar Publishers, Jaipur
11. Randhawa GS & Mukhopadhyay A. 1986. *Floriculture in India*. Allied Publishers.
12. Sabina GT & Peter KV. 2008. *Ornamental Plants*. New India Publishing Agency, New Delhi.
13. Sundaram,V. 2016. *Textbook on Commercial flowers and Ornamental Gardening*. Kalyani Publishers.
14. Syamal, M.M. 2014. *Commercial Floriculture*. Jaya Publishing House, New Delhi.
15. Nambisan, K.M.P.1992. *Design element of landscape gardening*, company, New Delhi



**SEMESTER III**



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Microbiology and Phycology</b>				
<b>Type of Course</b>	<b>DSC A</b>				
<b>Course Code</b>	<b>M24BO3DSC200</b>				
<b>Course Level</b>	<b>200</b>				
<b>Course Summary</b>	The course will give an insight towards the diversity of microbes and algal flora. The study of microbiology provides a comprehensive understanding of microbes, its principles, and its applications in various fields, whereas phycology deals with the study of algae. Being the primary producers, both micro and macroalgae play a significant role in aquatic ecosystems. Students learn its salient/ diagnostic features and its importance to ecosystems. It also focuses on the economic and ecological significance and its applications.				
<b>Semester</b>	3	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		3	-	1	-
<b>Pre-requisites, if any</b>	Basic botanical knowledge and laboratory skills				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PO No
1	Understand the world of microbes and its significance	U	1
2	Examine the range of thallus structure, pigment composition, photosynthetic end products and reproduction in various algal groups.	An	1,3
3	Analyse the identifying features of microbes and algae	An	1,3

4	Demonstrate a comprehensive understanding of the economic importance of microbes and algae.	U	4,5
5	Examine the ecological significance and research potential of algae	An	5
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>Module 1 (15 hours)</b>				
1	1.1	Bacteria: General characters and classification based on staining, morphology and flagellation. Ultra structure of bacteria. Reproduction - binary fission. Genetic recombination in bacteria - conjugation, transformation and transduction.	6	1,3
	1.2	Viruses: General characters of viruses, viroids and prions. Structure of TMV and Bacteriophage ( $\lambda$ ). Multiplication of $\lambda$ phage – lytic and lysogenic cycle.	4	1,3
	1.3	Microbial interactions in ecosystems, Applications of microbes in industry, agriculture, food and medicine. Microbes in environmental conservation, waste management and as biocontrol agents. Harmful aspects of microorganisms (Brief account)	5	1,4
<b>Module 2 (15 hours)</b>				
2	2.1	History of algal classification, study of classification by Fritsch (1945); Brief introduction to the modern classification by Lee (1989) [up to class]. Algae Base (Brief study)	2	2
	2.2	Distribution, habitat diversity, range of thallus structure, pigment composition and photosynthetic end products in various groups of algae. Reproduction - vegetative, asexual and sexual reproduction. Outline of major life cycle patterns found in algae.	2	2
	2.3	Salient features and thallus structure of algae in the following groups with special reference to the type(s) mentioned: Cyanophyceae - <i>Nostoc</i> ; Chlorophyceae - <i>Volvox</i> , <i>Spirogyra</i> , <i>Cladophora</i> ,	11	2,3

		<i>Chara</i> Bacillariophyceae - <i>Pinnularia</i> ; Phaeophyceae– Sargassum; Rhodophyceae – <i>Polysiphonia</i> Reproductive structures of <i>Chara</i> , <i>Sargassum</i> and <i>Polysiphonia</i> (Brief account)		
	<b>Module 3 (15 hours)</b>			
<b>3</b>	3.1	Useful aspects of algae: Food, SCP, Biofertilizers, Medicine Exploration of algae as source of valuable commercially important products-carrageenan, agar-agar, alginate, diatomite Harmful effects of algae: Algal blooms, eutrophication, neurotoxins.	6	4
	3.2	Algae as primary producers and ecosystem engineers Algal associations and its significance (Parasitic algae, Symbiotic algae-association of algae with fungi, bryophytes, pteridophytes, gymnosperms, angiosperms, invertebrates) Algae based wastewater treatment for biodiesel production Role of algae as bioremediation agents. Role of algae in N <sub>2</sub> fixation	6	4,5
	3.2	Role of algae in scientific research - <i>Chlorella</i> Brief overview on cultivation of macroalgae and microalgae.	3	5
<b>4</b>	<b>Practical (30 hours)</b>			
	<b>Microbiology (10 hours)</b>			
	4.1	Gram staining – curd/ root nodules. Isolation of microbes from soil through serial dilution	8	1,4
	4.2	Demonstrate the culture of bacteria.	1	1,4
	4.3	Type of fermentation and respective microbes - wine, vinegar, curd	1	1,4
	<b>Phycology (20 hours)</b>			
4.4	Conduct a field visit to any one of the ecosystems rich in algae to experience algal diversity. Submit a report with photographs or	3	2,3,4	

		Collect algae from diverse habitats, observe through microscope and prepare microphotographs and submit a report.		
	4.5	Make micro preparations of thallus structures of the types mentioned in the syllabus.	16	2,3,4
	4.6	Familiarizing the technique of algal collection and preservation.	1	2,3,4
5	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

## References

1. Adetunji, C. O., Oloke, J. K., Dwivedi, N., Ummalyma, S. B., Dwivedi, S., Hefft, D. I., & Adetunji, J. B. (Eds.). (2023). *Next-Generation Algae, Volume 1: Applications in Agriculture, Food and Environment*. John Wiley & Sons.
2. Bold, H. C., & Wynne, M. J. (1985). *Introduction to the algae: structure and reproduction*. Prentice Hall.
3. Campbell, R. (1987). *Plant Microbiology*. ELBS Edward Arnold, London.
4. Dube, H.C. (2008). *Fungi, Bacteria and Viruses*. Agrobios.
5. Fritsch, F.E. (1945). *The structure and reproduction of Algae, Volume I*. Cambridge University Press.
6. Guiry M.D., & Guiry, G.M. (2024). AlgaeBase. World-wide electronic publication. National University of Ireland, Galway. <http://www.algaebase.org>

7. Kim, S.K. (2011). *Marine medicinal foods: Implications and Applications of micro and macroalgae*. Academic Press, New York.
8. Lee, R. E. (1989). *Phycology, 2nd Edition*, Cambridge University Press, New York.
9. Linda, E.G., & Lee, W.W. (2000). *Algae*, Prentice Hall, New Jersey.
10. Moheimani, N. R., McHenry, M. P., De Boer, K., & Bahri, P. A. (2015). Biomass and biofuels from microalgae. *Biofuel and biorefinery technologies*. Springer International Publishing Switzerland.
11. Pelczar, M.J., Reid, R.D., & Chan, E.C.S. (1983). *Microbiology*. Fourth Edition. Tata McGraw-Hill, New York..
12. Sahho, D., & Seckback, J. (2015). *The algae world*. Springer Dordrecht Heidelberg New York.
13. Sharma, O.P. (2011). *Algae*. Tata McGraw- Hill, New Delhi.
14. Smith, G.M. (1955). *Cryptogamic Botany (Vol. 1): Algae and Fungi*. Tata McGraw Hill, New York.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Mycology, Lichenology and Crop Pathology</b>					
<b>Type of Course</b>	<b>DSC A</b>					
<b>Course Code</b>	<b>M24BO3DSC201</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	The course provides a comprehensive exploration of the intricate worlds of fungi and plant diseases. Students delve into the morphology and ecological roles of fungi, gaining insights into their diverse functions as decomposers, symbionts, and pathogens. The curriculum also encompasses the study of plant diseases, investigating the interactions between plants and various pathogenic organisms, including fungi, bacteria, viruses, and nematodes. Through this course, students acquire the skills and knowledge necessary for disease diagnosis, prevention, and control, contributing to the sustainable management of plant populations in diverse settings.					
<b>Semester</b>	3	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Basic knowledge in botany					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Determine the diversity, reproductive behaviour and applications of fungi	A	1
2	Determine the diversity, reproductive behaviour and applications of lichens	A	1
3	Identify ecological and economical significance of fungi	U	1,2

	and lichens		
4	Describe the basic aspects of plant pathogen interaction	U	2
5	Recognize the plant diseases and suggest control measures	K	2,4
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>1</b>	<b>Module 1 (15 hours)</b>			
	1.1	Introduction and general characters of fungi. Classification based on Ainsworth (1973); Assembling the Fungal Tree of Life (AFTOL) - a brief account. Mycobank (Brief account)	3	1
	1.2	Salient features of each group, thallus and reproductive structures of the genera mentioned in each group below; Myxomycotina - General Characters	1	1
	1.3	Mastigomycotina – <i>Albugo</i> (Difference between Oomycete and true fungi)	2	1,3
	1.4	Zygomycotina – <i>Rhizopus</i>	2	1,3
	1.5	Ascomycotina: - <i>Xylaria</i>	2	1,3
	1.6	Basidiomycotina – <i>Agaricus, Puccinia</i>	4	1,3
	1.7	Deuteromycotina - <i>Fusarium</i>	1	1,3
<b>2</b>	<b>Module 2 (15 hours)</b>			
	2.1	Economic importance of Fungi – Beneficial and detrimental aspects.	3	1,3
	2.2	Fungi of Agricultural importance – mycoherbicides, myconematicides, mycoparasites, Mycorrhiza – types, function, and significance.	3	1,3

	2.3	Mushrooms- edible and poisonous types. Cultivation technique-Spawn production of Oyster mushroom, cultivation of Oyster mushroom (General Outline)	4	1,3
	2.4	General account, economic and ecological importance of lichen	1	2,3
	2.5	Classification of lichens based on thallus and its significance	2	2,3
	2.6	Structure and life cycle of <i>Parmelia</i> .	2	2,3
	<b>Module 3 (15 hours)</b>			
3	3.1	History of crop pathology (Brief study)	1	4
	3.2	Classification of plant diseases based on causative organisms and symptoms	2	4
	3.3	Plant-Pathogen Interaction (general outline)	1	4,5
	3.4	Defense mechanisms in Plants	2	4,5
	3.5	Mechanism of infection, transmission, and dissemination of plant diseases.	2	4,5
	3.6	Prophylaxis - quarantine measures, seed certification; Therapeutic – physical therapy, chemotherapy.	1	4,5
	3.7	Biological control of plant diseases	1	4,5
	3.8	Study of following diseases with emphasis on symptoms, cause, and control: <ul style="list-style-type: none"> <li>● Bunchy top of Banana</li> <li>● Bacterial blight of Paddy</li> <li>● Root wilt of Coconut</li> <li>● Abnormal leaf fall of Rubber</li> <li>● Leaf mosaic disease of Tapioca</li> <li>● Quick-wilt of pepper.</li> <li>● Nut fall of Arecanut.</li> </ul>	5	5
4	<b>Practical (30 hours)</b>			
	<b>Mycology (20 hours)</b>			




	4.1	Identification of the following types by making suitable micro preparations: <i>Albugo</i> , <i>Rhizopus</i> , <i>Xylaria</i> , <i>Puccinia</i> , <i>Agaricus</i> , and <i>Fusarium</i> .	8	1
	4.2	Staining of endomycorrhiza or fungus using Trypan Blue.	2	1, 3
	4.3	Collection/identification of common macrofungi (5 types).	10	1
<b>Plant Pathology (10 hours)</b>				
	4.4	Identify the diseases mentioned in the syllabus with respect to causal organisms and symptoms	3	4,5
	4.5	Submit specimens/ herbarium preparations of any three diseases/Geo tagged photos.	4	5
	4.6	Preparation of fungicides – Bordeaux mixture/ Tobacco decoction.	3	5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.</p> <p><b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks</p>

## References

1. Agrios, G. N. (2005). *Plant pathology*. Elsevier.
2. Ainsworth, G. C., & Sussman, A. S. (Eds.). (2013). *The fungal population: an advanced treatise*. Elsevier.
3. Alexopoulos, C. J., Mims, C. W., & Blackwell, M. (1996). *Introductory Mycology*. John Willey and Sons. Inc., New York, 868.
4. Borkar, S. G. (2017). *History of plant pathology*. CRC Press.
5. Bush, J. (2019). *Genetics of Plant Diseases*. Scientific e-Resources.
6. Campbell, R. (1987). *Plant Microbiology*. ELBS Edward Arnold, London
7. Deacon, J.W. (2013). *Fungal biology*. John Wiley & Sons.
8. Gogoi, R., Rathaiah, Y., Borah T. R. (1990). *Mushroom Cultivation Technology*. Scientific Publishers (India).
9. Gupta, V. K., Paul, T. S. (2004). *Fungi & Plant diseases*. Kalyani publishers, New Delhi
10. Hale M E, 1983. *The Biology of Lichen* (III Edn). Edward Arnold, London.
11. Jim Deacon (2006). *Fungal Biology* (IV Edn). Blackwell Publishing.
12. Mehan, V. K., & Sharma, V. K. (2010). Plant Pathology in India–Historical Developments. *Plant Pathology in India*, 19.
13. Mehrotra, R.S. and Aneja, K.R. (1990). An introduction to mycology. New Age International.
12. Varma, A., Abbott, L., Werner, D., & Hampp, R. (Eds.). (2007). *Plant surface microbiology*. Springer Science & Business Media.
14. Vasishta, P. C. (1995). *Botany (for Degree Students)*. S. Chand Limited.
15. Nita, B (2002). *Hand book on Mushrooms*, Oxford & IBH Publishing C. Pvt. Ltd. New Delhi.
16. Sharma, P. D. (2004). *The Fungi*, 2<sup>nd</sup> Edition, Rasthogi publication

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>						
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>						
<b>Course Name</b>	<b>Analytical Techniques in Plant Sciences</b>						
<b>Type of Course</b>	<b>DSE</b>						
<b>Course Code</b>	<b>M24BO3DSE200</b>						
<b>Course Level</b>	<b>200</b>						
<b>Course Summary</b>	This course will provide an overview of the various preparative methods and analytical techniques used in plant science. Students will learn the principles of different analytical techniques and its practical applications in plant research.						
<b>Semester</b>	3		Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	60	
		4	-	-	-		
<b>Pre-requisites, if any</b>	Basic knowledge in science						

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Outline the methods and procedures in microscopy	U	3,4
2	Explain working and application of various separation and analytical techniques	U	3,4
3	Articulate the principles underlying different instruments employed in plant science research	A	3,4
4	Apply the techniques in enumeration, analysis and purification of plant samples	A	3,4
5.	Acquire expertise in various preparative methods and analytical techniques used in plant science	A, S	3,4

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			

	1.1	Introduction to analytical techniques in plant science, Collection, preservation (dry & Wet) and preparation of plant materials: squash, smear, whole mount, maceration, and Sectioning. Retaining the natural colour of the plant samples (brief study).	4	1, 5
	1.2	Killing and fixing: properties of good fixative: types of fixative and fixation; killing and fixing agents and their composition (Carnoy's fluid and FAA)	2	1
	1.3	Sectioning- free hand and microtomy, applications of microtome - rotary microtome, sledge microtome, and cryostat	3	1
	1.4	Stains and staining techniques – different types of stains and their composition- safranin, acetocarmine; vital stains - neutral red, Evans blue, types of staining - Single staining and Double staining.	4	1,5
	1.5	Mounting and preparation of slides - mounting media: glycerine, DPX, and Canada balsam; preparation of slides: temporary and permanent	2	1,5
	<b>Module 2 (15 Hours)</b>			
2	2.1	Principle, working, and application: Light microscopy – Compound and dissection microscopes, parts and uses. Phase contrast microscopy, Electron microscopy (Brief account).	6	1,3
	2.2	Enumeration Techniques: Haemocytometer, Sedgewick-rafter.	2	3,4
	2.3	Micrometry	2	3,4
	2.4	<b>Activity:</b> 1. Temporary mounting of a hand-sectioned single-stained specimen 2. Maceration of a given specimen (Cucurbita stem)	5	1,5
	<b>Module 3 (15 Hours)</b>			
3	3.1	Centrifugation - Principle, working, and application of high-speed centrifuge and ultracentrifuge (preparative and analytical model) Different types of centrifuges: Fixed angle, swinging bucket	4	3,4
	3.2	Photometric Analysis – principle, working, and application of colorimeter and spectrophotometer. Application of UV-visible	6	3,4


		spectroscopy in plant science and related fields.		
	3.3	Principle, working, and application of pH meter	2	3,4
	3.4	<b>Activity</b> 1. Prepare a standard graph and estimate the concentration of a solution using a colorimeter 2. Adjust the pH of a solution using pH meter/ pH pen	3	5
	<b>Module 4 (15 Hours)</b>			
	4.1	Chromatography Techniques: - principle, working, and application of paper chromatography, TLC, column chromatography.	5	2,3
4	4.2	Electrophoresis: Electrophoretic mobility, factors affecting electrophoretic mobility. working and application of SDS-PAGE and agarose gel electrophoresis	4	2,3
	4.3	<b>Activity:</b> 1. Visit a recognized instrumentation lab or research lab and submit a report. 2. Separate plant pigments using paper/thin layer chromatography	6	5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester Examination (ESE)</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## REFERENCES

- Berlyn, G.P. & Miksche, J.P. (1976). *Botanical Microtechnique and Cytochemistry*. Wiley-Blackwell.

2. Campbell, I.D. & Dwek, R. A. (1984). *Biological Spectroscopy*. Benjamin Curmmings Publication Co. Inc.
3. Chang R (1971). *Basic principles of spectroscopy*. McGraw Hill.
4. Gray (1964). *Handbook of Basic Microtechnique*. McGraw Hill co.
5. Glasel, J. & Deutscher, M. B. (1995). *Introduction to Biophysical Methods for Protein and Nucleic acid Research*. Academic Press.
6. Huang, B.Q. & Yeung, E.C. (2015). *Chemical and Physical Fixation of Cells and Tissues: An Overview*. In E.C.T. Yeung, C. Stasolla, M.J. Sumner & B.Q. Huang (Eds.) *Plant Microtechniques and Protocols* (pp. 23-44), Springer.
7. Johanson D A (1940). *Plant microtechnique*. McGraw Hill co.
8. John E Sass (1967). *Botanical Microtechnique*. Oxford IBH Publ. Company.
9. Khandpur, R.S. (2006). *Handbook of analytical instruments*. Tata Mc Graw Hill.
10. Khasim, S.M. (2002). *Botanical Microtechnique: Principles and Practice*. Capital Publishing Company.
11. Krishnamurthy K V (1987). *Methods in Plant Histochemistry*. S Viswanathan printers, Anand book depot, Madras.
12. Nakara, B.C. & Choudhari, K.K. (2003). *Instrumentation measurements and analysis*. Tata Mc Graw Hill.
13. Pattabhi, N.V. & Gautham, N. (2002). *Biophysics*. Narosa Publishing House.
14. Pavia, D. L., Lampman, G. M., Kriz, G. S., & Vyvyan, J. R. (2015). *Introduction to spectroscopy*.
15. Perkampus H (1992). *UV-VIS Spectroscopy and its applications*. Springer-Verlag.
16. Prasad, M.K. and Prasad, M.K. (1972). *Outlines of Botanical Microtechnique*. Emkay Publishers.
17. Ruzin, S.E. (1999). *Plant Microtechnique and Microscopy*. Oxford University Press, New York, U.S.A.
18. Toji Thomas (2005). *Essentials of botanical microtechnique* (II Edn). Apex infotech publishing company.
19. Willard, H.H., Merritt L.L. Dean J.A. & Settle F.A. (1986). *Instrumental Methods of Analysis*. 7<sup>th</sup> Ed., Wadsworth Publishing Co.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Horticulture and post-harvest technology</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO3DSE201</b>				
<b>Course Level</b>	<b>200</b>				
<b>Course Summary</b>	Students are expected to gain knowledge on various Horticultural disciplines including gardening, field management and postharvest technologies. They will also develop an understanding of Regulatory Laws related to food safety and quality control along with exploring the entrepreneurial aspects within the field of Horticulture.				
<b>Semester</b>	3	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		4	-	-	-
<b>Pre-requisites, if any</b>	Familiarity with basic plant science, soil science and environmental science				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Develop a comprehensive understanding of horticulture, importance and its branches	U	1,5
2	Apply crop management techniques in horticulture including soil preparation, irrigation and pest control	A	4
3	Develop expertise in postharvest handling techniques to minimize losses and enhance the shelf life	A	4
4	Administer storage and transportation practices to maintain freshness and nutritional quality	A	4

5	Develop new value addition strategies and entrepreneurial skills required for the horticultural industry	C	4,5
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT


Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Introduction, Scope and Importance, Branches of horticulture.	3	1
	1.1	Components of soil: Organic, Inorganic & physiological-types and its importance.	2	2
	1.2	Classification of soil: Criteria for classification - soil profile- soil types - red soil, black soil, alluvial soil, laterite soil, coastal soil, sandy soil, serpentine soil, sodic soil, problematic soil, acidic and alkaline.	4	2
	1.3	Irrigation: Principles. Methods of irrigation - surface, subsoil and overhead irrigation system – types.	2	2
	1.4	Manuring: organic and Synthetic manures - Classification. Methods of manuring- broadcast, seed treatment, foliar application	3	2
	1.5	Estimation of soil pH using pH meter.	1	2
2	<b>Module 2 (15 Hours)</b>			
	2.1	Gardening: styles of gardens - English, Mughal, Japanese, Persian, French and Italian gardens - characteristics and components (Brief account Only). Garden tools and Implements – Types. Garden designing and layout. Different types of gardens: Outdoor, indoor garden, water garden, rockery.	6	1
	2.2	Landscape architecture: types - Contemporary, Environmental, Industrial, institutional and	5	1,5



		playground landscaping.		
	2.3	Plant propagation methods: Budding, Grafting, Layering and Tissue culture.	4	2
2	<b>Module 3 (15 Hours)</b>			
	3.1	Major branches of horticulture: Floriculture: definition and significance, Components – Cut flower, loose flower, dry flower, Floral oil. Olericulture: definition and significance; Types of vegetables: Warm season and cool season vegetables, types of vegetable farming - kitchen, garden, terrace garden, market garden, truck garden. Pomology: Types of fruits – Tropical, Subtropical and Temperate. General care of fruit crops - techniques for planting, pruning and training, pest management.	5	1
	3.2	Practice different types of grafting (approach, whip and tongue, cleft), T budding/ Patch Budding.	5	1
	3.3	Visit a garden and identify the components, plants, and prepare a report. Collect, familiarize and identify ornamental plant groups.	5	2
4	<b>Module 4 (15 Hours)</b>			
	4.1	Importance of post-harvest management. Postharvest handling methods: Washing, Grading, Waxing. Storage methods: Pre-cooling. Controlled atmospheric storage, Modified atmospheric storage – Low pressure storage and cold chain concept	3	3,4
	4.2	Packaging of fresh and processed products: general principles and methods of preservation - dehydration, thermal processing, chemical preservatives, fermentation, ionizing, radiation, Preparation of jams, jellies, squashes, pickles, salads, syrups and beverages.	4	4,5
	4.3	Government policies, regulations and specifications for fresh and processed products, Food safety and quality control-FSSAI. Export promotion agencies and their role on export of	3	5

		fresh and processed products.		
	4.4	Importance and scope of processing industry in India. General guidelines for the establishment of small and large scale processing units. Business opportunities, Role of HortiCorp and VFPCCK.	2	5
	4.5	Training on making jams, jellies, squashes, pickles, salads, syrups and beverages	3	5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b></p> <p>Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b></p> <p><b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar</p> <p><b>B. End Semester Examination (ESE)</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Angiosperm Taxonomy and Economic Botany</b>				
<b>Type of Course</b>	<b>DSC B</b>				
<b>Course Code</b>	<b>M24BO3DSC202</b>				
<b>Course Level</b>	<b>200</b>				
<b>Course Summary</b>	The course provides a thorough understanding about the morphological and reproductive characters of angiosperm plants utilized for identification and classification. The course elaborates the classification of angiosperms by Bentham and Hooker.				
<b>Semester</b>	3	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		3	-	1	-
<b>Pre-requisites, if any</b>	Basic knowledge in plant science				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Explain the plant morphological terminologies.	K	1
2	Appraise the system of angiosperm classification and nomenclature.	E	1,3
3	Demonstrate herbarium technique.	E	4
4	Identify plants to their families on the basis of key characters.	R	1,3
5.	Evaluate the medicinal and economic importance of selected angiosperms	E	5

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>1</b>	<b>Module 1 (15 hours)</b>			
	1.1	Morphology Leaf - simple, compound; venation and phyllotaxy. Flower as a modified shoot,	5	1

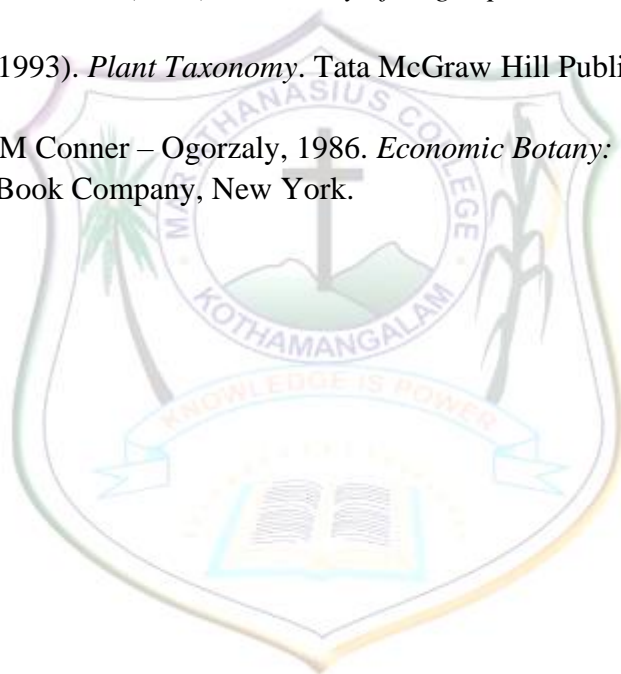
		structure of flower - floral parts, their arrangement, relative position; cohesion and adhesion of floral parts, symmetry of flowers; types of aestivation and placentation; floral diagram and floral formula.		
	1.2	Inflorescence Racemose - simple, spike, spadix, catkin, corymb, umbel and head; cymose - simple, monochasial, helicoid and scorpid; special types – cyathium, verticillaster.	6	1
	1.3	Fruits: Simple: Fleshy - drupe, berry, hesperidium; Dry - Dehiscent - legume, capsule; Indehiscent - Caryopsis, Cypsella, Schizocarpic - lomentum, carcerulus, regma, cremocarp with examples. Aggregate. Multiple: sorosis, syconus	4	1
	<b>Module 2 (15 hours)</b>			
<b>2</b>	2.1	Plant classification and Herbarium techniques Importance of plant classification, types of classification - artificial, natural and phylogenetic (brief account only)	3	2
	2.2	Binomial nomenclature; ICBN (Brief account only). Bentham and Hooker's system of classification (up to series) and its merits and demerits.	3	2
	2.3	Cytotaxonomy and chemotaxonomy (brief account only). Herbarium techniques; importance of herbarium.	2	2,3
	2.4	Angiosperm families Study of the following families of Bentham and Hooker's system of classification with special reference to major identifying characters and economic importance: Annonaceae, Malvaceae, Rutaceae, Leguminosae (Mimosaceae, Caesalpiniaceae and Fabaceae), Apiaceae (Umbelliferae)	7	2,4
	<b>Module 3 (15 hours)</b>			
<b>3</b>	3.1	Rubiaceae, Asteraceae (Compositae), Apocynaceae, Lamiaceae (Labiatae), Euphorbiaceae, Arecaceae (Palmae), Poaceae (Gramineae).	7	2,4


	3.2	Classification of economically important plants based on their uses. Study of the following groups of plants with special reference to their botanical name, family, morphology of useful part, economic products and uses: Cereals - Paddy, Wheat; Pulses - Green gram, Bengal gram; Tuber crops-Tapioca; Spices - Pepper, Cardamom; Beverages - Tea, Coffee; Oil yielding plants - Coconut, Groundnut; Fibre yielding plants - Cotton, Coir; Timber yielding plants - Teak, Rose wood; Latex yielding plants - Para rubber; Bio pesticides - Neem, Tobacco; Ornamental plants - Rose, Orchids, Anthurium	8	5
<b>Module 4 (Practical 30 hrs)</b>				
4	4.1	Examine vegetative and floral features of different plants and assign them to respective families mentioned in the syllabus.	15	1
	4.2	Prepare and submit any 5 herbarium sheets	5	3
	4.3	Study of the groups of plants mentioned in the economic botany syllabus with special reference to their botanical name, family, morphology of useful part, economic products and uses	10	5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc. <b>B. End Semester Examination</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

## References

1. Chrispeels (2003). *Plants, Genes, & Crop Biotechnology* Ch. 8. Plant Growth & Development (Botany fundamentals)
2. Eames A J, (1969). *Morphology of Angiosperms*. McGraw Hill, New York.
3. Hill A F, (1952). *Economic Botany: A Text book of Useful Plants and Plant Products*. Tata McGraw-Hill Publishing Company Limited, New Delhi.
4. Jain S K, (1987). *A Manual of Ethnobotany*.Scientific Publishers, Jodhpur.
5. Jain S K, 1987. *A Manual of Ethnobotany*.Scientific Publishers, Jodhpur
6. Kochhar S L, (1981). *Economic Botany in the Tropics*.Macmillan India Limited, Delhi.
7. Lawrence G H M, (1951). *Taxonomy of Vascular Plants*.Oxford & IBH, New Delhi.
8. Naik V N, (1984). *Taxonomy of Angiosperms*. Tata McGraw Hill Publishing Co, New Delhi.
9. Ones & Dangl (2006). *The plant immune system*. Nature 444:323-9
10. Pandey S N, S P Misra, (2008). *Taxonomy of Angiosperms*.Ane Books India, New Delhi.
11. Sharma O P, (1993). *Plant Taxonomy*. Tata McGraw Hill Publishing Co Ltd., New Delhi.
12. Simpson B S, M Conner – Ogorzaly, 1986. *Economic Botany: Plants in Our World*. McGraw Hill Book Company, New York.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Agri-based Microenterprises</b>					
<b>Type of Course</b>	<b>MDC</b>					
<b>Course Code</b>	<b>M24BO3MDC200</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	This course is designed to equip participants with the knowledge and skills necessary to establish and manage successful agri-based microenterprises. Focusing on key sectors such as organic farming, horticulture, tissue culture, and mushroom cultivation, the course provides a comprehensive understanding of sustainable and profitable agribusiness practices.					
<b>Semester</b>	3	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	-	-	45
<b>Pre-requisites, if any</b>	Basic knowledge about agriculture					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Summarize key principles in organic farming, horticulture, tissue culture and mushroom cultivation.	U	2,4
2	Illustrate fruits and vegetable technology including sustainable practices and business considerations.	U	4,5
3	Develop hands-on skills in composting techniques, artificial vegetative propagation practices, tissue culture techniques and mushroom cultivation	S	4
4	Apply the skills of organic farming, horticultural practices, tissue culture techniques, fruits and vegetable technology and mushroom cultivation, as an entrepreneurial venture.	A	4,5
5	Administer a mushroom cultivation project in a small scale level	A	4,5

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Introduction to Organic farming- Advantages of Manures over fertilizers. NPK value- Definition and significance.	2	1
	1.2	Common organic manures – bone meal, cow dung, poultry waste, oil cakes, Green manure (special reference to major element in the composition) Preparation of compost- vermicompost, vermiwash; familiarize KAMBA compost Biofertilizers-Definition and Types –, <i>Rhizobium</i> , <i>Mycorrhiza</i> , <i>Blue green algae</i> and <i>Azolla</i> . Activity-Hands on training on Vermicomposting Activity-Preparation of compost and establishing a small kitchen garden. Submit a report with geotagged photos	4	1,3
	1.3	Biological control Agents- <i>Trichoderma</i> , <i>Bacillus</i> ; Biopesticides – Tobacco and Neem decoction. Activity-Prepare and submit any one Biopesticide formulation.	2	1,3
	1.4	Types of soil, preparation of potting mixture, Garden tools and implements Methods of plant propagation- Sexual (seed propagation) and Asexual; Artificial methods (cutting, grafting, budding and layering); Use of growth regulators for rooting. Hands on training on Artificial methods of propagation - budding and grafting. Activity-Demonstration of budding (T and Patch)	7	1
2	<b>Module 2 (15 Hours)</b>			
	2.1	Gardening - Types of gardens– Ornamental and Landscape garden, kitchen garden Water garden and aquascaping, Aquarium plants and its propagation Garden components (Brief account only), Bonsai, terrarium, Kokedama. <b>Activity-</b> Submit a self made terrarium/ kokedama/ aquarium (use only natural materials)	6	1,3,4
	2.2	Concept of totipotency, definition of explant, callus. Infrastructure of a tissue culture laboratory. Solid and liquid media – basic components of tissue culture medium.	3	1,3



	2.3	Sterilization of explants'. inoculation and incubation. Micro propagation: different stages, organogenesis and embryogenesis Visit to a well established tissue culture lab/nursery/ mushroom cultivation unit	6	1,3
3	<b>Module 3 (15 Hours)</b>			
	3.1	Scope and Significance of Mushroom cultivation, Edible and poisonous mushroom. Health benefits	1	1
	3.2	Types of commercially cultivated mushrooms - button mushroom, oyster mushroom and milky mushroom, Spawn -Definition.	1	1
	3.3	Cultivation methodology of Oyster mushroom – using paddy straw and saw dust Layout and set up of a mushroom house (small scale). Processing of mushrooms and Value added products- mushroom - pickle, candy, dried mushroom	4	1,3,4
	3.4	Elementary knowledge on horticultural types of fruits and vegetables, Concept of shelf life and perishable fruits, Ripening and biological ageing, Storage and preservation concerns.	2	2
	3.5	Fruits preservation-Room temperature (Juice, syrup, squash), heat treatment (Jelly, jams), Dehydration (sun drying, application of sugar syrup,salt), freezing Vegetable preservation-packaging and storage, dehydration techniques, vegetable products ( flakes, chips, dried powder), frozen vegetables, Preservation by Canning and bottling. <b>Activity-</b> Prepare and submit any one fruit/vegetable product using methods prescribed in the syllabus Visit and submit an audio visual documentary on any one small scale entrepreneurship activity with reference to the skills mentioned in the syllabus Submit a proposal on any plant based entrepreneurship activity (other than mentioned in syllabus).	7	2,3,4
4	<b>Teacher specific course component</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar <hr/> <b>B. Semester End examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

### References

1. Sharma, Arun K. 2002. A Handbook of Organic farming. Agrobios, India.
2. Sathe, T.V. 2004, Vermiculture and Organic Farming. Daya Publishers.
3. Alvares, C. 1996. The Organic Farming Source Book. The Other India Press, Mapusa, Goa.
4. Gopal Chandha De, 2002. Fundamentals of Agronomy. Oxford and IBH Publishing House.
5. George Acquiah, 2004. Horticulture: Principles and Practices (II Edn). Prentice Hall. India.
6. Hudson T, Hartmann, Dale E Kester, 2001. Plant Propagation, Principles and Practices (VI Edn). Prentice Hall, India.
7. Kaul T N, 2002. Biology and Conservation of Mushroom. Oxford and IBH Publishing Co.
8. Pandey R K, S K Ghosh, 1996. A Handbook on Mushroom Cultivation. Emkey Publications.
9. Adams C R, Early M P, 2004. Principles of Horticulture. Elsevier, N. Delhi.
10. Barton West R, 1999. Practical Gardening in India. Discovery Pub. House, New Delhi.

### SUGGESTED READINGS

1. Edmond J B, Senn T L, Andrews F S, Halfacre P G, 1975. Fundamentals of Horticulture (IV Edn). TMH, New Delhi.
2. Purohit S S, 2005. Plant Tissue Culture. Student Edition.
3. Rema L P, 2006. Applied Biotechnology. MJP Publishers
4. Kalyan Kumar De, 1996. Plant Tissue Culture. New Central Book Agency (P) Ltd.
5. Razdan M K, 1995. Introduction to Plant Tissue Culture (II Edn). Oxford and IBH Publishing Co.
6. Sharma R R, 2005. Propagation of Horticultural Crops. Kalyani Publishers.
7. Singh B D, 1996. Biotechnology. Kalyani Publishers.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Bioethics and IPR</b>					
<b>Type of Course</b>	<b>VAC</b>					
<b>Course Code</b>	<b>M24BO3VAC200</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	This course focus on systematic outline of the bioethics and Intellectual Property Rights. This will provide the core principles in the interaction of IPR and Bioethics, also give overview of the domestic and international legal regime dealing with intellectual property law.					
<b>Semester</b>	3			Credits		Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
<b>Pre-requisites, if any</b>	Nil					

#### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Apply ethical principles in biological research	A	2
2	Discuss intellectual property rights and its benefit to society	U	2
3	Choose fundamental aspects of Intellectual Property Rights in development and management of innovative projects	A	2,4
4	Interpret knowledge on IPR, patents, patent regime and registration aspects in India and abroad	U	2,4
5	Appraise the current trends in IPR and Govt. steps in fostering IPR	E	2,4

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

#### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	Module 1 (15 Hours)			

	1.1	Bioethics – Need, issues (social and cultural) and applications; Misuse of modern molecular biology tools and techniques.	2	1
	1.2	Bioethics & Biodiversity: Convention on protecting Biodiversity, Protocols in exchanging Biological material across borders	2	1
	1.3	Issues and concerns pertaining to Genetically modified foods & food crops, Harm to the environment - potential impact of GMOs on the ecosystem.	3	1
	1.4	Bioethics in Medicine & Cloning: Protocols of ethical concerns related to prenatal diagnosis, gene therapy, organ transplantation, Xenotransplantation, ethics in patient care, informed consent	3	1
	1.5	Patenting biotech inventions: objective, applications, concept of novelty, concept of inventive steps	2	1 4
	1.6	Use of plants in research, human volunteers for clinical research, moral issues in patenting biotechnological inventions, Ethics related to professional streams.	3	1 2
	<b>Module 2 (15 Hours)</b>			
<b>2</b>	2.1	Meaning of Intellectual Property Rights – Introduction to TRIPS and WTO – IPR in India and the world	4	3 4
	2.2	Kinds of Intellectual property rights - Copy Right, Patent, Trade Mark, Trade Secret and trade dress	3	2 4
	2.3	Design, Layout Design, Geographical Indication, Plant Varieties and Traditional Knowledge.	3	4 5
	2.4	<b>Activity – 1</b> Geographical Indication - Meaning & significance of GI, How to file GI.	5	4, 5
	<b>Module 3 (15 hours)</b>			
<b>3</b>	3.1	Origin, Meaning of Patent, Types, Inventions which are not patentable	3	3, 4
	3.2	Registration Procedure, Rights and Duties of Patentee, Patent Infringement.	3	4, 5
	3.3	Copyright - Definition, Terms & Types of Copyright, Piracy. Information technology related IPR (computer software, database and data protection)	3	4, 5
	3.4	Trade Marks - Meaning & Nature of Trade Marks, Types, Infringement & Remedies, Offenses relating to Trade Marks.	3	4, 5

	3.5	<b>Activity – 2</b> Traditional Knowledge - Meaning, importance of TK, Sources of TK, TKDL (Traditional Knowledge Digital Library).	3	4, 5
<b>4</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar <b>B. Semester End examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## References

1. Ahuja, V.K. (2017). Law relating to Intellectual Property Rights. India, In: Lexis Nexis.
2. Benjamin A Pierce (2008). *Genetics: A conceptual approach* (IV Edn). W H Freeman and Company
3. Bernard R Glick, Jack J Pasternak, Cheryl L Pattern (2010). *Molecular biotechnology: Principles and applications of recombinant DNA*. ASM press.
4. Burton E Tropp (2012). *Molecular biology: Genes to Proteins* (IV Edn). Jones and Bartlett Learning.
5. Government of India's Patents Website: [patinfo.nic.in](http://patinfo.nic.in)
6. Intellectual property India: [www.ipindia.nic.in](http://www.ipindia.nic.in)
7. Neeraj, P. and Khusdeep, D. (2014). Intellectual Property Rights. India, In: PHI learning Private Limited.
8. Nithyananda, K.V. (2019). Intellectual Property Rights: Protection and Management. India, In: Cengage Learning India Private Limited.
9. Parulekar, Ajit & D'Souza, Sarita, (2006). Indian Patent Law : Legal and Business Implications, Macmillan India publication,
10. Santaniello, V., Evenson, R.E., Zilberman, D. and Carlson, G.A. (Eds) (2003). *Agriculture and Intellectual Property Rights*, University Press publication,
11. Sateesh, MK (2008), *Bioethics & Biosafety*, IK International publications,
12. Subramanian, N., & Sundararaman, M. (2018). Intellectual Property Rights – An Overview. Retrieved from <http://www.bdu.ac.in/cells/ipr/docs/ipr-eng-ebook.pdf>

13. USPTO Web Patent Databases at: [www.uspto.gov/patft](http://www.uspto.gov/patft)





## **SEMESTER IV**



**Mar Athanasius College (Autonomous), Kothamangalam**  
**FYUGP SYLLABUS**

<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Archegoniatas</b>				
<b>Type of Course</b>	<b>DSC A</b>				
<b>Course Code</b>	<b>M24BO4DSC200</b>				
<b>Course Level</b>	<b>200</b>				
<b>Course Summary</b>	The course offers a fundamental introduction to the evolutionary importance, classification, morphology, and distinctive features of Archegoniatas. Additionally, it provides a basic perspective on the ecological and economic relevance of Archegoniatas.				
<b>Semester</b>	4	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		3	-	1	-
<b>Pre-requisites, if any</b>	Basic knowledge in botany				

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Explain the general characters of archegoniatas	U	1
2	Classify archegoniatas to different plant groups	A	1,3
3	Compare the structure of gametophyte and sporophyte of Archegoniatas	AN	3
4	Assess the economic and ecological significance of Archegoniatas	E	5.6
5	Discuss the recent trends in archegoniate research	U	5

*\*Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

#### **COURSE CONTENT**

<b>Module</b>	<b>Units</b>	<b>Course description</b>	<b>Hrs</b>	<b>CO No.</b>
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1	<b>Module 1 (15 hours)</b>			
	1.1	Common features of archegoniates; Adaptation to thrive in land habit; Alternation of generations.	2	1
	1.2	Evolution/ transition of the sporophytic and gametophytic phase of Bryophytes, Pteridophytes and Gymnosperms	3	1
	1.3	Bryophytes General characters, Classification by Rothmaler 1951 (up to family)	2	1
	1.4	Type study: Morphology and anatomy of thallus, Reproduction and life cycle of <i>Riccia</i> , <i>Anthoceros</i> and <i>Pogonatum</i> (Developmental details of sex organs and embryo not needed).	7	1, 2, 3
	1.5	Ecological and economic importance of bryophytes.	1	4
2	<b>Module 2 (15 hours)</b>			
	2.1	Pteridophytes General characters, Classification of up to classes by Smith (2006) and PPG system (Brief account only)	3	1, 2
	2.2	Morphology and anatomy of thallus, Reproduction and life cycle of <i>Psilotum</i> , <i>Selaginella</i> and <i>Pteris</i> (Developmental details of sex organs not needed).	7	1, 3
	2.3	Stelar evolution in pteridophytes, Heterospory and seed habit	3	3
	2.4	Ecological and economic importance of Pteridophytes, Ornamental pteridophytes	2	4
3	<b>Module 3 (15 hours)</b>			
	3.1	Gymnosperms General characters Classification by Sporne (1965) (up to family), Brief account of classification by Christenhuez (2011)	4	1
	3.2	Morphology, anatomy, and reproduction of <i>Cycas</i> , <i>Pinus</i> and <i>Gnetum</i> (Developmental details of sex organs not needed)	8	1, 2, 3

	3.3	Economic importance of Gymnosperms, Ornamental Gymnosperms	3	4
4	<b>Module 4 Practical (30 hours)</b>			
	4.1	Conduct a field visit and submit a report with geo-tagged photos / images of gametophytes and/or sporophytes of archegoniates in your locality.	5	1, 2, 3, 4
	4.2	Collect three recent research publications on archegoniates and submit a comparison report.	2	5
	4.3	Collect, identify the genus, and submit gametophytes and/or sporophytes of any five archegoniates.	5	1, 2, 3
	4.4	<i>Riccia</i> and <i>Anthoceros</i> – Morphology and anatomy of thallus. <i>Pogonatum</i> - Morphology of the sporophyte and gametophyte	6	1, 2, 3
	4.5	<ul style="list-style-type: none"> <li>● <i>Psilotum</i>- Morphology of sporophyte and synangium</li> <li>● <i>Selaginella</i>- Morphology of sporophyte, transverse section of the stem.</li> <li>● <i>Pteris</i>- Morphology of sporophyte, transverse section of sporophyll</li> </ul>	8	1, 2, 3
	4.6	<ul style="list-style-type: none"> <li>● <i>Cycas</i>- Morphology of coralloid roots and reproductive structures; TS of leaflet.</li> <li>● <i>Pinus</i>- Morphology of male and female cones; TS of the needle.</li> <li>● <i>Gnetum</i> - Morphology of male and female cones</li> </ul>	4	1, 2, 3
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b></p> <p>Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
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<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b></p> <p><b>A. Continuous Comprehensive Assessment (CCA)</b>  Theory Total = 25 marks  Quiz, Test Papers, seminar  Practical Total = 15 marks  Lab performance, record, field report etc.</p>
	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10marks  Part B (Short essay) – 4 out of 6 x 5 = 20marks  Part C (Long essay) – 2 out of 4 x 10 = 20marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>

### References

1. Bhatnagar, S.P. & Moitra, A. (1996). Gymnosperms. New Age International (P) Ltd Publishers, New Delhi, India.
2. Chandra S, Srivastava M. (2003). Pteridology in New Millennium. Kluwer Academic Publishers.
3. Chopra R. N., Kumar, P. K. (1988). Biology of Bryophytes. Wiley Eastern Ltd. New Delhi.
4. Coutler J. M., Chamberlain C. J. (1958). Morphology of Gymnosperms. Central book depot. Allahabad.
5. Parihar N. S. (1965). An Introduction to Bryophyta. Central Book Depot, Allahabad.
6. Parihar N. S. (1977). Biology and Morphology of Pteridophytes. Central Book Depot, Allahabad.
7. Rashid A. (1976). An Introduction to Pteridopyta. Vikas publ. Co., New Delhi.
8. Shaw J. A., Goffinet B. (2000). Bryophyte Biology. Cambridge University Press.
9. Smith G. M. (1938). Cryptogamic Botany Vol. II. Bryophytes and pteridophytes. McGraw Hill Book Company, London.
10. Sporne K. R. (1967). The Morphology of Bryophytes. Hutchinson University Library, London.
11. Sporne K. R. (1967). The Morphology of Gymnosperms. Hutchinson and Co. Ltd. London.
12. Sporne K. R. (1967). Morphology of Pteridophytes. Hutchi University Library, London.
13. Sreevastava H. N. (1980). A Text Book of Gymnosperms. S Chand and Co. Ltd., New Delhi.
14. Sreevastava H. N. A textbook of Pteridophyta. S Chand and Co., New Delhi.
15. Vasishta B. R. (1993). Pteridophyta. S Chand and Co., New Delhi.
16. Vasishta B. R. Bryophyta. S Chand and Co. New Delhi.
17. Vasishta P. C. (1980). Gymnosperms. S Chand and Co., Ltd., New Delhi.
18. Watson E.V. (1971). The structure and life of Bryophytes. Hutchinson University Library, London.

## Websites

<http://www.artdata.slu.se/guest/SSCBryo/SSCBryo.html>

<http://www.northernontarioflora.ca/links.cfm?val=bryophytes>

<http://bryophytes.plant.siu.edu/>

<http://worldofmosses.com/>

<http://www.unomaha.edu/~abls/>

<http://www.anbg.gov.au/bryophyte/index.html>

<http://www.bryoecol.mtu.edu/>

<http://www.mobot.org/MOBOT/tropicos/most/Glossary/glosefr.html>

[http://www.fairhavenbryology.com/Master\\_Page.html](http://www.fairhavenbryology.com/Master_Page.html)


<http://www.gymnosperms.org/>

<http://www.plantapalm.com/vce/toc.htm>

<http://www.cycad.org/conservation.htm>

[http://allwebhunt.com/cgi.cfm/Top/Science/Biology/Flora\\_and\\_Fauna/Plantae/Cycadophyta/Cycadopsida/Cycadaceae/Cycas](http://allwebhunt.com/cgi.cfm/Top/Science/Biology/Flora_and_Fauna/Plantae/Cycadophyta/Cycadopsida/Cycadaceae/Cycas)



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Plant Cell and Molecular Biology</b>				
<b>Type of Course</b>	<b>DSC A</b>				
<b>Course Code</b>	<b>M24BO4DSC201</b>				
<b>Course Level</b>	<b>200</b>				
<b>Course Summary</b>	Cell and Molecular biology play a crucial role in shaping understanding of life. The course emphasizes the basic principles that buttress the processes unique to living organisms at the molecular and cellular levels. Students will acquire a basic understanding of architecture of plant cells, organization of genetic material, the storage, transfer, and regulation of genetic information etc. The course envisages the application of modern molecular and cellular biology in Plant Sciences and provides a solid foundation for further studies in the areas of molecular life sciences, bioengineering, and biotechnology.				
<b>Semester</b>	4	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		3	-	1	-
<b>Pre-requisites, if any</b>	Basic understanding of cell structure in plants, process of cell division and knowledge of experiments that led to the discovery of genetic material				

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Outline the historical developments in cell and molecular biology	U	3
2	Illustrate the structure and function of plant cell wall and	A	3, 5

	cell organelles		
3	Describe the function of the nucleus and chromosome condensation process and their role in heredity	U	3, 4, 5
4	Assess the gene regulatory network and inheritance in organisms and the role of enzymes regulating cell activities	E	3
5	Examine how Cell division and programmed cell death occur within a plant cell	An	3
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
	<b>Module 1 (15 hours)</b>			
	1.1	History and Scope of Cell and Molecular biology; Major developments in Cell and Molecular Biology.	1	1
	1.2	Composition, structure and functions of cell membrane – Fluid mosaic model	2	2
1	1.3	Structure and Major Functions of the following cell organelles: Endoplasmic Reticulum, Lysosomes, Dictyosomes, Vacuole, Ribosomes (Brief Account) and Cytoskeleton. Major Components and Definitions of GERL and Endomembrane System. Structure and Major Functions of Semi-autonomous Cell Organelles - Chloroplast, Mitochondria.	6	2
	1.4	Ultra-Structure of Nucleus, Nuclear Envelope, Nuclear Pore Complex (NPC). Structure and Function of Nuclear lamina and Nucleolus. Morphology of a typical chromosome, Organization of genetic material in chromosomes. Structural organization: Histones, Non-histone proteins, Nucleosomes, Chromatosomes. Chromatin organization in eukaryotes	6	3
2	<b>Module 2 (15 hours)</b>			

	2.1	Special Chromosomes: Structure and Function of Polytene and Lamp brush chromosomes.	2	3
	2.2	Types and Organization of Chromatin: Heterochromatin, Euchromatin, Karyotype, Idiogram	1	3
	2.3	Eukaryotic Cell cycle (G1, S, G2, M) Mitosis, meiosis and their significance	3	5
	2.4	Programmed Cell Death (Apoptosis) (Overview). <b>Activity:</b> Students may submit appropriate illustrations with short descriptions to explain the different stages of mitosis and meiosis	2	5
	2.5	Basic understanding of Genetic material Types of DNA: A, B and Z DNA, Detailed structure of B DNA, Plastome - Chloroplast DNA. Types and functions of RNA: hnRNA, mRNA, tRNA, rRNA, snRNA and microRNA <b>Activity:</b> Prepare a comparative account on the types of RNA and submit for evaluation	3	4
	2.6	DNA replication (prokaryotic): Messelson and Stahl experiment, Role of enzymes - DNA Polymerases, Primases, Helicases, Ligases and DNA Topoisomerases. Brief account of enzymes involved in eukaryotic DNA replication	4	4
<b>Module 3 (15 hours)</b>				
		Point Mutations: Definitions of Transition Mutations, Transversion Mutations, Silent mutations, Missense mutations, Nonsense Mutations. Molecular basis of point mutations. Definition and Significance of Frameshift mutations. Significance of DNA repair mechanisms in cells. <b>Activity:</b> Discuss how mutation in a single nucleotide leads to altered phenotype citing suitable examples.	4	4
<b>3</b>	3.1	Gene expression: Central dogma of molecular biology. Basic mechanism of Transcription in Prokaryotes. Perspective of transcription in Eukaryotes: Split genes, Introns, Exons, Spliceosomes (Definitions and significance). Genetic code, Wobble hypothesis. Post transcriptional modification of mRNA in Eukaryotes; capping, poly adenylation and splicing	7	4

		(brief account). Translation in Prokaryotes and Eukaryotes (brief account)		
	3.2	Regulation of gene expression in prokaryotes by Operons: <i>Lac</i> and <i>Trp</i> operon.	3	4
	3.3	Endosymbiont hypothesis (Overview), Significance of chloroplast and nuclear DNA in the biosynthesis of RUBISCO.	1	4
	<b>Module 4 Practical (30 hours)</b>			
<b>4</b>	4.1	Study of mitosis by squash preparation of <i>Allium</i> sp. root tip	30	2, 3, 5
	4.2	Calculate mitotic index of root tips prepared by squash preparation		
	4.3	Identification of various stages of meiosis I using appropriate illustrations		
	4.4	Isolation of plant DNA from appropriate plant specimen		
	4.5	Demonstration (any one) of <ul style="list-style-type: none"> <li>● Cell viability using tri-phenyl tetrazolium chloride (TTC).</li> <li>● Cell counting using hemocytometer</li> <li>● Observation of cyclosis and Chloroplast in leaf of <i>Hydrilla</i> or Staminal hairs of <i>Rheo discolor</i></li> </ul>		
	4.6	Submit a report of beneficial/harmful aspects of mutation.		
<b>5</b>	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
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<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc
	<b>B. End Semester Examination</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

### References

1. Aggarwal S K, 2009. Foundation Course in Biology (II Edn). Ane Books Pvt. Ltd.
2. Avinash, Kakoli Upadhyay, 2005. *Basic Molecular Biology*. Himalaya Publishing House, Mumbai.
3. Cohn N S, 1964. Elements of Cytology. Brace and World Inc., New Delhi.
4. Darlington C D, 1965. Cytology. Churchill, London.
5. Darnel J, Lodish, Hand Baltimore D, 1991. Cell and molecular biology. Lea and Fibiger, Washington.
6. De Robertis E D P, Robertis E M P, 1991. Cell and molecular biology. Scientific American books.
7. Dobzhansky B, 1961. Genetics and origin of species. Columbia University Press, New York.
8. Gardner E J, Snustad D P, 1984. Principles of Genetics. John wiley, NewYork.
9. Gerald Karp, 2006. Cell Biology. McGraw Hill company.
10. Gupta P K. Genetics. Rastogi Publications.
11. Lewin B, 1999. Genes. Oxford University Press, NewYork.
12. Lewis W H, 1980. Polyploidy. Plenum Press, NewYork.
13. Roy S C, Kalayan Kumar De, 1997. Cell biology. New central Boos, Calcutta.
14. Sandhya Mitra, 1998. Elements of Molecular biology. Macmillan, India Ltd.
15. Sharma A K, Sharma A, 1980. Chromosome technique: Theory and practice. Aditya Books, NewYork.
16. Veer Bala Rastogi, 2008. Fundamentals of Molecular Biology. Ane Books Pvt. Ltd.
17. Wayne M Becker, Lewis J Kleinsmith, Jeff Hardin, 2004. The World of Cell. Pearson Education.
18. Waseem Ahammede Ffaridi, 2013. Genetics and Genomics. Pearson.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Introduction to Biotechnology</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO4DSE200</b>				
<b>Course Level</b>	<b>200</b>				
<b>Course Summary</b>	The course intended to provide an understanding on the process and the current developments in the field of Biotechnology. The course also highlights the application of biotechnology for human welfare..				
<b>Semester</b>	4	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		4	-	-	-
<b>Pre-requisites, if any</b>	Basic knowledge in cell and molecular biology				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Understand the fundamental principles and techniques of plant tissue culture.	U	3
2	Demonstrate proficiency in the practical aspects of micropropagation.	U	4,5
3	Evaluate the various methods of micropropagation and assess their applications in plant propagation and crop improvement.	E	3,4
4	Illustrate the principles of recombinant DNA technology (rDNA) to manipulate genetic material.	A	2,4,5
5.	Discuss the diverse applications of biotechnology.	U	2,5

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>1</b>	<b>Module 1 (15 hours)</b>			
	1.1	Biotechnology - an overview. Plant tissue culture - basic concepts, totipotency,	3	1

		differentiation, de-differentiation and re-differentiation.		
	1.2	Tissue culture media: Components, role of plant growth regulators in tissue culture. Types of media and examples -solid, liquid, MS, White's, B5 media; Sterilization of equipments, glassware and culture medium, surface sterilization of explants.	5	1,2
	1.3	Micropropagation: Definition, selection and sterilization of explant, inoculation, culturing, hardening and transplantation.	4	3
	1.4	Learning activity 1. Familiarize with basic steps in micropropagation	3	2
	<b>Module 2 (15 hours)</b>			
2	2.1	Methods of micropropagation: - axillary bud proliferation, adventitious regeneration – shoot organogenesis and somatic embryogenesis - direct and indirect. Callus and cell suspension culture.	6	2,3
	2.2	Types of culture and applications: Embryo culture, anther and pollen culture, Protoplast culture. Production of synthetic seeds.	4	3
	2.3	Advantages and disadvantages of micropropagation - somaclonal variations.	2	3
	2.4	Learning activity 1. Immobilization of whole cells or tissues in sodium alginate	3	3
	<b>Module 3 (15 hours)</b>			
3	3.1	rDNA technology – introduction. Tools used in rDNA technology. Restriction endonucleases and ligases; Cloning vectors and their desirable properties; plasmids, cosmids, phage vectors, Phasmids, artificial chromosomes.	5	4
	3.2	Steps in rDNA technology Isolation of DNA, Cutting and joining of DNA molecules. Transformation and selection of transformants - using antibiotic resistances markers and complementation.	5	4
	3.3	Agarose gel electrophoresis, Southern blotting, PCR and its applications. Uses of refrigerated centrifuges, UV trans-illuminator, gel documentation system and Laminar Air Flow chamber (brief account only).	5	4
	<b>Module 4 (15 Hours)</b>			


4	4.1	Biotechnology in agriculture: Bt crops, nematode resistant crops, herbicide resistant crops, biofortification	4	5
	4.2	Biotechnology in medicine: Human insulin and gene therapy	3	5
	4.3	Biotechnology in marine resources: pharmaceuticals, enzymes	3	5
	4.4	Biotechnology in dairy processing: Bio-preservation (Bacteriosin), Probiotics	3	5
	4.5	Biotechnology in environmental monitoring – super bugs	2	5
5	<b>Teacher specific course components</b>			
<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.			
<b>Assessment types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar <b>B. End Semester Examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks			

## References

1. David W Mount (2001). *Bioinformatics: Sequence and genome analysis*. CBS publishers & distributors.
2. David P Clark (2010). *Molecular biology*. Elsevier.
3. G Smita Rastogi and Neelam Pathak. Genetic Engineering. Oxford Higher Education.
4. George Acquah (2005). *Understanding biotechnology*. Pearson.
5. James D. Watson, Amy A. Caudy, Richard M. Myers, Jan A. Witkowski (2007). *Recombinant DNA* (III Edn). W H Freeman.
6. Jeremy W Dale, Malcolm von Schantz (2002). *From genes to genomes*. John Wiley & Sons Ltd.
7. Leland H Hartwell, Leroy Hood, Michael L Goldberg, Ann E Reynolds, Lee M Silver, Ruth C Veres (2004). *Genetics: From genes to genomes* (II Edn). McGraw Hill.
8. R Keshavachandran and K V Peter. Plant Biotechnology: Methods in Tissue Culture and Gene Transfer. Orient Blackswan.
9. Robert F Weaver (2002). *Molecular biology* (II Edn). McGraw Hill.
10. Robert J Brooker (2009). *Genetics: Analysis & principles* (III Edn). McGraw Hill.

11. S. B. Primrose, R. M. Twyman (2006). *Principles of gene manipulation and genomics* (VII Edn). Blackwell publishing.
12. Susan R. Barnum (1998). *Biotechnology: an introduction*. Thomson Brooks/cole.
13. William J Thieman, Michael A Palladino (2009). *Introduction to biotechnology* (II Edn). Pearson.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Phytotechnology</b>					
<b>Type of Course</b>	<b>DSE</b>					
<b>Course Code</b>	<b>M24BO4DSE201</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	The present course focuses mainly on common herbal plants in our locality, their morphological peculiarities, nutritive and medicinal properties. This course also aims for the extraction of major principles of herbal plants in their crude form, also their cultivation, conservation practices and their applied aspects (Herbal Dyes, Organic pesticides, Biofuels).					
<b>Semester</b>	4	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		4	-	-	-	60
<b>Pre-requisites, if any</b>	Knowledge in plant biology					

#### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Identify the common herbal plants in our locality.	U	1
2	Familiarize the cultivation practices and conservation of the herbal plants and homely application against common diseases.	U	2,3
3	Examine the different herbal plants based on the medicinal and nutritive values.	An	3,4
4	Develop the skills for extracting the various phytochemicals in crude form.	C	5

5	Evaluate the major chemical components present in the selected herbal plants.	E	3
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>1</b>	<b>Module 1 (15 hours)</b>			
	1.1	Overview of Herbal Medicine: Definition, history, and significance in traditional and modern healthcare.	2	1
	1.2	Botanical Fundamentals: Introduction to plant anatomy, morphology, and physiology relevant to herbal medicine.	3	1
	1.3	Plant Identification: Techniques for identifying medicinal plants in the wild and cultivated settings. Ethnobotany: Exploration of cultural and traditional uses of plants in various regions.	4	3
	1.4	Indian medicine system: Ayurveda, Siddha, Unani, Homeopathy	6	3
<b>2</b>	<b>Module 2 (15 hours)</b>			
	2.1	Introduction to Cultivation: Basics of soil preparation, planting, and maintenance for medicinal plants.	3	1, 5
	2.2	Environmental Factors: Understanding the impact of climate, soil type, and geography on plant growth. Organic Cultivation Practices: Sustainable and environmentally friendly methods for growing medicinal plants	5	4
	2.3	Harvesting Techniques: Timing, methods, and best practices for harvesting medicinal plants to preserve potency.	4	2
	2.4	Post-Harvest Handling: Drying, storage, and processing techniques to maintain quality and efficacy.	3	2

3	<b>Module 3 (15 hours)</b>			
	3.1	Principles of Extraction: Understanding various extraction methods including maceration, infusion, decoction, and distillation.	3	4
	3.2	Solvents for extraction: Different solvents, solvent selection, concentration, and extraction efficiency.	4	4
	3.3	Quality Control: Methods for assessing the quality, purity and potency of herbal extracts..	4	4
	3.4	Formulation Development: Basics of herbal formulation including dosage and forms, compatibility, and stability	4	5
4	<b>Module 4 (15 hours)</b>			
	4.1	Visit to a well-maintained herbal garden such as JNTBGRI, Malabar Botanical Garden and other recognized institutes. (1 day)	7	1, 2
	4.2	Visit to scientific labs regarding extraction, identification of phytochemicals. (1 day)	5	1, 2
	4.3	Submit any 5 medicinal plants with their medicinal uses	3	1, 2
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar



	<p><b>B. End Semester Examination</b>  Theory Total = 70 marks, Duration 2 hrs  Part A (Short answer) – 10 out of 12 x 2 = 20 marks  Part B (Short essay) – 6 out of 9 x 5 = 30 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>
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### References

1. Daniel, M., Arun, A., Raole, V.M. (2007). *Herbal Technology: Recent Trends and Progress*, Scientific Publishers.
2. Sujanal, P.; Prabhu N.H., Pius, O.L., Sajeev, V.B. (2008). *Susthira Oushadha Sasya Krishi*, State Medicinal Plants Board, Thrissur, Kerala.
3. Agarwal, P., Alok, S., Fatima, A and A. Verma. (2013) *Current scenario of Herbal Technology worldwide: An overview*. Int J Pharm Sci Res; 4(11): 4105-17.
4. Dotto, J.M., S. A. Abihudi. (2021). *Nutraceutical value of Carica papaya: A review*. Scientific African 13 e00933.
5. Kokate, C.K., Purohit, A.P., Gokhale, S. B. (1999). *Pharmacognosy*. Nirali Prakashan.
6. Green, A. (2000) *Principles of Ayurveda*, Thomsons, London.
7. Arber, A. (1999). *Herbal plants and Drugs*, Mangal Deep Publications.
8. Chopra, R.N., Nayar, S.L., and Chopra, I.C., (1956). *Glossary of Indian medicinal plants*, C.S.I.R, New Delhi.
9. Sivarajan, V.V., and Balachandran, I.(1994). *Ayurvedic drugs and their plant source*. Oxford IBH publishing Co.
10. Chen, SL., Yu, H., Luo, HM. Wu, Q., Li, C., & Steinmetz, A., (2016) *Conservation and sustainable use of medicinal plants: problems, progress, and prospects*. Chin Med 11, 37. <https://doi.org/10.1186/s13020-016-0108-7>.
11. [https://www.researchgate.net/publication/327304552\\_an\\_overview\\_of\\_major\\_classes\\_of\\_phytochemicals\\_their\\_types\\_and\\_role\\_in\\_disease\\_prevention](https://www.researchgate.net/publication/327304552_an_overview_of_major_classes_of_phytochemicals_their_types_and_role_in_disease_prevention)
12. [https://www.researchgate.net/deref/https%3A%2F%2Fdoi.org%2F10.31881%2FJLR.2021.09?\\_tp=eyJjb250ZXh0Ijp7ImZpcnN0UGFnZSI6InB1YmxpY2F0aW9uIn9](https://www.researchgate.net/deref/https%3A%2F%2Fdoi.org%2F10.31881%2FJLR.2021.09?_tp=eyJjb250ZXh0Ijp7ImZpcnN0UGFnZSI6InB1YmxpY2F0aW9uIn9)



**Mar Athanasius College (Autonomous), Kothamangalam**

**FYUGP SYLLABUS**

<b>Programme</b>	<b>BOTANY</b>					
<b>Course Name</b>	<b>Plant anatomy and Microtechnique</b>					
<b>Type of Course</b>	<b>DSC B</b>					
<b>Course Code</b>	<b>M24BO4DSC202</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	This course is an introduction to the basic internal structure of plants, including their cells, tissues, and organs. Students get an insight into the organization of various tissue systems in a plant. Knowledge about the anatomical structure of stem and root of monocot and dicots, changes leading to secondary growth and a basic understanding of wood anatomy will be attained by the end of this course. Students will also develop skills in microscopy and Microtechnique for efficiently handling plant specimens.					
<b>Semester</b>	4	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Basic knowledge about plant structure					

**COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PO No</b>
1	Discuss the structural features of plant cell, tissues, vascular bundles and stomata	U	1
2	Describe the anatomy and ecological significance of epidermal and secretory structures	U	1
3	Differentiate stelar patterns in stem and root of vascular plants with normal and anomalous secondary growth	An	1,3
4	Demonstrate the methods used in preparing microscopic slides of plant specimens	A	4
5	Combine the anatomical knowledge with other research disciplines in plant science	An	5

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

**COURSE CONTENT**

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Introduction and scope of Plant Anatomy; Structure of plant cell, structure of cell wall	2	1
	1.2	Tissues and tissue systems: Structure and functions of meristematic tissues and permanent tissues (simple and complex), concept of tissue systems: dermal tissues, ground tissues and vascular tissues.	4	2
	1.3	Secretory tissues: glands, glandular hairs, nectaries, hydathodes, Laticifers –articulated and non-articulated	4	2
	1.4	Classification of meristems: Based on location (apical, intercalary and lateral), origin (promeristem, primary and secondary meristem) and plane of division (rib, plate and mass meristem). Apical meristems: Structure of shoot and root apex	3	1
	1.5	Vascular bundles- types: collateral, bicollateral, concentric and radial	1	1,3
	1.6	Stomata types- anomocytic, anisocytic, paracytic and diacytic	1	1
2	<b>Module 2 (15 hours)</b>			
	2.1	Primary structure of stem, root and leaf: Dicot and Monocot.	6	3
	2.2	Secondary Growth: Normal Secondary growth in stem and root.	4	3
	2.3	Wood anatomy: hard wood, soft wood. Growth rings-ring porous and diffuse porous wood. sap wood and heart wood, Tyloses	3	1, 2, 3,5
	2.4	Applications of anatomy in Plant systematics, forensics, Pharmacognosy and Dendrochronology	2	5
3	<b>Module 3 (15 hours)</b>			
	3.1	Microtechnique - Brief Introduction, Scope	1	4
	3.2	Microscopy: Simple and Compound, Parts of compound and dissection microscope	2	4
	3.3	Killing and fixing: Killing and fixing agents and their composition (Farmer's fluid and FAA)	2	4

	3.4	Sectioning-Free hand sectioning-TS, RLS, TLS	2	4
	3.5	Microtomy: rotary and sledge microtome, serial sectioning, applications of microtomy.	2	4
	3.6	Staining – types of stains: acidic, basic and neutral, vital stains, single staining, double staining, Preparation of stains-acetocarmine, safranin and crystal violet	3	4
	3.7	Mounting-temporary, semi-permanent and permanent, mounting media- glycerine, canada balsam, DPX	3	4
	<b>Module 4 Practical (30 hours)</b>			
<b>4</b>	4.1	Primary structure of stem and root (dicot and monocot)	6	1, 3
	4.2	Anatomy of dicot and monocot leaf (mesophytes)	6	1, 3
	4.3	Identification of stomatal types	6	1, 3
	4.4	Normal Secondary growth in dicot stem and root	6	1, 3
	4.5	Familiarize with stains, mounting medium and instruments used in microtechnique	6	4
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc
	<b>B. End Semester Examination</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

## References

1. Beck, C. B. (2010). *An Introduction to Plant Structure and Development-Plant Anatomy for the Twenty-First Century*. Cambridge University Press, ISBN: 9781139486361.
2. Bonham, D. (2018). *Plant Anatomy*. Larsen & Keller education, ISBN: 9781635496468
3. Crang, R., Lyons-Sobaski, S., & Wise, R. (2018). *Plant Anatomy A Concept-Based Approach to the Structure of Seed Plants*. Springer International Publishing, ISBN: 9783319773155.
4. Cutler, D. F., Botha, T., & Stevenson, D.W. (2009). *Plant Anatomy: An Applied Approach*, Wiley, ISBN: 9781444300468.
5. Esau, K. (2006). *Anatomy of seed plants*, 2<sup>nd</sup> Edition, Wiley India Pvt. Limited, ISBN: 9788126508204
6. Evert, R.F., Eichhorn, S.E. (2006). *Esau's Plant Anatomy Meristems, Cells, and Tissues of the Plant Body: Their Structure, Function, and Development*, Wiley, ISBN: 9780470047378
7. Foster, A. S. (2015). *Practical plant anatomy*. Creative Media Partners, LLC, ISBN: 9781341784507
8. Jain, A.P., & J. K. Maheshwari J. K. (2001). *Recent researches in plant anatomy and morphology*, Scientific Publishers, ISBN: 9788172332693
9. Marimuthu, R. (2019). *Microscopy and Microtechnique*. MJP Publisher, ISBN: 9788180940354
10. Mishra, S.R. (2009) *Understanding Plant Anatomy*, Discovery Publishing House Pvt. Limited, ISBN: 9788183564571.
11. Pandey, B.P. (2001). *Plant anatomy*, S Chand Limited. ISBN: 9788121901451.
12. Peterson, R. L., Peterson, C. A., & Melville, L.H. (2008). *Teaching plant anatomy through creative laboratory exercises*. NRC Press, ISBN: 9780660197982
13. Ruzin, S. (1999). *Plant Microtechnique and Microscopy*. Oxford University press Inc., Oxford
14. Sanderson, J. (2020). *Biological Microtechnique*. CRC Press, ISBN: 9781000140941
15. Yeung, E.C. T., Stasolla, C., Sumner, M.J., & Huang, B. Q. (Eds.) (2015). *Plant microtechniques and protocols*, Springer.



**Mar Athanasius College (Autonomous), Kothamangalam**

**FYUGP SYLLABUS**

<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Biofertilizers and Biocontrol Agents</b>					
<b>Type of Course</b>	<b>SEC</b>					
<b>Course Code</b>	<b>M24BO4SEC200</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	The course Biofertilizers and Biocontrol agents is designed in such a way to develop skills in graduate-level students to prepare various types of eco – friendly bioformulations for sustainable agriculture. The course deals with important categories of micro and macroscopic agents that can act as biofertilizers and biocontrol agents, their preparation and application methods.					
<b>Semester</b>	4	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	-	-	45
<b>Pre-requisites, if any</b>	Knowledge in biology					

**COURSE OUTCOMES (CO)**

CO No.	Expected Course Outcome	Learning Domains *	PSO No.
1	Explain the different concepts and approaches of sustainable agriculture	U	2
2	Articulate the knowledge of various organisms in sustainable agricultural practices.	A	2,5
3	Compare and evaluate the role of various components of bioformulations.	An	4
4	Practice bioformulation production and their application methods.	A	4
5	Illustrate the knowledge acquired to develop compost from household waste.	A	2,4

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

**COURSE CONTENT**

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			

	1.1	<p>Sustainable agricultural practices: Definition and concepts, Different approaches of sustainable agriculture/ natural farming: organic farming, Whole farm planning, Minimal cultivation, Environment-friendly agriculture .</p> <p><b>Learning activity:</b></p> <ol style="list-style-type: none"> <li>1. Group discussion/Debate – conventional and sustainable agriculture.</li> <li>2. Prepare and submit a report on various agricultural practices in an agricultural field based on a field visit.</li> </ol>	5	1
	1.2	<p>Types of household wastes, manufacturing of biofertilizers using household waste: Procedure – sorting of household waste, composting (biodegradation) – enzymatic method, backward method, composting by microbial inoculation and biological beneficial organisms. Methods to improve the quality of household compost – mineral additives and plant hormones.</p> <p><b>Learning activity:</b></p> <p>Conduct the preparation of compost from household wastes using the Garden pot composting method or Pipe composting method.</p>	10	5
	<b>Module 2 (15 Hours)</b>			
2	2.1	<p>Brief history and concept of Biofertilizers, status, scope, and importance of Biofertilizers. Classification of Biofertilizers – (a) Nitrogen-fixing (b) Phosphorus-solubilising bio-fertilizers or PSB (c) Potash-solubilising bio-fertilizers (d) Plant growth promoting microbes (PGPR). Major groups of microbial biofertilizers – Bacteria (Rhizobium, Pseudomonas) and Fungi (AM mycorrhiza and ectomycorrhiza). Blue-green algae (BGA), Plant-based biofertilizer – Azolla.</p> <p><b>Learning activity:</b></p> <p>1. Field exploration for macroscopic biofertilizers.</p>	8	2
	2.2	<p>Brief history and development of Biocontrol agents, Types: Macro biocontrol agents – egg parasitoids (<i>Trichogramma</i>) and Microbial biocontrol agents – (a) Bioinsecticides – <i>Bacillus thuringiensis</i>, (b) Bio fungicides – <i>Trichoderma</i>. Plant-based biopesticides: Neem and tobacco-based products (Brief account only). Commercially available botanical biopesticides – Pyrethrum, <i>Eucalyptus</i> essential oil.</p> <p><b>Learning activity:</b></p> <ol style="list-style-type: none"> <li>1. Collect recipes, uses and modes of action of various types of plant-based biopesticides.</li> </ol>	7	3

		2. Conduct a presentation/group discussion on the recipes they collected.		
3	<b>Module 3 (15 Hours)</b>			
	3.1	Bioformulations: Definition, components (Active ingredient, carrier material, additive), Types of bioformulations: Solid (granules, wettable powders, wettable granules, dust) liquid (suspension concentrate), encapsulation. Bioformulations for the uptake of nutrients like - Nitrogen, Phosphorus, Potassium, and Iron. Bioformulations as biocontrol agents/ biopesticides: Bacterial, Fungal and Viral. <b>Learning activity:</b> 1. Visit a biofertilizer/ pesticide manufacturing industry. 2. Make a comparison chart of the components of commercially available biofertilizers/ biopesticides.	8	4
	3.2	Rhizobium-based biofertilizer production steps: Selection of strain, Mass culture, Carrier preparation, Inoculant production. Formulation of <i>Trichoderma</i> as biocontrol agents. Delivery methods of various biofertilizer and biocontrol agents – seed treatment, soil amendment, soil drench, aerial spraying, root dip method. <b>Learning activity:</b> 1. Field exploration for plants with root nodules 2. Practice various methods of biofertilizer and biocontrol agent application.	7	4
4	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar



	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x1 =10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>
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## References

1. Mason, J. (2003). Sustainable agriculture. Landlinks Press.
2. Kaushik, B. D., Kumar, D., & Shamim, M. (Eds.). (2019). Biofertilizers and biopesticides in sustainable agriculture. CRC Press.
3. Rai, M. (Ed.). (2006). Handbook of microbial biofertilizers. CRC Press.
4. Borkar, S. G. (2015). Microbes as bio-fertilizers and their production technology. Woodhead Publishing India Pvt, Ltd.
5. Bosch, R., Messenger, P. S., & Gutierrez, A. P. (1982). An introduction to biological control (No. 632.96/B742). Springer US.
6. El-Wakeil, N., Saleh, M., & Abu-hashim, M. (Eds.). (2020). Cottage industry of biocontrol agents and their applications: practical aspects to deal biologically with pests and stresses facing strategic crops (pp. 133-155). Springer International Publishing.
7. Nollet, L. M., & Rathore, H. S. (Eds.). (2023). Biopesticides handbook. CRC Press.
8. Rajeshwari, R., & Appanna, V. (Eds.). (2021). Biopesticides in Horticultural Crops. CRC Press.
9. Singh, D. (Ed.). (2014). Advances in plant biopesticides. Springer.
10. Dalavayi Haritha, M., Bala, S., & Choudhury, D. (2021). Eco-friendly plant based on botanical pesticides. Plant Archives, 21(1), 2197-2204.
11. Hall, F. R., & Menn, J. J. (1999). Biopesticides: use and delivery. Humana Press Inc..
12. Nick, B. & Glare, T. (2020). Biopesticides for sustainable agriculture. Burleigh Dodds Science Publishing.
13. Arora, N. K., Mehnaz, S., & Balestrini, R. (Eds.). (2016). Bioformulations: for sustainable agriculture (pp. 1-283). Berlin: Springer.
14. Giri, B., Prasad, R., Wu, Q. S., & Varma, A. (Eds.). (2019). Biofertilizers for sustainable agriculture and environment. Cham: Springer International Publishing.
15. Kannaiyan, S. (Ed.). (2002). Biotechnology of biofertilizers. Springer Science & Business Media.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b>					
<b>FYUGP SYLLABUS</b>						
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Conservation Biology and Sustainable Development</b>					
<b>Type of Course</b>	<b>VAC</b>					
<b>Course Code</b>	<b>M24BO4VAC200</b>					
<b>Course Level</b>	<b>200</b>					
<b>Course Summary</b>	The course provides a basic overview regarding the concepts in conservation biology. It also gives a basic outlook towards the need for biodiversity conservation and sustainable development. It also creates an awareness regarding the transition to green growth.					
<b>Semester</b>	4	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		3	-	-	-	45
<b>Pre-requisites, if any</b>	Basic understanding about biodiversity and its significance					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Describe the concepts in conservation biology	U	2
2	Identify a variety of tools used by conservation biologists	U	2,4
3	Explain the concept and importance of sustainability and sustainable development	U	2
4	Examine the threats and adopt creative measures for biodiversity conservation	A	2,5
5	Assess the current status of biodiversity	E	2

***f\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)***

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>1</b>	<b>Conservation Biology (15 hours)</b>			
	1.1	Introduction to conservation Biology –Definition, career prospects in conservation biology, Conservation and management practices	3	1

	1.2	Conservation Techniques-Principles of conservation - ex-situ and in-situ conservation techniques, ecological restoration. Statistical and computational tools used in conservation biology- Population Viability Analysis (PVA), Minimum Viable Population, Decision Analysis and Multiple-Criteria Approaches	7	2
	1.3	Ecotourism-Ecotourism as a tool for conservation and sustainable development, difference between ecotourism and mainstream tourism, guidelines and green practices for ecotourism, impacts of tourism on culture and environment and its management-Examples, positive and negative impacts	5	1, 4
2	<b>Biodiversity (15 hours)</b>			
	2.1	Definition, types and importance	3	4
	2.2	Biodiversity loss- Causes, extinction, IUCN account of biodiversity, red data book, rare, endangered and threatened species (RET).	5	4,5
	2.3	Concept of endemism, Biodiversity hotspots in India.	2	4,5
	2.4	Biodiversity documentation- Case study- Students have to submit a brief report with geo-tagged photographs of the biodiversity of the nearby locality.	5	5
3	<b>Sustainable development (15 hours)</b>			
	3.1	Introduction -aim and impact of sustainable development	3	3
	3.2	Sustainable development - Basic characteristics, Core elements, Principles and Goals	5	3
	3.3	Strategies and policies for sustainable development Examples of Sustainable development in daily life –Wind energy, solar energy, sustainable forestry, bio-composting, biogas production, water efficient fixtures, green spaces and sustainable construction.	7	3
4	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning,
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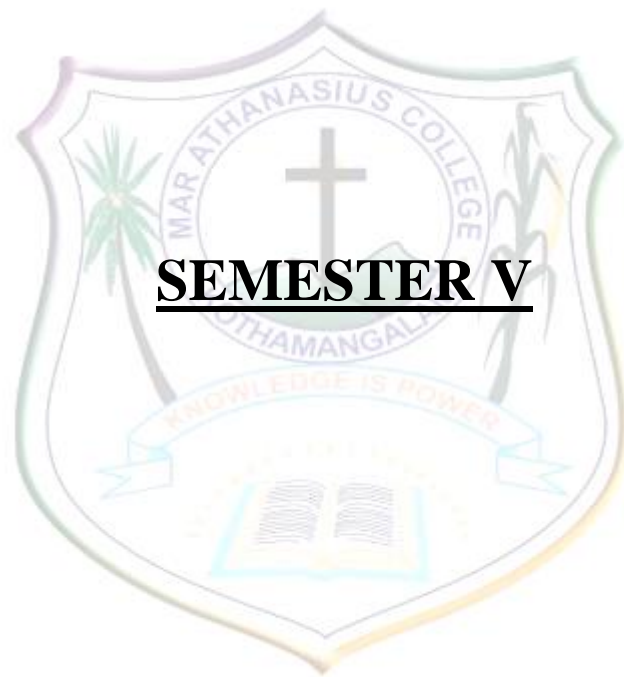
	and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar
	<b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x1 =10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## References


1. Ahmedullah M, Nayar M P (1987). Endemic plants of India
2. Elliott, Jennifer. 2012. An Introduction to Sustainable Development. 4th Ed. Routledge, London.
3. Gilpin, M. E. & Soulé, M. E. "Minimum viable populations: Processes of species extinction." In Conservation Biology: The Science of Scarcity and Diversity, ed. M. E. Soulé (Sunderland: Sinauer & Associates, 1986): 19–34.
4. Rogers, Peter P., Kazi F. Jalal, and John A. Boyd. "An introduction to sustainable development." (2012).
5. Wilson E O (1988). Biodiversity. The national academic press. 37. Wilson E O (1999). The diversity of life. W.W. Norton and Company
6. <https://asuonline.asu.edu/newsroom/online-learning-tips/what-is-conservation-biology-ecology/>
7. <https://www.nature.com/scitable/knowledge/library/conservation-biology-16089256/>
8. <https://sumas.ch/5-examples-of-sustainable-development/>

## SUGGESTED READINGS

1. IUCN (2007). The 2000 IUCN red list of threatened species. IUCN. England.
2. Jain S K, Sastry A R K (1984). The Indian plant red data book. BSI, Calcutta
3. Our Common Journey: A Transition Toward Sustainability. National Academy Press, Washington D.C. Soubbotina, T. P. 2004.
4. Primack, R. B. (1993). Essentials of Conservation Biology. Sunderland, MA: Sinauer & Associates.
5. Richard T. Wright, Dorothy F. Boorse (2017). Environmental Science: Toward A Sustainable Future, Pearson, 13th Edition



**SEMESTER V**

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Angiosperm Systematics and Economic Botany</b>				
<b>Type of Course</b>	<b>DSC</b>				
<b>Course Code</b>	<b>M24BO5DSC300</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	Angiosperm systematics focuses on systematic arrangement of flowering plants, their evolutionary relationships, and interrelations among different species. Economic botany deals with the morphological characteristics of plant parts that are economically significant, focusing on their usefulness and real-world applications.				
<b>Semester</b>	5	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		3	-	1	-
<b>Pre-requisites, if any</b>	Knowledge in general characters of angiosperms				

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Comprehend the general principles of angiosperm systematics and plant nomenclature	U	1
2	Summarize taxonomic information from available resources	U	5
3	Compare the morphological characters of plants belonging to different plant families	An	1,4
4	Execute field collections and plant specimen preparations scientifically	An	3,4
5	Utilize the knowledge in plant systematics for the benefit of	A	2,5

	science and society		
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## COURSE CONTENTS

Module	Units	Course description	Hours	CO No.
	<b>Module 1 (15 hours)</b>			
<b>1</b>	1.1	Plant Morphology Leaf morphology- Different types of leaves, Phyllotaxy Inflorescence Racemose-Simple Raceme, Spike, Catkin, Spadix, Corymb, Umbel, Head; Cymose- Simple cyme, monochasial- helicoid and scorpioid, dichasial and polychasial cymes; Special types- Cyathium, Verticillaster, Thyrsus, Hypanthodium and Panicle <b>Learning activity:</b> Collect and submit any 5 types of leaves and inflorescence mentioned in the syllabus.	4	3
	1.2	Flower – as a modified shoot Floral Whorls - arrangement, relative position- Symmetry, Aestivation, Placentation. Cohesion and adhesion of essential organs. Floral diagram and Floral Formula.	3	3
	1.3	Types of Fruits Simple fruits - Fleshy, Dry – Dehiscent, Indehiscent and Schizocarpic fruits Aggregate fruits and Multiple fruits. <b>Learning activity:</b> Collect and submit different types of fruits mentioned in the syllabus.	3	3
	1.4	Plant Taxonomy History of Plant Classification systems- Artificial System- (Linnaeus - Brief account), Natural System (B & H system- Detailed account), Phylogenetic Systems (E & P system- Brief study), APG (brief account).	3	2
	1.5	Botanical Literature- Floras- Regional and National Floras, Revision & Monographs (Brief account). Online Taxonomic Databases: International Plant Names Index (IPNI), Plants of the World Online (POWO), Botanicus.org (Brief account).	2	5

<b>Module 2 (15 Hours)</b>				
2	2.1	Herbarium technique -Steps in preparation of herbarium, Importance of Herbaria, Major Herbaria - National and International, Virtual Herbaria- Index herbariorum, Botanical Survey of India.	3	4
	2.2	Plant Nomenclature- Binomial, ICN - Introduction & Principles (Brief study), Rule of priority, Author citation, Homonym, Synonym, Basionym.	2	1
	2.4	Type concept- (Holotype, Isotype, Lectotype).	3	1
	2.5	Taxonomic keys- Bracketed and Indented keys (Brief account).	2	1
	2.6	Study the following families of Bentham and Hooker's System with special reference to their vegetative and floral characters; special attention should be given to common and economically important plants within the families Annonaceae, Malvaceae, Rutaceae, Anacardiaceae, Cucurbitaceae.	5	3
<b>Module 3 (15 hours)</b>				
3	3.1	Study the listed families from Bentham and Hooker's System, paying special attention to their vegetative and floral traits, along with noteworthy economically important plants within each family. Leguminosae (Mimosaceae, Caesalpiniaceae and Fabaceae) Apiaceae, Rubiaceae, Asteraceae, Sapotaceae,	6	3
	3.2	Apocynaceae, Solanaceae, Acanthaceae, Lamiaceae, Euphorbiaceae, Orchidaceae, Arecaceae, Poaceae.	6	3
	3.3	<b>Economic Botany</b> Study following plants with special reference to the botanical name, family, morphology of useful parts and uses - Cereals (Rice, Wheat), Millets (Ragi, Fox tail millet), Pulses (Green gram, Bengal gram), Sugar Yielding (Sugar Cane), Fruits (Banana, Guava/Mango), Vegetables (Carrot, Ladies finger), Tuber crops (Tapioca, Greater Yam), Beverages (Tea, Coffee), Oil yielding plants (Coconut, Ground nut), Fibre yielding (Coir, Cotton), Gums and resins (White dammar, Gum Arabic, Asafoetida) Insecticide yielding plants (Tobacco, Neem).	3	2



<b>4</b>	<b>Module 4 Practical (30 hours)</b>		
	Practical (30 hours) 1. Explore habitats to identify the inflorescence and fruit types mentioned in the syllabus. 2. Conduct field work for a period of not less than 3 days to familiarize plants under the guidance of faculties and submit a field report with geotagged photos. 3. Visit a recognized herbarium, practice herbarium technique and submit 15 herbarium sheets with a field book. 4. Examine vegetative and floral features of different plants and assign them to respective families mentioned in the syllabus. 5. Collect, identify and submit morphologically useful parts of any 10 plants mentioned in economic botany.	30	4
<b>5</b>	<b>Teacher specific course components</b>		

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

## REFERENCES

1. Davis, P.H. & Heywood, V.H. (1967). Principles of Angiosperm Taxonomy. Oliver and Boyd, Edinburgh.
2. Eames, A.J. (1961). Morphology of Angiosperms. McGraw Hill, New York.
3. Foster, A.S. & Giffard, E.M. (1962). Comparative morphology of vascular plants.


- Allied Pacific Pvt. Ltd. Bombay.
4. Harris, J.G & M.W. Harris (1994). Plant Identification Terminology -An illustrated Glossary, Spring lake publishing, Spring lake, Utah.
  5. Heywood, V. H. (1967). Plant Taxonomy. Edward Arnold, London.
  6. Hill, A. F. (1982). Economic Botany. McGraw Hill, New York.
  7. Jain, S.K. & Rao, R. (1976). A handbook of field and herbarium technique. Today and tomorrow, Publishers, New Delhi.
  8. Jeffery, C. (1968). An Introduction to Plant Taxonomy. J and A Churchill, London.
  9. Lawrence, G. H. M. (1951). Taxonomy of Vascular Plants. Macmillan, New York.
  10. Maheshwari, P. & Singh, U. (1965). Dictionary of Economic Plants in India. ICAR, New Delhi.
  11. Naik, V.N. (1984). Taxonomy of angiosperms. Tata Mc Graw- Hill Publishing Company, New Delhi.
  12. Narayanan, M. K. R., Shaju T, Sunil C. N., Abdussalam A. K. and AbdulJaleel V. (2013) Orchids of Wayanad. Lead Books.
  13. Pandey, S. N. & Misra, S. P. (2008). Taxonomy of Angiosperms. Ane Books India, New Delhi.
  14. Simpson, M.G. (2006). Plant Systematics. Elsevier Academic Press publications, USA.
  15. Singh, G. (2010). Plant systematics - an integrated approach (3rd Edn) Science Publishers.
  16. Singh, V. & Jain, D.K. (1989). Taxonomy of Angiosperms. Rastogi Publication, Meerut.
  17. Sivadasan, M., Anil Kumar, N., and Ravi, N. (2005). Flora of Pathanamthitta (Western Ghats, Kerala). Daya Publishing House.
  18. Sivarajan, V.V. (1991). Introduction to the Principles of Plant Taxonomy. Oxford IBH Publishing Co. Pvt. Ltd., New Delhi.
  19. Sreemali, J. L. (1979). Economic Botany. Kitab Mahal, Allahabad.
  20. Turland, N. (2013). The Code Decoded. A user's guide to the International Code of Nomenclature for algae, fungi, and plants. Koeltz Scientific Books, Königstein, Germany. 169 pp
  21. <https://www.ipni.org/>
  22. <https://powo.science.kew.org/>
  23. [www.botanicus.org](http://www.botanicus.org)
  24. <https://sweetgum.nybg.org/science/ih/>

### **SUGGESTED READINGS**

1. Beentje, H. (2016). The Kew Plant Glossary- An illustrated dictionary of plant terms (2<sup>nd</sup> Edn). Kew Publishing. Royal Botanic Garden, Kew, England.
2. Bell, A.D (1991). Plant form- An illustrated guide to Flowering plant morphology. Oxford University Press, New York, Tokyo.
3. Chase, M. W. *et al.* (2016). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. J. Linn. Soc., Bot., 181(1): 1-20.

4. Henry A. N. and Bose M. C. (2001). An aid to the International Code of Botanical Nomenclature. Botanical Survey of India, Coimbatore.
5. Prenner, G., Bateman, R. & Rudall, P. (2010). Floral formulae updated for routine inclusion in formal taxonomic descriptions. *Taxon*. 59. 241-250. 10.2307/27757066.
6. Rendle, A. B. (1979). Classification of flowering plants, Vols. I & II. Vikas Publishing House, U.P.
7. Sambamurthy, A. (2005). Taxonomy of Angiosperms. I.K. International Pvt. Ltd, New Delhi.
8. Sharma, O. P. (1996). Plant Taxonomy. Tata McGraw Hill, New Delhi.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Plant Physiology and Biochemistry</b>				
<b>Type of Course</b>	<b>DSC</b>				
<b>Course Code</b>	<b>M24BO5DSC301</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	The course aims at introducing the physiology of plant systems and indulges the student in finding out various processes that function within the plant body. The course also deals with various biomolecules.				
<b>Semester</b>	5	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		3	-	1	-
<b>Pre-requisites, if any</b>	Concept of a plant cell and cell components, Basic chemistry of compounds				

#### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No.</b>
1	Describe various biomolecules in the living system.	U	3
2	Summarize the physiology of different plant life processes.	U	3,4
3	Categorize the factors affecting physiological processes	An	3,4
4	Evaluate the role of biotic and abiotic components in plant stress	E	5
5	Examine intricacies of protein structure and diversity	A	3

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Plant water relations: Diffusion, imbibition, osmosis, water potential; Absorption of water - active and passive, apoplastic and symplastic pathways. Role of aquaporins.	3	2
	1.2	Ascent of sap: Cohesion-tension theory. Transpiration – types and significance; anti-transpirants. Guttation.	3	2,3
	1.3	Nutrient elements: Classification based on biochemical functions and Physiological roles. Beneficial elements. Mineral uptake - passive (ion exchange) and active (carrier concept).	3	2
	1.4	Carbohydrates: Classification: mono (glucose and fructose), di (sucrose) and polysaccharides (starch); general structure (Haworth Projection) and functions. Lipids: General features, roles and types of lipids (Simple and Compound, structural and storage lipids).	6	1
2	<b>Module 2 (15 Hours)</b>			
	2.1	Proteins: General account of proteins - amino acid, peptide bond. Structural levels of proteins - primary, secondary, tertiary, and quaternary; General functions of proteins Enzymes: classification and nomenclature, mechanism of action (Lock and Key Hypothesis, Induced fit theory). Enzyme inhibition and Factors affecting enzyme action.	5	1,5
	2.2	Photosynthesis: Pigments, Photosystems; Light Reactions - cyclic and non-cyclic photophosphorylation. Dark reactions - C3, C2, C4 pathway, CAM. Factors affecting photosynthesis.	8	2,3
	2.3	Translocation of solutes: Phloem loading and unloading, polymer trapping (brief account)	2	2

3	<b>Module 3 (15 Hours)</b>			
	3.1	Respiration: Anaerobic and Aerobic; Glycolysis, Kreb's cycle, Mitochondrial Electron Transport system, ATP synthesis - chemi-osmotic hypothesis, Respiratory quotient; Factors affecting respiration.	9	2,3
	3.2	Plant hormones: Physiological effect and practical applications - Auxins, Gibberellins, Cytokinins, ABA, and Ethylene.	2	2
	3.3	Stress Physiology: Abiotic (drought, salinity and heavy-metal), Biotic (pathogen) stress, Plant stress response (brief study).	2	2,4
	3.4	Physiology of flowering: Photoperiodism, Vernalization	2	2
4	<b>Practical (30 Hours)</b>			
	4.1	<p><b>Plant Physiology (20 Hours)</b> Core Experiments (any 3):</p> <ul style="list-style-type: none"> <li>● Preparation of molal, molar, normal, and percentage solutions and their dilutions</li> <li>● Separation of plant pigments by TLC/Paper/Column chromatography.</li> <li>● Estimation of plant pigments by colorimetry.</li> <li>● Estimation of Proline in plant tissue under abiotic stress.</li> <li>● Estimation of Phenol in plant tissues under biotic stress.</li> <li>● Calculation of stomatal index in mesophytes and xerophytes</li> <li>● Estimation of rate of photosynthesis</li> <li>● Measurement of growth using various parameters</li> </ul> <p>Demonstration experiments: (ANY 4)</p> <ul style="list-style-type: none"> <li>● Demonstration of plasmolysis.</li> <li>● Demonstration of osmosis using osmoscope.</li> <li>● Demonstration of Oxygen evolution during Photosynthesis.</li> <li>● Measurement of transpiration rate using Ganong's potometer/Farmer's potometer</li> <li>● Mohl's half leaf experiment.</li> </ul>	20	1,2,3

	4.2	<b>Biochemistry (10 Hours)</b> <ul style="list-style-type: none"> <li>● General test for carbohydrates – Molisch’s test, Benedict’s tests / Fehling’s test.</li> <li>● Colour test for starch - Iodine test.</li> <li>● Colour tests for proteins in solution – Biuret test/Million’s test</li> <li>● Quantitative estimation of protein using a colorimeter.</li> </ul>		
	4.3	<b>Activity</b> (Any one) <ul style="list-style-type: none"> <li>● Design and perform an experiment related to plant physiology. Prepare and submit a report with geotagged photos.</li> <li>● Prepare and submit a report with your views and conclusions on the latest research in physiology / biochemistry based on journal publications on any topic mentioned in the syllabus (A copy of the original publication has to be submitted with the report.</li> <li>● Design models representing physiological or biochemical processes taking place in plants and submit them for evaluation.</li> <li>● Prepare a review article in a selected research area in Physiology and biochemistry and submit for evaluation.</li> <li>● Retrieve 5 research articles on any selected topic in Physiology/ biochemistry and submit them for evaluation.</li> </ul>	5+5	2,3
5	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. End Semester Examination</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x1 =10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks

Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks
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## REFERENCES

1. Dayananda B, 1999. Experiments in Plant Physiology. Narosa Publishing House, New Delhi.
2. Hopkins W G, Norman P A Huner, 2008. Introduction to plant physiology. John Wiley and sons. New York.
3. Jain J L, Sanjay Jain, Nitin Jain, 2005. Fundamentals of Biochemistry. S Chand, New Delhi.
4. Lehninger A L, 1961. Biochemistry. Lalyan publishers, Ludhiana.
5. Nelson D L, Cox M M, 1993. Principles of Biochemistry. MacMillan Publications.
6. Pandey S N, Sinha B K, 2006. Plant Physiology. Vikas Publishing House Pvt. Ltd.
7. Plummer D T, 1988. An introduction to practical biochemistry. Tata McGraw-Hill Publishing Company, New Delhi.
8. Sadasivam S, Manickan A, 1996. Biochemical Methods. New Age International Ltd. New Delhi.
9. Salisbury F B, Ross C W, 1992. Plant Physiology. CBS Publishers and Distributers, Delhi.
10. Srivastava H S, 2005. Plant Physiology. Rastogi publications, Meerut.
11. Verma V, 2007. Textbook of Plant Physiology. Ane Books India, New Delhi.
12. Taiz L, Zeiger E, Moller I, Murphy A 2023. Plant Physiology and Development (VII Edn). Oxford University Press



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Plant Biotechnology and Introduction to Bioinformatics</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO5DSE300</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	The course is designed as a comprehensive exploration to the field of Plant Biotechnology and Bioinformatics. The course aims to familiarize students with the key developments in the sphere of Plant Biotechnology and to discuss the potential applications of biotechnology in crop improvement and for novel uses for plants.				
<b>Semester</b>	5	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		4	-	-	-
<b>Pre-requisites, if any</b>	General overview and key concepts of Biotechnology				

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Understand the process of rDNA technology.	U	3, 5
2	Develop knowledge in fundamental gene cloning techniques.	A	3
3	Compare different gene transfer methods based on efficiency and specificity.	An	3
4	Explain the applications of plant genetic engineering in the field of agriculture, medicine, environment, and industry.	An	3,5

5	Utilize bioinformatics tools and databases.	A	4,5
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>Module 1 (15 Hours)</b>				
<b>1</b>	1.1	Introduction to rDNA technology: Steps in recombinant DNA technology Restriction Endonucleases (Types I-IV, biological role and application); T4 DNA Ligase. Cloning Vectors: properties of ideal cloning vector, features of cloning vectors: Plasmids, Cosmids, Phage vectors and artificial chromosomes. pCAMBIA, Ti plasmid, Lambda phage, YAC, BAC, Expression vectors, Shuttle vector- Brief account only.	8	1, 2
	1.2	Methods of gene transfer: direct gene transfer - electroporation, microinjection, microprojectile /particle bombardment, In-direct gene transfer- Agrobacterium mediated gene transfer.	4	1, 3
	1.3	Selection of recombinant clones: screening of recombinant DNA- Antibiotic resistance markers, complementation (Blue white screening). Reporter genes (GUS, GFP).	3	1, 2
<b>Module 2 (15 Hours)</b>				
<b>2</b>	2.1	Herbicide resistant plants (RoundUp Ready soybean); transgenic crops with improved quality traits (Golden rice); improved horticultural varieties (Moondust carnations)	3	4
	2.2	Role of transgenics in bioremediation (Superbug); Industrial enzymes (Aspergillase, Protease, Lipase); Genetically Engineered Products–Edible vaccine.	2	4

	2.3	PCR and its applications, Agarose gel electrophoresis and UV transilluminator Biotechnology instrumentation and Lab visit	10	1,2
3	<b>Module 3 ( 15 Hours)</b>			
	3.1	Gene editing tools (CRISPR- Cas9) and its role in transgenic plant development and gene function studies (Brief account only) Synthetic biology and plant metabolic engineering for improved crop traits, Developing climate resilient crops (Brief account only)	10	2,4,5
	3.2	Ethical considerations in plant biotechnology Biosafety considerations and IPR associated with GM crops	5	4,5
4	<b>Module 4 (15 HRS)</b>			
	4.1	An introduction to bioinformatics, objectives and applications of bioinformatics. Biological data bases: types - primary, secondary and composite databases; nucleotide sequence databases – NCBI (GenBank), EMBL, DDBJ; Protein Sequence databases - SWISS-PROT/UNIPROT, PIR; Protein structure database – PDB; Bibliographic database – PubMed, Chemical Structure database:- Pubchem.	8	5
	4.2	Sequence analysis and molecular phylogeny Sequence analysis tools - BLAST and FASTA, Molecular visualisation tools - Pymol /Rasmol . Sequence alignment - Scoring matrices, global and local alignment, Pairwise and multiple sequence alignment; common software used in alignment - MEGA Molecular phylogeny - homologs, orthologs and paralogs; phylogenetic tree - rooted and unrooted tree, advantages of phylogenetic tree, use of MEGA/IQ-TREE	7	5
5	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b>  Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b>  <b>A. Continuous Comprehensive Assessment (CCA)</b>  Theory Total = 30 marks  Quiz, Test Papers, seminar</p>
	<p><b>B. End Semester examination</b>  Theory Total = 70 marks, Duration 2 hrs  Part A (Short answer) – 10 out of 12 x 2 = 20 marks  Part B (Short essay) – 6 out of 9 x 5 = 30 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>

## References

1. Acquaah, G. (2005). *Understanding biotechnology*. Pearson.
2. Brown, T. A (2002). *Genomes* (II Edn). Bios.
3. Chawla H. S (2009): *Introduction to Plant Biotechnology* 3<sup>rd</sup> Edition, Oxford & IBH Publishing Co. Pvt. Ltd. New Delhi.
4. DeLisi, C. (2019). The role of synthetic biology in climate change mitigation. *Biol Direct* **14**, 14
5. Desmond S. T Nicholl (2008): *An Introduction to Genetic Engineering; Studies in Biology*. Cambridge University Press. 3<sup>rd</sup> Edition.
6. Ignasimuthu, S (2009). *Basic Bioinformatics*. Narosa Publications.
7. Keshavachandran R and Peter K V (2008): *Plant Biotechnology: Methods in Tissue Culture and Gene Transfer*. Orient Blackswan.
8. Liu, D., Hu, R., Palla, K. J., Tuskan, G. A., & Yang, X. (2016). Advances and perspectives on the use of CRISPR/Cas9 systems in plant genomics research. *Current Opinion in Plant Biology*, *30*, 70-77.
9. Liu, W., & Stewart, C. N. (2015). Plant synthetic biology. *Trends in Plant Science*, *20*(5), 309-317.
10. Paul G Higgs, Teresa K Attwood (2005). *Bioinformatics and molecular evolution*. Blackwell publishing.
11. Razdan M K. (2015): *Introduction to Plant Tissue Culture*. Oxford and IBH, MKM Publishers 2<sup>nd</sup>. Edition
12. Razzaq, A., Saleem, F., Kanwal, M., Mustafa, G., Yousaf, S., Imran Arshad, H. M., & Joyia, F. A. (2019). Modern trends in plant genome editing: an inclusive

- review of the CRISPR/Cas9 toolbox. *International Journal of Molecular Sciences*, 20(16), 4045.
13. Sensen, C. W. (2002). *Genomics and Bioinformatics*. Wiley – VCH.
  14. Smita Rastogi and Neelam Pathak (2009). *Genetic Engineering*. Oxford University Press. New Delhi.
  15. Timir Baran Jha and Biswajit Ghosh (2016): *Plant Tissue Culture*. Platinum Publishers. Revised 2<sup>nd</sup> Edition. Kolkata
  16. William J Thieman, Michael A Palladino (2009). *Introduction to biotechnology* (II Edn). Pearson.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Plant Tissue Culture</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO5DSE301</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	The course aims to offer a comprehensive understanding of the principles, techniques, and types of tissue culture, along with its applications and benefits.				
<b>Semester</b>	5	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		4	-	-	-
<b>Pre-requisites, if any</b>	Knowledge in plant cell biology, physiology and biochemistry				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Understand the basic principles and techniques of plant tissue culture	U	3
2	Explain somatic embryogenesis and its applications	U	3, 5
3	Analyze various techniques of germplasm conservation and its significance.	An	3, 4
4	Interpret the somaclonal and ploidy variations	U	5
5	Compare the types of tissue culture, their significance, and analyze secondary metabolite production from various cultures.	E	5

**\*Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Plant Tissue Culture: Historical perspective Basic infrastructure of tissue culture lab Differentiation, Dedifferentiation and redifferentiation, Totipotency; Organogenesis (direct and indirect); Embryogenesis. Methodology - Sterilization (physical and chemical methods), Culture media - solid and liquid media, Gelling agents, Murashige and Skoog's (MS medium), Nutrient and hormone requirements (role of vitamins and hormones). Preparation of explants and inoculation, callus induction, subculturing, hardening.	6	1
	1.2	Somatic embryogenesis: General aspects, initiation of embryogenic cultures, maturation of somatic embryos, regeneration of plants, factors regulating somatic embryogenesis. Differences between somatic and zygotic embryos. Encapsulation of somatic embryos, synthetic seed production; desiccated and hydrated types. Applications and limitations of synthetic seeds.	6	2
	1.3	Origin of somaclonal variation – pre-existing variability, in vitro induced variability.	3	4
2	<b>Module 2 (15 hours)</b>			
	2.1	Reasons for somaclonal variation– changes in ploidy level, changes in chromosome structure, gene mutations, gene amplifications, changes in extra nuclear genes, activation of transposable elements, DNA methylation.	7	4
	2.2	Isolation of somaclonal variants. Applications of somaclonal variation.	2	4

	2.3	Protoplast culture: Isolation and purification of protoplasts, culture of protoplasts, cell division and callus formation, plant regeneration.	3	1,5
	2.4	Protoplast fusion (somatic hybridization) – chemical, mechanical, electrofusion. Selection and isolation of heterokaryons, cybrids and their applications. Applications of protoplast culture.	3	4,5
	<b>Module 3 (15 hours)</b>			
3	3.1	Anther culture – Androgenesis, pretreatment of anther/pollen grains, media and growth regulators, Induction and stage of pollen development, regeneration, androgenic embryos, factors affecting androgenesis. Microspore culture - protocol, advantages over anther culture. Ovary and ovule culture - Gynogenesis: Developmental stage at inoculation, in vitro maturation of embryo sacs, triggering factors – pretreatment, medium. Uses and limitations of haploid plants.	6	4,5
	3.2	Endosperm culture - Conventional production of triploid plants, Triploid plants and their importance, Advantages and limitations of endosperm culture. Meristem culture - methods, advantages and applications. Embryo culture, embryo rescue techniques and its applications.	5	4,5
	3.4	Production of secondary metabolites Culture conditions for producing secondary metabolites, selection of high yielding lines, elicitation, immobilization of cells. Hairy root culture – advantages of using hairy root culture, establishment of hairy root culture and production of secondary metabolites.	4	5
	<b>Module 4 (15 hours)</b>			
4	4.1	Germplasm conservation Importance, methods of conservation <i>In vitro</i> conservation, short and medium term storage. Cryopreservation technique – importance of cryopreservation, pretreatment, freezing methods, cryoprotectants, vitrification.	6	3



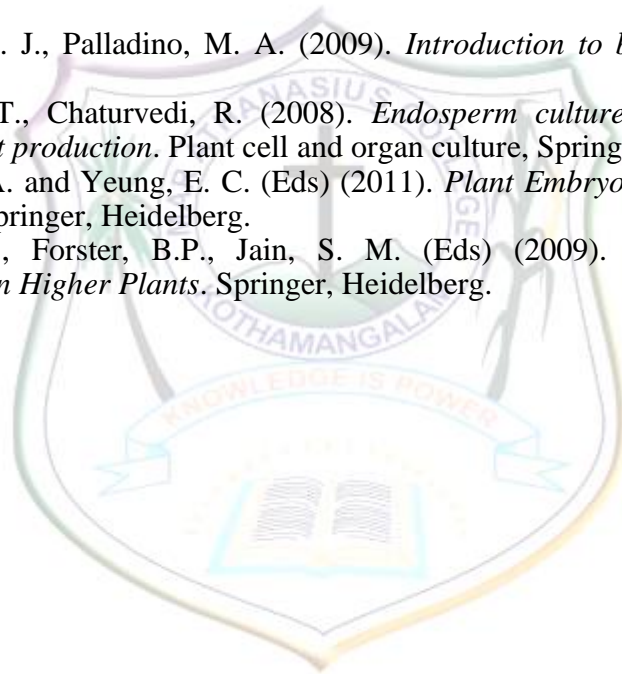
	4.2	Tissue culture and its economic prospects. Micropropagation of orchids, forest plants and horticultural plants.	4	5
	4.3	<u>Activity:</u> Sterilization techniques – Surface sterilization, Hot air oven, autoclave, Laminar Air Flow, Filtration Media preparation Inoculation Immobilization of whole cells/tissues in sodium alginate Isolation of anther for anther culture Visit to a reputed biotechnology/Tissue culture laboratory.	5	1
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## References

1. Arditti, J. (2008). *Micropropagation of Orchids* (Vol. I). Blackwell publishing.
2. Barnum, S. R. (1998). *Biotechnology an introduction*. Thomson Brooks/cole.
3. Benson, E. E. (1999). *Plant Conservation Biotechnology*. Taylor and Francis.
4. Bhojwani, S. S., Razdan, M. K. (1996). *Plant tissue culture: Theory and Practice*. Elsevier.
5. Collin, H.A., Edwards, S (1998). *Plant tissue culture*. Bios scientific publishers.
6. Davey, M. R., Anthony, P. (2010). *Plant Cell Culture: Essential Methods*. Wiley-Blackwell A John Wiley & Sons, Ltd.
7. Dixon, R. A., Gonzales, R. A. (2004). *Plant cell culture, a practical approach* (II Edn). Oxford University Press.

8. Edwin F. George, Michael A. Hall, Geert-Jan De Klerk (2008). *Plant Propagation by Tissue Culture (Vol I): The Background*. Springer.
9. Evans, D. E., Coleman, J. O. D., Kearns, A. (2003). *Plant Cell Culture*. BIOS Scientific Publishers.
10. Gamborg, O.L., Philips, G. C. (Eds.) (2005). *Plant cell, tissue and organ culture: Fundamental methods*. Narosa Publishinh House.
11. Hvoslef-Fide, A. K., Preil, W. (Eds) (2005). *Liquid Culture Systems for in vitro Plant Propagation*. Springer.
12. Ignacimuthu, S. (2006). *Biotechnology: An introduction*. Narosa Publishing House.
13. Jain, S. M., Häggman, H. (Eds) (2007). *Protocols for Micropropagation of Woody Trees and Fruits*. Springer, Heidelberg.
14. Pritchard, H. W. (2004). *Modern methods in orchid conservation: The role of Physiology, Ecology and Management*. Cambridge University Press.
15. Ratledge, C. and Kristianson, B. (2001). *Basic biotechnology*. Cambridge University press.
16. Reed, B.M. (2008). *Plant Cryopreservation: A Practical Guide*. Springer, Heidelberg.
17. Thieman, W. J., Palladino, M. A. (2009). *Introduction to biotechnology (II Edn)*. Pearson.
18. Thomas, D.T., Chaturvedi, R. (2008). *Endosperm culture: A novel method for triploid plant production*. Plant cell and organ culture, Springer science.
19. Thorpe, T. A. and Yeung, E. C. (Eds) (2011). *Plant Embryo Culture: Methods and Protocols*. Springer, Heidelberg.
20. Touraev, A., Forster, B.P., Jain, S. M. (Eds) (2009). *Advances in Haploid Production in Higher Plants*. Springer, Heidelberg.





**Mar Athanasius College (Autonomous), Kothamangalam**

**FYUGP SYLLABUS**

<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Bioactive Phytochemicals</b>					
<b>Type of Course</b>	<b>DSE</b>					
<b>Course Code</b>	<b>M24BO5DSE302</b>					
<b>Course Level</b>	<b>300</b>					
<b>Course Summary</b>	This course will provide an understanding about the diversity of bioactive compounds present in plants. Students will also get an insight into the pathways of biosynthesis of these compounds and their significance.					
<b>Semester</b>	5	<b>Credits</b>				<b>Total Hours</b>
<b>Course Details</b>	<b>Learning Approach</b>	<b>Lecture</b>	<b>Tutorial</b>	<b>Practical</b>	<b>Others</b>	
		4	-	-	-	60
<b>Pre-requisites, if any</b>	Knowledge in physiology and biochemistry of plants					

**COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Understand the diversity of bioactive phytochemical compounds in plants	U	3
2	Describe the structure and functions of phytochemicals in plants	U	3
3	Compare the metabolic pathways leading to the biosynthesis of phytochemicals	An	3
4	Evaluate the significance of bioactive phytochemicals in various fields	E	3,5
5	Plan analytical tests to identify the presence of phytochemicals in plant extracts	An	4

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

**COURSE CONTENT**

<b>Module</b>	<b>Units</b>	<b>Course description</b>	<b>Hours</b>	<b>CO No.</b>
<b>1</b>	<b>Module 1 (15 Hrs)</b>			

	1.1	Introduction: Plants as sources of bioactive compounds, historical perspectives. Common plant families with active principles, parts, Examples of plants with bioactive compounds.	4	1
	1.2	Active principles in plants, chemical diversity of bioactive compounds, secondary metabolites	3	1
	1.3	Primary metabolites vs Secondary metabolites – interconnected pathways of synthesis	2	3
	1.4	Brief study of biosynthetic pathways of Alkaloids, Terpenoids and Phenolics	6	3
	<b>Module 2 (15 Hrs)</b>			
2	2.1	Alkaloids – introduction, properties, occurrence, structure, classification, functions	3	2
	2.2	Terpenoids – introduction, properties, occurrence, structure, classification, functions	3	2
	2.3	Phenolics – introduction, properties, occurrence, structure, classification, functions	3	2
	2.4	Flavonoids – introduction, properties, occurrence, structure, classification, functions	3	2
	2.5	Coumarins – introduction, properties, occurrence, structure, classification, functions	3	2
	<b>Module 3 (15 Hrs)</b>			
3	3.1	Pharmacological uses of plant bioactive compounds	3	4
	3.2	Phytochemicals in cosmetics and Food industry	3	4
	3.3	Ecophysiological significance of phytochemicals	3	4
	3.4	Role of Secondary metabolites in plant taxonomy	3	4
	3.5	Biotechnological approaches to produce bioactive compounds	3	4
	<b>Module 4 (15 Hrs)</b>			
4	4.1	Extraction of active principles from plants: methods – hot and cold extraction, Soxhlet extraction, Clevenger; Common solvents used for extraction	5	5
	4.2	Qualitative analysis of secondary metabolites – methods for preliminary analysis	4	5
	4.3	<b>Activity</b> Extraction of bioactive components from plant samples: Hot and cold extraction, Soxhlet extraction	6	5

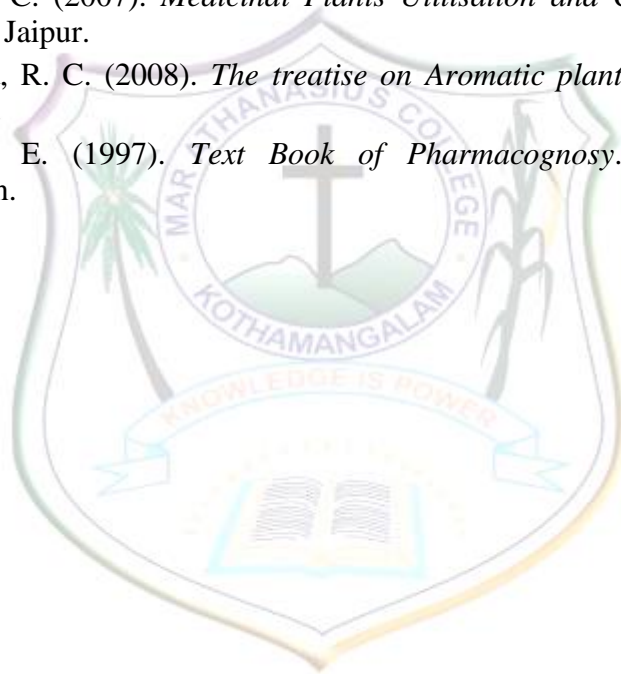
		Primary screening of bioactive compounds Extraction and estimation of selected secondary metabolites		
5		<b>Teacher specific course components</b>		


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

#### References:

- Asolkar, L. V., Kakkar, K. K., & Chakre, O. J. (1992). Glossary of Indian medicinal plants with active principles. *CSIR, New Delhi, 1*, 187.
- Buchanan, B. B., Gruissem, W., & Jones, R. L. (Eds.). (2015). *Biochemistry and molecular biology of plants*. John Wiley & Sons.
- Daniel M, (1991). *Methods in plant chemistry and Economic Botany*. Kalyani publishers, New Delhi.
- Egbuna, C., Ifemeje, J. C., Udedi, S. C., & Kumar, S. (Eds.). (2018). *Phytochemistry: Volume 1: Fundamentals, Modern Techniques, and Applications*. CRC Press.
- Indian Medicinal Plants (5Vols)* (1994). Arya Vaidya Sala Kottackal, Orient longoman New Delhi.
- Irfan Ali Khan, (2008). *Medicinal and Aromatic plants of India*, Ukaaz Publishers, Hyderabad.
- Jain S. K. (2004). *A Manual of Ethnobotany*, Scientific Publishers, India.
- Kapoor, L. D. (2018). *CRC handbook of Ayurvedic medicinal plants*. CRC press.
- Krishnaswamy N, R. (2003). *Chemistry of Natural Products*, Universities press, Hyderabad.
- Matsumura, E., Matsuda, M., Sato, F., Minami, H., Ramawat, K. G., & Mérillon, J. M. (2013). Natural products: phytochemistry, botany and metabolism of alkaloids, phenolics and terpenes.

11. Murthy, H. N., Paek, K. Y., & Park, S. Y. (2024). Bioactive Compounds in the Storage Organs of Plants.
12. Pati, S., Sarkar, T., & Lahiri, D. (Eds.). (2023). *Recent Frontiers of Phytochemicals*. Elsevier.
13. Pushpangaden, P., Nyman, U. L. F., & George, V. (1996). *Glimpses of Indian Ethno Pharmacology*. The Royal Danish School of Pharmacy Copenhagen, Denmark.
14. Raaman, N. (2006). *Phytochemical techniques*. New India Publishing.
15. Sadasivam, S. & Manickam, A. (1996). *Biochemical methods* (II Edn). New age international Publishers.
16. Shah, B. N. (2009). *Textbook of pharmacognosy and phytochemistry*. Elsevier India.
17. Evans, W. C. (2009). *Trease and Evans' pharmacognosy*. Elsevier Health Sciences.
18. Trivedi, P. C. (2007). *Medicinal Plants Utilisation and Conservation*, Avishkar Publishers, Jaipur.
19. Upadhyaya, R. C. (2008). *The treatise on Aromatic plants*. Anmol Publications, New Delhi.
20. Wallis, T. E. (1997). *Text Book of Pharmacognosy*. CBS Publication & Distribution.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Food science and quality control</b>					
<b>Type of Course</b>	<b>DSE</b>					
<b>Course Code</b>	<b>M24BO5DSE303</b>					
<b>Course Level</b>	<b>300</b>					
<b>Course Summary</b>	In this course, students will be familiarized with the components of food and the changes leading to spoilage. They acquire an in-depth understanding of the technologies used to produce safe and nutritious foods as well as the importance of food security. The course will also provide information about the regulations to be followed in food industries and food-related sectors.					
<b>Semester</b>	5		Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		4	-	-	-	60
<b>Pre-requisites, if any</b>	Basic understanding of the structure of carbohydrates, proteins and fats as components of food					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Describe the food components and issues relevant to food processing and food quality management systems.	U	3
2	Explain food adulteration, food borne diseases and methods to identify food adulteration	U	3,5
3	Discuss the spoilage and deterioration mechanisms in foods and methods to control spoilage.	U	4,5
4	Evaluate the principles of food science to assure the quality of food products.	E	2
5	Employ the principles of food science in practical, real-world situations and problems.	A	2,5

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Composition and Types of food (14 hours)</b>			
	1.1	Introduction and scope of Food science Composition of food: <ul style="list-style-type: none"> <li>● Carbohydrates- Major sources and functions.</li> <li>● Proteins-Major sources and functions.</li> <li>● Lipids-Saturated and unsaturated fatty acids, Dietary sources, functions of fats.</li> </ul> Fiber – Dietary sources, functions	5	1
	1.2	Minerals- Calcium, Phosphorus, Magnesium, Sodium, Potassium etc Vitamins- fat soluble and water soluble	2	1
	1.3	Enzymes- Amylase, Protease, Lipase, Phytase, Lipoxygenase, Pectic enzyme Pigments-Chlorophylls, Carotenoids	3	1
	1.4	Types of food- Nutraceuticals, Probiotics, Prebiotics, GM food, Organic food, Traditional food, Fermented food	4	1
2	<b>Food additives, Food adulteration and Food borne diseases (15 hours)</b>			
	2.1	Food additives: Food colours, Sweeteners, Gelling agents, Flavour enhancers, Surface acting agents, Bleaching agents, Stabilizers, and Thickeners Activity: Carry out a market survey of additives used in different types of foods, classify them based on their role and present your findings as PowerPoint presentations.	5	1,2
	2.2	Food adulteration: Definition, Common adulterants in food, Reasons for adulteration	1	2
	2.3	Testing adulteration in milk, ghee, sugar, salt, tea, coffee, chili powder, turmeric powder, sweets, poultry and fish (Brief account) Hands on training on Adulteration testing of milk, chilli powder and tea (market sample)	9	2
	2.4	Harmful effects of food adulteration	1	2
	2.5	Food borne illness and diseases associated: Food poisoning, Botulism, Ergotism, Staphylococcal intoxication, Mycotoxicosis	3	1,2
3	<b>Food spoilage and preservation (14 hours)</b>			
	3.1	Food spoilage: reasons for food spoilage, Physical and Chemical changes in food that affect texture, flavour, odour, stability and nutritive value during processing and storage.	2	3
	3.2	Food preservation methods: asepsis, removal of microorganisms, Drying, smoking, low temperature, high temperature, Canning, vacuum filling, UV radiation Activity: Familiarize with different preservation	8	3,4




		methods employed for preservation of vegetables, fruits, cereals, and pulses- Submission of report		
	3.3	Food Preservatives: Salt, Vinegar, Sugar, Benzoates, Sorbates, Nitrates, Propionates, Antioxidants, Antibiotics, Antifungal preservatives	4	4
<b>4</b>	<b>Quality control in Food industry (13 hours)</b>			
	4.1	Quality control (QC) in food industry, major concepts of QC, Significance	3	1,3,5
	4.2	Food safety Standards and Regulations-ISO 22000, HACCP, FSSAI, GMP, AGMARK Visit any Food industry/Food processing unit that follows food safety standards and regulations and submit a report	7	1,3,5
	4.3	Quality control activities-Sampling and Inspection, Certification, Testing laboratories	3	1,3,5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

### References

1. Adams, M.R. & Mass, M.D. (2008). Food Microbiology, New age international Pvt Ltd.
2. Banwart, G.T. (2017). Basic Food Microbiology, 2<sup>nd</sup> edition, CBS Publications, New Delhi ISBN-13 978-8123906461
3. Black, J.G. (1999). Microbiology Principles and exploration, 4<sup>th</sup> Edition. John Wiley and sons Inc. ISBN-13 978-0471377320
4. Frazier, W.C. (1989). Food Microbiology. 4<sup>th</sup> Edition, McGraw Hill Education. ISBN-13 978-0071004367

5. Grumezescu, A. M., & Holban, A. M. (2017). *Microbial Contamination and Food Degradation*. Academic Press.
6. Jay, J. M. (2005) *Modern Food Microbiology*. Fourth edition, CBS Publishers, and Distributors Pvt. Ltd. ISBN-13 978-8123904757
7. Jay, J.M., Lossner, M.J.& Golden, D.A. (2008). *Modern food Microbiology*. 7<sup>th</sup> edition, Springer, ISBN0387231803.
8. Jha, S. N. (2015). *Rapid detection of food adulterants and contaminants- Theory and practice* Academic Press (Elsevier). ISBN 978-0-12-420084-5
9. Lara, W (2019). *Food science and quality control* ED Tech press, ISBN-978-1-83947-260-2
10. Liu, D. (Ed.) (2018). *Handbook of Foodborne diseases*. CRC Press
11. Pelczar, J.M., Chan, E. C.S., & Kreig, N.R. (2023). *Microbiology* 5<sup>th</sup> Edition, Affiliated East West Press ISBN-13 978-8176711234
12. Damodaran, S., Parkin, K.L. & Fennema, D.R. (Eds.) (2007). *Fennema's Food Chemistry*, 4<sup>th</sup> Edition, CRC press.
13. Guthrie, H.A. (1983). *Introductory nutrition*. 5<sup>th</sup> edition. Mosby, 5<sup>th</sup> Edition, St. louis, ISBN-13 978-0801619977
14. Meyer, L.H. (2004). *Food Chemistry*, Textbook publishers. ISBN: 0758149204
15. Mudambi, S.R., Rao, S.M.& Rajagopal, M.V. (2006). *Food Science*. 2<sup>nd</sup> edition, *New Age International*. Pvt. Ltd. *Publishers*
16. Mudambi, S.R. & Rajagopal, M.V. (2001). *Fundamentals of Food and Nutrition*, 4<sup>th</sup> edition. New Age International Publishers
17. Shakuntla, M.N.& Shadaksharaswamy, M. (2013). *Food facts and principles*. New Age International
18. Srilakshmi, B. (2003). *Food science*, 3<sup>rd</sup> edition. New Age International. ISBN-13 978-8122414813
19. Swaminathan, M. (2022) *Advanced text book on Food and nutrition*, Vol II, The Bangalore Press.
20. Swaminathan, M. (2018) *Handbook of Food and Nutrition*, 5<sup>th</sup> Edition, The Bangalore Press.
21. Swaroop, A., Bagchi, D., & Preuss, H.G. (2015). *Nutraceuticals and Functional Foods in Human Health and Disease Prevention* CRC Press, ISBN- 9781482237221
22. Watson, R. R., & Victor R Preedy, V.R. (Eds.) (2015). *Probiotics, Prebiotics, and Synbiotics: Bioactive Foods in Health Promotion* Academic press (Elsevier) ISBN- 978-0-12-802189-7

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Mushroom Production and Value Addition</b>					
<b>Type of Course</b>	<b>SEC</b>					
<b>Course Code</b>	<b>M24BO5SEC300</b>					
<b>Course Level</b>	<b>300</b>					
<b>Course Summary</b>	The present course encompasses various aspects of mushrooms focusing on its importance as a valuable food supplement. The course also deals with various aspects of mushroom cultivation including the process, requirements and post-harvest steps. The value addition and marketing strategies connected to this field is also included.					
<b>Semester</b>	5	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		3	-	-		-
<b>Pre-requisites, if any</b>	General awareness in science					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Explain the importance of mushrooms and distinguish between edible and poisonous mushrooms	U	1
2	Appreciate the nutritive value and health benefits of mushrooms and implement edible mushroom cultivation techniques	A	2, 3
3	Analyze the possibilities of value addition in mushrooms	An	4, 5
4	Develop entrepreneurship skills through product design	S	5
5	Create marketing strategies for value-added products	C	5

	of mushrooms		
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>Module 1 (15 hours)</b>				
<b>1</b>	1.1	General characters and morphology of mushrooms. Distinguishing characters of button, oyster and milky mushrooms.	3	1
	1.2	Identification of mushrooms - edible and poisonous. Scope and significance of mushroom cultivation	3	1
	1.3	Nutritional profile of mushrooms- Carbohydrates, proteins, amino acids, vitamins, minerals, fats and fibre.	2	2
	1.4	Health benefits of Mushrooms-anti-tumour, antiviral and antibacterial effect, in therapeutic diet (brief study)	2	2
<b>Module 2 (15 hours)</b>				
<b>2</b>	2.1	Mushroom cultivation: Requirements, structure and construction of mushroom house, Sanitation and sterilization	2	2, 4
	2.2	Spawn preparation- requirements, spawn substrate selection, isolation of pure culture and nutrient media for pure culture, maintenance and storage of spawn. <b>Learning activity:</b> Demonstration of mushroom bed preparation/spawn preparation	5	2, 4
	2.3	Cultivation of Milky Mushroom ( <i>Calocybe indica</i> ), and Oyster Mushroom ( <i>Pleurotus</i> spp.) using paddy straw. <b>Learning activity:</b> Training in Oyster mushroom cultivation	5	2,4
	2.4	Pest and disease management in mushroom cultivation (brief account), Spent mushroom substrate utilization-fodder, compost.	3	1, 2
<b>3</b>	<b>Module 3 (15 hours)</b>			

	3.1	Post-harvest processing of mushrooms-refrigeration / instant packing, freeze drying, dehydration, canning	2	3, 4
	3.2	Value-added products from mushrooms – soup powder, biscuits, chutney powder, pickles.	2	3, 4, 5
	3.3	Marketing strategies for mushroom products	1	4, 5
	3.4	Major problems in mushroom cultivation and solutions. self-employment schemes, Government aids <b>Learning activity:</b> Expert interaction and live bed preparation	10	4, 5
<b>4</b>	<b>Teacher-specific course components</b>			

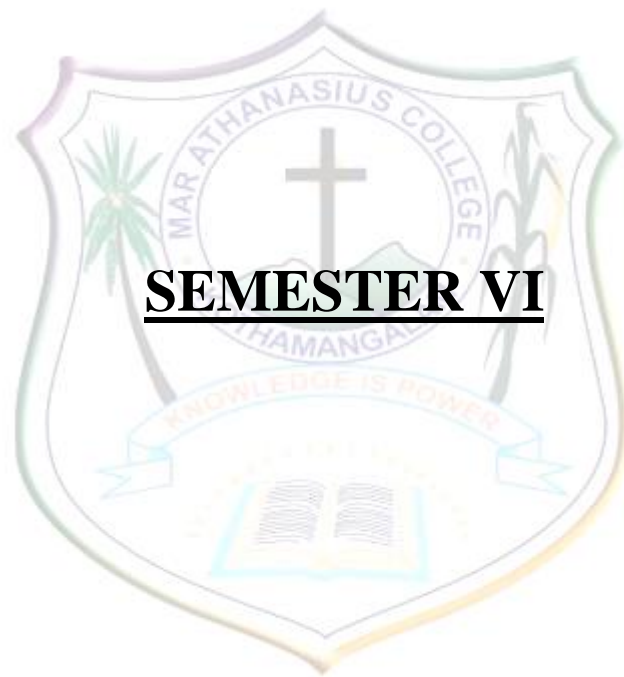
<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar
	<b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks


## References

1. Kaul, T. N.(2002). *Biology and Conservation of Mushroom*, Oxford and IBH Publishing Co.
2. Aneja, K.R. *Experiments in Microbiology, Plant Pathology, Tissue Culture and Mushroom Production Technology*. New Age International (P) Limited Publishers, Bangalore
3. Bahl,N. (2002). *Hand book on Mushrooms*. Oxford & IBH Publishing C. Pvt.
4. Chang, S.F., Miles, P.G. and Chang, S.T. (2004). *Mushrooms Cultivation, nutritional value, medicinal effect and environmental impact*. 2nd ed., CRC press.
5. Chang, S.T., Miles, P.G.(1979). *Edible Mushrooms and their Cultivation*. Boca Raton; CRC Press.

6. Marimuth et al., (1991). *Oyster Mushrooms*. Dept. of Plant pathology, TNAU, Coimbatore.
7. Nair, M.C. (1990). *Mushroom Technical Bulletin 17*. Kerala Agricultural University, Mannuthy.
8. Nita Bahl. (1988). *Hand book of Mushrooms*, 2nd Edition, Vol I & II.
9. Pandey, R.K. and Ghosh, S.K.(1996). *A HandBook on Mushroom Cultivation*. Emkey Publications.
10. Rai, R.D. and Arumuganathan, T. (2008) *Post Harvest Technology of mushrooms*, Technical bulletin, NRCM, ICAR, Chambaghat, Solan 1731213, Himachal Pradesh
11. Stamets, P. and Chilton, J.S. (2004). *Mushroom cultivation- A practical guide to growing mushrooms at home*, Agarikon Press.
12. Tewari, S.C. and Kapoor, P. (1993). *Mushroom cultivation*. Mittal Publication. Delhi.
13. <https://dmrsolan.icar.gov.in/>
14. <https://kau.in/institution/department-plant-pathology-0>





	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Plant Anatomy and Developmental Biology</b>					
<b>Type of Course</b>	<b>DSC</b>					
<b>Course Code</b>	<b>M24BO6DSC300</b>					
<b>Course Level</b>	<b>300</b>					
<b>Course Summary</b>	The course Plant anatomy and reproductive botany equips students with a deep understanding of the intricate structures and developmental processes in plants, enabling them to appreciate the complexity and beauty of plant life and its significance in the natural world.					
<b>Semester</b>	6	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Knowledge about structure of vegetative and reproductive parts of plants					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PO No
1.	Identify and differentiate tissues of plant organs	K, U	1
2	Relate the structural complexity of the cell wall and its applications.	U	1,2
3	Differentiate various anatomical changes under developmental stages and habitat conditions.	An	1,2
4	Interpret the applied aspects of anatomical studies in other branches of plant science.	A	5
5	Describe the structure and development of reproductive parts in angiosperms.	U	3

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
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1	<b>Module 1 (15 hours)</b>			
	1.1	Plant cell general view; Cell wall: Definition, Functions, Chemical composition of cell wall. Structure and function of plasmodesmata, simple and bordered pits, Growth of cell wall - apposition, intussusception.	3	1
	1.3	<b>Tissues:</b> Meristematic tissue – characteristic features, functions and classification. Permanent tissues - structure and function of simple and complex tissues. Secretory tissues: external secretory tissue - glands and nectaries; internal secretory tissues - laticifers.	4	2
	1.4	<b>Tissue systems:</b> Epidermal tissue system - epidermis, cuticle, trichomes; stomata – structure, types; bulliform cells. Ground tissue system - cortex, endodermis, pericycle, pith and pith rays. Vascular tissue system - structure of xylem and phloem, different types of vascular bundles and their arrangement in root and stem.	3	2
	1.5	Primary structure of stem and root (monocot and dicot) Normal secondary growth in dicot stem and root. Steps in secondary thickening: Intra-stelar secondary thickening, formation of cambium, structure and function of cambium, activity of cambium, role of cambium in wound healing, budding and grafting.	5	3
	<b>Module 2 (15 hours)</b>			
	2.1	Extra stelar secondary thickening: periderm – structure and development, bark, lenticels; factors affecting cambial activity, Seasonal activity of cambium, annual rings. Dendrochronology.	5	3
	2.2	Anomalous secondary thickening (general account).	2	3
	2.3	Types of wood; heartwood, sapwood, hard wood - porous nature, softwood - non porous nature (Brief study). Reaction wood: tension wood and compression wood.	2	4
	2.4	Identification of wood – using physical, microscopic, and macroscopic features. Identification of - fragmentary plant material as adulterants in crude drugs, food adulterants and contaminants. Archaeological plant remains and prediction of ancient	6	4

		<p>climatic conditions. Forensic investigations evidence, and taxonomic significance characters. Wood modification technologies for industry (Brief account only). Relevance of anatomical studies in crop science.</p>		
3	<b>Module 3 (15 hrs)</b>			
	3.1	Flower as a reproductive structure, floral parts, and their roles.	1	5
	3.2	Microsporangium and male gametophyte, Microsporangium: structure and development of anther, microsporogenesis, Male gametophyte development, dehiscence of anther, structure of pollen.	3	5
	3.3	Megasporangium and female gametophyte, Megasporangium: Types of ovules – anatropous, orthotropous. Megasporeogenesis – female gametophyte – structure of a typical embryo sac, types of embryo sacs - monosporic (Polygonum type).	4	5
	3.4	Fertilization: Mechanism of pollination, agents of pollination. Pollinators and global food security. Pollen pistil interaction, germination of pollen grains; double fertilization.	3	5
	3.5	Endosperm and Embryo development: Endosperm: types – cellular, nuclear and helobial. Embryogeny, structure of dicot and monocot embryo, fruit & seed formation. Polyembryony; Apomixis.	4	5
4	<b>Module 4. Practical (30 hrs)</b>			
	4.1	<p>I. Select and conduct any two of the following learning activities a/b/c/d (Individual/Group):</p> <p>a. Submission of an assignment on anatomical organization of the plant body based on the higher secondary level syllabus.</p> <p>b. Collect herbaceous members of dicot and monocot – prepare stained sections of root, stem, leaves, and flower bud.</p> <p>c. Prepare photographs of each and locate – Tissue types, epidermal, ground, and vascular tissue systems.</p> <p>d. Identify locally available plants with secretory tissues and prepare a report/ poster/audiovisual document.</p> <p>I. Micro preparation of dicot and monocot root and stem (Primary/Secondary).</p>	20	1, 3, 4


		II. Micro preparation of <i>Bignonia/Boerhavia</i> stem after secondary thickening.		
	4.2	I. Dissect a flower and document (photograph/illustration) II. Identification of C.S of the anther. III. Identification and documentation of anther dehiscence pattern in five locally available plants. IV. Pollen viability tests – Acetocarmine test / Tetrazolium test. V. Stigma receptivity test VI. Pollen germination test - Sugar solution test. VII. Dissection of dicot embryo.	10	5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc
	<b>B. End Semester Examination</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

### References

1. Beck, C. B. (2010). An introduction to plant structure and development: plant anatomy for the twenty-first century. Cambridge University Press.
2. Bhojwani, S. S, Bhatnagar, S. P., & Dantu, P. K. (2015). The embryology of angiosperms. Vikas Publishing House.
3. Cutler, D. F., Botha, C. E. J., Stevenson, D. W. (2008). Plant Anatomy: An Applied Approach. United Kingdom: Wiley.
4. Easu, K. (1977). Anatomy of seed plants (II Edn). Wiley Eastern, New York.

5. Evert, R. F. (2006). *Esau's plant anatomy: meristems, cells, and tissues of the plant body: their structure, function, and development*. John Wiley & Sons.
6. Hoadley, R. B. (1990). *Identifying Wood: Accurate Results with Simple Tools*. United States: Taunton Press.
7. Maheshwari, P. (1971). *An introduction to the embryology of angiosperms*. Tata McGraw Hill Publishing Company Ltd., New Delhi.
8. Maiti R, Satya P, Rajkumar D, and Ramaswamy A. (2012). *Crop plant anatomy*. CABI
9. Murphy, D. B., & Davidson, M. W. (2012). *Fundamentals of light microscopy and electronic imaging*. John Wiley & Sons.
10. Pandey, B. P. (2001). *Plant anatomy*. S. Chand Publishing.
11. Pandey, S. N. (2009). *Plant Anatomy and Embryology*. India: Vikas Publishing House Pvt Limited.
12. Panigrahi S and Rout, S (2020) Wood modification: An alternative strategy for use and protection of wood In *Agriculture and Forestry: Current Trends, Perspectives, Issues-I*. PP - 267 -286
13. Sandberg, D., Kutnar, A., & Mantanis, G. (2017). Wood modification technologies- a review. *Iforest-Biogeosciences and forestry*, 10(6), 895.
14. Stern, W. L. (1976). Multiple uses of institutional wood collections. *Curator: The Museum Journal*, 19(4), 265-270.
15. Vasishta, P. C. (2016). *Plant Anatomy*. Pradeep publication, Jalandhar.
16. Wiedenhoef, A. (2014). Curating xylaria. J. Salick, K. Konchar & Ma. Nesbitt (eds.), 127-134.
17. Yeung, E. C. T., Stasolla, C., Sumner, M. J., & Huang, B. Q. (Eds.). (2015). *Plant microtechniques and protocols* (No. 11831). Cham, Switzerland: Springer International Publishing.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Research Methodology, Biostatistics and Computer Application</b>					
<b>Type of Course</b>	<b>DSC</b>					
<b>Course Code</b>	<b>M24BO6DSC301</b>					
<b>Course Level</b>	<b>300</b>					
<b>Course Summary</b>	The course discusses various aspects of research – like how to find a research problem, the major sources of literature for research, the major steps in research, methods of report writing, use of ICT and statistics in research.					
<b>Semester</b>	6	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	1	-	
<b>Pre-requisites, if any</b>	Knowledge in science, numerical skills, and exposure to computer					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO
1	Discuss the basic concepts of research.	U	5
2	Identify and compile the various sources of literature for research.	U	5,6
3	Outline a research problem in Biology, design and write research report based on it.	An	3,6
4	Familiarize various available operating systems. Operate various tools in MS office/Libre Office to generate and present research reports.	A	4
5	Evaluate the data using various statistical tools and interpret the results.	A	6

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

**COURSE CONTENT**

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Objectives of research. Types of research - pure and applied. Identification of research problem, formulation of hypothesis – Null hypothesis and alternate hypothesis.	3	1
	1.2	Major steps, purpose, literature sources, names of reputed National and International journals in life science (Minimum 2 international & 3 national); reprint acquisition – INFLIBNET, PubMed, NCBI.	5	1,2
	1.3	Definition of the problem; Identification of the objective(s); literature review (brief account only), introducing working hypothesis, design of the study – basic principles and significance; sampling for data – methods, Identification and collection of data, types of data – Primary and Secondary; Collection of primary data – observation method, interview method, questionnaire method, through schedules; analysis and interpretation of data, Report writing (Brief account).	7	2,3
2	<b>Module 2 (15 hours)</b>			
	2.1	Preparation of dissertation - IMRAD system - Preliminary pages – Title pages – Certificate, Declaration, Acknowledgement, Table of contents, Abstract; Main text - Introduction and review of literature, Materials and methods, Results, Discussion, Conclusion; End matter – Bibliography and Appendix.	3	3
	2.2	Basic components of a computer – concept of Hardware and Software, Major Operating Systems: Proprietary: Windows, Macintosh and Open source: Linux. Application suit – M.S Office (Brief introduction).	4	4
	2.3	MS WORD - Word Processing - creating a new document, saving a document, exporting to pdf, opening an existing document, basic text editing; Editing tools – cut, copy, paste, find, and replace,	4	4

		undo and redo; Formatting tools – font formatting, paragraph formatting, bullets and numbering, styles, page formatting. Track change		
	2.4	MS EXCEL - creating worksheet, data entry, sorting data. Statistical tools (SUM, AVERAGE, MEDIAN and MODE). Preparation of graphs and diagrams (Bar diagram, Pie chart, Line chart, Histogram).	4	3,4
3	<b>Module 3 (15 hours)</b>			
	3.1	MS-POWERPOINT: Steps of preparation of presentation based on a topic from biology, which includes Tables, Charts, and Images. Ideal characteristics of a presentation slide set for scientific purposes using a model template.	2	4
	3.2	LibreOffice – Writer, Calc, Impress; Open Office (brief study). Search engines: Google.com; meta-search engine – Metacrawler; academic search engine - Google scholar. Educational sites related to biological science – Scitable, DNAi.	4	4
	3.3	Statistical terms, and symbols (Brief study only). Sampling: concept of sample, sampling methods - random and non-random sampling.	2	5
	3.4	Diagrammatic and graphical representation - line diagram, bar diagram, pie diagram, histogram, frequency curve.	2	5
	3.5	Measures of central tendency: mean, median, mode, (discrete and continuous series). Measures of dispersion: standard deviation. Probability and distribution patterns: normal distribution, binomial distribution. Tests of significance (t-test/Chi-square test).	5	5
4	<b>Module 4 Practical (30 hours)</b>			
	4.1	Preparation of a list of references (not less than 10) on a given topic of biological science. Preparation of Review on a given topic using online and print resources.	5	1,2
	4.2	Collect information on a topic related to biological science using the internet and make a report based on the collected information (Using M.S WORD / Libre Office Writer)	3	2,3

	4.3	<p>Collect a compound leaf with at least 25 leaflets of varying sizes from a plant, measure the length of each leaflet, and conduct the following works using M.S Excel/ Libre Office Calc and record:</p> <p>(a) Prepare data table/frequency table in M.S Excel / Libre Office Calc</p> <p>(b) Prepare bar diagram</p> <p>(c) Prepare Line chart</p> <p>(d) Prepare a Pie chart</p> <p>(e) Prepare Histogram</p> <p>Collect data on a particular topic using online or print questionnaires and perform the following activities in M.S Excel / LibreOffice Calc and record.</p> <p>(a) Calculate the average of variables</p> <p>(b) Calculate the median of variables</p> <p>(c) Calculate the mode of variables.</p> <p>Prepare a worksheet using a set of data collected and find out the SUM.</p>	15	3,4
	4.4	<p>Preparation of PowerPoint presentation using M.S PowerPoint / LibreOffice – Impress, based on a given topic.</p> <p>Problems related to</p> <p>a. Measures of central tendency</p> <p>b. Measures of dispersion</p> <p>c. Probability</p> <p>d. Test of significance (t – test/Chi-square test)</p>	7	4,5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b></p> <p>Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b></p> <p><b>A. Continuous Comprehensive Assessment (CCA)</b></p> <p>Theory Total = 25 marks</p> <p>Quiz, Test Papers, seminar</p> <p>Practical Total = 15 marks</p> <p>Lab performance, record, field report etc.</p>



	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>
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### References


1. Arora, P. N., Malhan, P. K. (2010). *Biostatistics*. India: Himalaya Publishing House.
2. Banerjee, P. K. (2007). *Introduction to biostatistics (a textbook of biometry)*. S. Chand Publishing.
3. Courter, G., Marquis, A. (2006). *Mastering Microsoft Office 2003 for Business Professionals*. Germany: Wiley.
4. Creswell, J. W., Creswell, J. D. (2017). *Research Design: Qualitative, Quantitative, and Mixed Methods Approaches*. United States: SAGE Publications.
5. Documentation Team, L. (2018). *LibreOffice 6.0 Writer Guide*. Australia: Friends of OpenDocument, Incorporated.
6. Foulkes, L. (2020). *Learn Microsoft Office 2019: A Comprehensive Guide to Getting Started with Word, PowerPoint, Excel, Access, and Outlook*. United Kingdom: Packt Publishing.
7. Gastel, B., Day, R. A. (2022). *How to Write and Publish a Scientific Paper*. United States: Bloomsbury Academic.
8. Goel, A. (2010). *Computer Fundamentals*. India: Pearson Education.
9. Holding, H., & Martin, C. (2001). *Mastering Microsoft Office*. Palgrave Master Series (Computing)
10. Holmes, D., Moody, P., Dine, D., & Trueman, L. (2017). *Research methods for the biosciences*. Oxford university press.
11. Jeffrey A Lee, (2009). *The Scientific Endeavour: Methodology and perspectives of sciences*. Pearson.
12. Khanal, A. B. (2015). *Mahajan's Methods in Biostatistics For Medical Students and Research Workers*. India: Jaypee Brothers Medical Publishers Pvt. Limited.
13. Khanna, R. (2008). *Basics of computer science*. New Age.
14. Kothari, C. R. (2004). *Research methodology: Methods and techniques*. New Age International.
15. Leete, G., Finkelstein, E., Leete, M. (2004). *OpenOffice.org For Dummies*. Germany: Wiley.
16. Rao, P. S., & Richard, J. (2012). *Introduction to biostatistics and research methods*. PHI Learning Pvt. Ltd..
17. Rosner, B. (2015). *Fundamentals of Biostatistics*. Brooks/Cole, Cengage Learning

### Websites

1. <https://www.inflibnet.ac.in/>
2. <https://pubmed.ncbi.nlm.nih.gov/>
3. <https://www.ncbi.nlm.nih.gov/>
4. <https://www.google.com/>

5. <https://www.metacrawler.com/>
6. <https://scholar.google.com/>
7. <https://www.nature.com/scitable/>
8. <https://www.dnai.org/>



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Microbial Biotechnology</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO6DSE300</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	The course in Microbial Biotechnology introduces students to the fundamental principles and techniques underlying the manipulation of microorganisms for various industrial and agricultural applications. Students explore the history and applications of fermentation processes, including aerobic and anaerobic fermentation, as well as submerged and solid-state fermentation, along with the factors influencing them.				
<b>Semester</b>	6	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		3	-	1	-
<b>Pre-requisites, if any</b>	Basic knowledge in microbiology and biotechnology				

#### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Understand applications of microbial biotechnology for the welfare of mankind.	U	5
2	Apply microbial biotechnology to solve problems in different sectors.	U	4,5
3	Analyze the potential of biotechnological tools for industrial applications.	An	1,4,5
4	Develop the skills for culturing microbes for industrial application.	C	4,5
5	Evaluate industrial and food microbiology processes.	E	5,6
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

#### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Microbial biotechnology, scope and techniques. Industrially important microorganisms, Gene transfer mechanisms in microbes, Transformation, transduction, conjugation and recombination.	5	1
	1.2	Genetic variability in microorganisms. Biotechnological tools to improve the microbial strains with respect to industry and agriculture.	2	1,2
	1.3	Microbial Growth: Definition, salient features of growth curve, generation time, specific growth rate, batch and continuous culture, synchronous growth, diauxic growth curve.	8	1,2
2	<b>Module 2 (15 hours)</b>			
	2.1	Brief history of fermentation, Fermentation-general concepts, Applications of fermentation	2	2,3
	2.2	Range of fermentation process- Microbial biomass, enzymes, metabolites, recombinant products, transformation process, Component parts of a fermentation process.	5	3,4,5
	2.3	Types of fermentations- Aerobic and anaerobic fermentation, Submerged and solid-state fermentation, Factors affecting submerged and solid-state fermentation, Substrates used in SSF and its advantages, Culture media- types, components and formulations. Sterilization: Batch and continuous sterilization.	8	4,5
3	<b>Module 3 (15 hours)</b>			
	3.1	Primary Metabolites- Production of commercially important primary metabolites like organic acids, amino acids and alcohols. Secondary Metabolites- Production processes for various classes of secondary metabolites: Antibiotics, Vitamins and Enzymes.	7	3,4

	3.2	<p>Exploitation of microorganisms and their products (curd, cheese, beer, alcohol, yoghurt).</p> <p>Single cell protein: microorganisms used; raw material used as substrate, condition for growth and production; nutritive value and uses of SCP.</p> <p>Baker's yeast; Production of probiotic biomass; and mold cultures. Mushroom production: cultivation of mushroom; edible mushroom.</p>	8	3,4
<b>Module 4 Practical (30 Hours)</b>				
4	4.1	<p>Laboratory Activities (Conduct Any Three)</p> <ol style="list-style-type: none"> <li>1. Isolation of microorganisms from different sources – air and water.</li> <li>2. Isolation of microbes by serial dilution and pour plate/ spread plate method</li> <li>3. Isolation of microbes by streak plate method</li> <li>4. Microbiological examination of foods. <ol style="list-style-type: none"> <li>i) Isolation and enumeration of bacteria and fungi from fresh and spoiled fruits.</li> <li>(ii) Isolation and enumeration of bacteria and fungi from fresh and spoiled vegetables.</li> <li>(iii) Isolation and enumeration of bacteria from fruit juices.</li> </ol> </li> <li>5. Effect of food preservatives on the growth of microbes.</li> <li>6. IMVIC test</li> <li>7. Oxidase test</li> <li>8. Catalase test</li> <li>9. Litmus milk test</li> <li>10. Hydrogen sulphide test</li> <li>11. Carbohydrate fermentation test</li> </ol>	20	4,5

	4.2	Laboratory/Industry visit: Students are expected to conduct a visit to a sophisticated biotechnology laboratory/research centre/biotechnology industry to have an idea on the type of work going on there.  A report of the visit should be prepared and submitted.	10	3,4
<b>5</b>		<b>Teacher specific course components</b>		


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

## References

1. Barnum, S. R., & Barnum, C. M. (2005). *Biotechnology: an introduction*. Thomson/Brooks/Cole.
2. Casida L E (2005). *Industrial Microbiology*. New Age International Limited.
3. Dube H C (2008). *Fungi, Bacteria and Viruses*. Agrobios.
4. El-Mansi, E. M. T. Bryce, C. F. A., Demain, A. L., Allman, A. R. (2007). *Fermentation Microbiology and Biotechnology (II Edn)*. Taylor & Francis.
5. Glick, B. R., & Pasternak, J. J. (2003). *Molecular biotechnology: principles and applications of recombinant DNA*.
6. Palmer, T., & Bonner, P. L. (2007). *Enzymes: biochemistry, biotechnology, clinical chemistry*. Elsevier.
7. Pandey, A. (2003). *Solid-state fermentation. Biochemical engineering*

- journal*, 13(2-3), 81-84.
8. Patel, A. H (2000). *Industrial Microbiology*. Macmillan Publishers.
  9. Pelczar Michael J, Adams M R, Chan E C S, Krieg Noel R (2000). *Microbiology*. Tata McGraw Hill.
  10. Prescott, S. C., & Cecil, G. D (2004). *Industrial Microbiology*. CBS publishers and distributors.
  11. Ratledge, C., & Kristiansen, B. (2001). *Basic biotechnology* cambridge university press.
  12. Sharma P D (2003). *Microbiology*. Restogi publishing.
  13. Stanbury, P. F., Whitaker, A., & Hall, S. J. (2013). *Principles of fermentation technology*. Elsevier



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Genetics and Evolution</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO6DSE301</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	This course provides a comprehensive exploration of the fundamental principles underlying genetics and evolution. Students will delve into the molecular basis of inheritance, the mechanisms of evolution, and the interconnectedness of these fields. Through theoretical discussions, practical applications, and case studies, participants will gain a deep understanding of how genetic processes drive evolutionary change.				
<b>Semester</b>	6	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		4	-	-	-
<b>Pre-requisites, if any</b>	History of genetics and contributions of Gregor Johann Mendel. Concept of gene and chromosome.				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PO No
1	Describe the basic principles of classical genetics and genetic interactions and discuss the non-mendelian patterns seen in nature	U	3,5
2	Estimate the linkage based genetic mapping in eukaryotes	E	3,5
3	Explain the types of sex determination mechanisms in higher organisms	U	3,5
4	Summarize the basics of population genetics	U	3,4,5
5	Transfer the concept of evolution in social inclusivity	A	2



**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	a) Terms & Concepts – chromosome, gene, allele-dominant and recessive, locus, genotype & phenotype, chromosome theory of inheritance, cross-monohybrid & dihybrid, testcross, backcross b) Principles of Mendelian Inheritance-Dominance, Segregation, and Independent Assortment. c) Model genetic organisms- <i>Neurospora crassa</i> , and <i>Arabidopsis thaliana</i> (mention only their importance in genetic study)	5	1
	1.2	Modifications of Mendelian ratios a) Incomplete dominance: Example - flower colour in <i>Mirabilis jalapa</i> . b) Co-dominance: Example - MN blood type in humans. c) Lethal genes: Example - pigmentation in Snapdragon. d) Epistasis: - Dominant epistasis: Example - fruit colour in summer squashes; Recessive epistasis – coat colour in mice <b><u>Learning activity:</u></b> <ul style="list-style-type: none"> <li>Workout Problems related to monohybrid cross, dihybrid cross, modified Mendelian ratios.</li> </ul>	10	1
2	<b>Module 2 (15 hours)</b>			
	2.1	a) Complementary gene interaction: Example - flower colour in <i>Lathyrus odoratus</i> . b) Multiple alleles: definition, example –Blood grouping in human ABO, Self-sterility in <i>Nicotiana tabaccum</i> . <b><u>Learning activity:</u></b> <ul style="list-style-type: none"> <li>Workout Problems related to complementary gene action and multiple alleles.</li> </ul>	5	1

	2.2	<p>a) Linkage – chromosome theory of linkage; complete and incomplete linkage.</p> <p>b) Crossing Over –mechanism of crossing over; types of crossing over – single, double and multiple; recombinant &amp; non-recombinant gametes</p> <p>c) Linkage mapping, recombination frequency &amp; map units; interference &amp; co-incidence.</p>	5	2
	2.3	<p>a) Extra chromosomal inheritance- cytoplasmic inheritance- Example: - leaf variegation in <i>Mirabilis jalapa</i>, Maternal effects - shell coiling in snail</p> <p>b) Quantitative inheritance: - polygenic; continuous traits. Example: ear size in maize; Quantitative trait Loci.</p>	5	1,2
<b>Module 3 (15 hours)</b>				
3	3.1	<p>a) Chromosomal mechanism of sex-determination: XX-XY, XX-XO, ZZ-ZW, Haplo-Diplo system, genic balance system.</p> <p>b) Environmental Sex Determination: Sex determination in slipper limpet and reptiles</p> <p>c) X-linked inheritance - Haemophilia in man; Y-linked inheritance – Holandric gene</p> <p>d) Sex-limited Inheritance – Example-feathering pattern in Fowl; Sex-influenced Inheritance - Example – Baldness in humans</p> <p>e) Mechanisms of sex determination in plants- <i>Melandrium</i> (emphasis on Epigenetic inheritance)</p>	10	3
	3.2	<p>Concept of Population, Allelic frequency, genotypic frequency; Hardy- Weinberg Equilibrium and the factors affecting the equilibrium.</p> <p><b><u>Learning activity:</u></b> Problems based on Hardy- Weinberg equation</p>	5	4
<b>Module 4 (15 hours)</b>				
4	4.1	<p>a.) Origin of life- biochemical origin of life (Miller’s Experiment). Theories of evolution -Darwin’s theory and modern synthetic theory. Evidences for evolution- (brief study)</p> <p>b.) Character evolution; Microevolution and macroevolution; Convergent, divergent, and parallel- evolution- (definition with examples)</p>	7	5

	4.2	<p>a.) Biological Species concept; speciation - genetic divergences and isolating mechanisms- geographical isolation &amp; reproductive isolation (prezygotic and postzygotic- isolation mechanisms)- (brief study)</p> <p>b.) Patterns of speciation- allopatric, sympatric, Peripatric, parapatric and quantum speciation - (brief study)</p> <p>c.) Population bottleneck and founder effect</p>	8	5
<b>5</b>	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b>  Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b>  <b>A. Continuous Comprehensive Assessment (CCA)</b>  Theory Total = 30 marks  Quiz, Test Papers, seminar</p> <hr/> <p><b>B. End Semester examination (ESE)</b>  Theory Total = 70 marks, Duration 2 hrs  Part A (Short answer) – 10 out of 12 x 2 = 20 marks  Part B (Short essay) – 6 out of 9 x 5 = 30 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>

## References

1. Coyne, A.J. (2009). Why Evolution is True, Viking Penguin, New York, USA.
2. Futuyma, D. J. (1998). Evolutionary biology. 3rd Sinauer Associates Inc, Publishers, Sunderland.
3. Gardner, E.J., Simmons, M.J., & Snustad, D.P. (1991). Principles of Genetics, John Wiley & sons, India. 8th edition.
4. Griffiths, A.J.F., Wessler, S.R., Carroll & S.B., Doebley, J. (2010). Introduction to Genetic Analysis. W. H. Freeman and Co., U.S.A. 10th edition.
5. Klug, W.S., Cummings, M.R., & Spencer, C.A. (2009). Concepts of Genetics. Benjamin Cummings, U.S.A. 9th edition.
6. Mayr, E. (1997). This is biology: The science of the living world. University Press.
7. Pandey S.N. (2006). A text book of Botany, Vikas Publishing House, New Delhi.
8. Pierce, Benjamin A. (2017). Genetics: A Conceptual Approach. W.H Freeman.

9. Simpson, G. G. (1953). The Major Features of Evolution, Oxford and IBH Publishing, New Delhi.
10. Snustad, D.P. & Simmons, M.J. (2010). Principles of Genetics, John Wiley & Sons Inc., India. 5th edition.
11. Strickberger, M. W. (2005). Evolution. Jones & Bartlett Learning.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Phytogeography, Forestry and Ecotourism</b>				
<b>Type of Course</b>	<b>DSE</b>				
<b>Course Code</b>	<b>M24BO6DSE302</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	The course 'Phytogeography, Forestry and ecotourism' deals with the study of distribution of plant community, its management and conservation.				
<b>Semester</b>	6	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		4	-	-	-
<b>Pre-requisites, if any</b>	Basic concepts of ecology				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Explain various theories and principles related to plant distribution.	U	1,2
2	Identify and categorize the interactions in the ecosystem and factors affecting the plant growth.	An	1,2
3	Describe the principles and practices in forest management.	U	2,5
4	Evaluate and appreciate the role of youth, Clubs, organizations in conservations.	Ap	2
5	Appreciate the role of ecotourism projects in nature conservations.	Ap	2,5,6

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
<b>1</b>	<b>Module 1 (15 hours)</b>			
	1.1	Ecological complexes and factors affecting plants growth and distribution. Biotic factors: interactions – positive and negative	3	2

	1.2	Topographic factors: altitude and aspects. Edaphic factors – soil profile and physical and chemical properties of soil, soil formation	4	2
	1.3	Climatic factors: temperature and pressure, water - precipitation, humidity, soil water holding capacity, light - global radiation	3	2
	1.4	Morphological, anatomical, and physiological adaptation of plants to the environment with references to biomes.	5	2
	<b>Module 2 (15 hours)</b>			
	2.1	Definition, principles governing plant distribution, factors affecting plant distribution	2	1
	2.2	Plant distribution- distribution of plants- continuous, discontinuous, and endemic. Theories of plant distribution – migration hypothesis, long distance dispersal hypothesis, theory of continental drift, age area hypothesis, land bridge theory.	5	1
	2.3	World Biomes - aquatic and terrestrial, Climatic, vegetational and botanical zones of India.	3	1
	2.4	Remote sensing - Definition and data acquisition techniques. Application of remote sensing in vegetation classification, understanding environmental issues and ecosystem management. Geographic information system (GIS).	5	1
	<b>Module 3 (15 hours)</b>			
	3.1	Introduction to forestry: Classification of forests (Champion and Seth, 1968). Major types of forests in India. Silviculture; principles and practices- clear felling system, coppice system. Common plants in silviculture. Sustainable forest management approaches with reference to Kerala - timber plantation, agroforestry, social forestry, JFM	5	3,4
	3.2	Forest Ecosystems and biodiversity- Forest ecology and ecosystem services. Biodiversity- definition, values of biodiversity, levels of biodiversity. Biodiversity loss, Concept of endemism. Types of endemism.	5	3
	3.3	Species extinction – Rate of species extinction, reasons to stop extinction- methods to save species. Threats to forest biodiversity, IUCN- threat categories. IUCN account of biodiversity, red data book and hot spots.	5	4
	<b>Module 4 (15 hours)</b>			
	4.1	Ecotourism definition, Elements and characteristics of ecotourism. Types of ecotourism – Heritage ecotourism, coastal ecotourism, cultural ecotourism, festival ecotourism, ayurvedic ecotourism. positive and negative impacts of ecotourism.	7	5
	4.2	Major ecotourism centers in Kerala – Gavi, Thattekadu, Thenmala.	4	5

		Learning activity: Visit an ecotourism center and identify the ecotourism components of the ecotourism and submit a report.		
	4.3	Wildlife tourism and its opportunities with reference to Kerala- Periyar tiger reserve, Tholpetty wildlife sanctuary	4	5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester examination (ESE)</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## References


1. Ballantyne, R., & Packer, J. (2013). International Handbook on Ecotourism. Edward Elgar Publishing.
2. Britannica Educational Publishing. (2010). Biomes and Ecosystems. Britannica Educational Publishing.
3. Köhl M., Magnussen, S. S., Marchetti, M., & Springerlink (Online Service. (2006). Sampling Methods, Remote Sensing and GIS Multiresource Forest Inventory. Springer Berlin Heidelberg.
4. Montagnini, F., & Jordan, C. F. (2005). Tropical Forest Ecology. Springer Science & Business Media.
5. Richardson, J., R. Björheden, P. Hakkila, Lowe, A. T., & Smith, C. T. (2006). Bioenergy from Sustainable Forestry. Springer Science & Business Media.
6. Schulze E. D., Beck, E., & Klaus Müller-Hohenstein. (2005). Plant ecology. Springer.
7. Stiling, P. D. (2015). Ecology: global insights & investigations. Mcgraw-Hill Education.
8. Woodward, S. L. (2009). Introduction to biomes. Greenwood Press.

## SUGGESTED READINGS

1. Begon, M., Harper, J. L., & Townsend, C. R. (2006). Ecology: individuals, populations, and communities. Blackwell Science.
2. Fennell, D. A. (2002). Ecotourism. Routledge.
3. Jennifer Hill, & Dr Tim Gale. (2012). Ecotourism and Environmental Sustainability. Ashgate Publishing, Ltd.
4. Newsome, D., Dowling, R. K., & Moore, S. A. (2005). Wildlife tourism. Channel View Publications.
5. Raven, C. (2006). Forestry. Infobase Publishing.
6. Wearing, S., & Neil, J. (2013). Ecotourism. Routledge.
7. Wearing, S., & Schweinsberg, S. (2019). Ecotourism: transitioning to the 22nd century. Routledge.





	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Entrepreneurial Botany</b>				
<b>Type of Course</b>	<b>SEC</b>				
<b>Course Code</b>	<b>M24BO6SEC300</b>				
<b>Course Level</b>	<b>300</b>				
<b>Course Summary</b>	The course aims to prepare the students for an entrepreneurial journey by giving an overview of entrepreneurship. The course discusses the process of developing an independent idea into ventures. Different areas of opportunity will also be addressed.				
<b>Semester</b>	6	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	
		3	-	-	-
<b>Pre-requisites, if any</b>	Basic knowledge in botany and its applications				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Demonstrate knowledge of diverse botanical entrepreneurship	A	5
2	Develop comprehensive business acumen for botanical ventures incorporating innovation, risk assessment and strategic solutions	A	4,5
3	Navigate and integrate government initiatives and support schemes in entrepreneurial endeavors	A	5
4	Analyze and evaluate real world success stories of entrepreneurs from government initiatives	An	4,5
5	Propose entrepreneurial ideas based on plant and	E,C	5,6

	plant-based product conducting preliminary research and evaluate the success stories in entrepreneurship		
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Types and Characterization of Botanical Entrepreneurship. Exploring various types, including agribusiness, bio ventures and aesthetics. Characterizing ventures based on botanical products.	3	1, 5
	1.2	Need, Entrepreneurial Values, Motivation, and Barriers- Analysing the socio-economic factors driving entrepreneurial endeavours in botany. - Identifying values, motivation factors, and overcoming	2	1,2
	1.3	Entrepreneurship as Innovation, Risk Assessment, and Solutions- Examining the role of innovation in botanical entrepreneurship. Assessing risks specific to botanical ventures and proposing strategic solutions.	2	2
	1.4	Overview of Key Botanical Industries in Kerala Exploration of <i>Spirulina</i> , edible mushrooms, cashew, spices and coconut industries within the Kerala context.	3	2,3
	1.5	Case studies on successful ventures like Jackfruit 360 ,Vegrow and Synthite Biotech startups and support mechanisms Zaara Biotech, KDISC, Bio 360- Life science park, BioNest	3	4
	1.6	Exploring the market for ornamental plants and flowers in Kerala. Opportunities and challenges specific to the aesthetics industry in the state.	2	2
2	<b>Module 2 (15 hours)</b>			
	2.1	Preservation and products: Canning and Processing of Fruits in Kerala -Overview of fruit canning processes and equipment, with a focus on Kerala's fruit varieties. Adapting processes to meet the unique demands of the Kerala fruit market.	2	2

	2.2	Fruit and vegetable-based products- production of juices, squashes and other fruit-based products considering Kerala's agricultural landscape	2	2
	2.3	Bamboo and cane-based products Plant based nutraceuticals, and oils- Herbal medicines and cosmetics	2	2
	2.4	Kerala Startup mission- schemes, support systems Start Up India. Overview of the Start Up India initiative, promoting innovation and entrepreneurship. Analyzing the support mechanisms provided to startups and success stories.	4	3,4
	2.5	MUDHRA Yojan- Overview, its role in funding micro enterprises. Practical insights into the application process and eligibility criteria. Stand Up India: Exploring the Stand-Up India scheme and its focus on promoting entrepreneurship among women and SC/ST communities.	5	3
	<b>Module 3 (15 hours)</b>			
3	3.1	Understanding the SC/ST Hub initiative and its role in supporting entrepreneurs from marginalized communities. Examining how the SC/ST Hub facilitates access to finance, markets, and capacity building.	3	3
	3.2	Navigating Government Support. Practical guidance on how entrepreneurs can navigate and access the above-mentioned government schemes	2	3
	3.3	Success Stories and Case Studies. Analysing real world success stories of entrepreneurs who have benefited from the mentioned government initiatives: BIRAC schemes, YIP, Atal innovation missions (Extracting key lessons and best practices.)	3	4
	3.4	Entrepreneurial Impact Assessment. Evaluating the impact of government schemes on entrepreneurial ventures. Discussing challenges faced and proposing solutions for improvement.	2	4
	3.5	<b>Activities</b> 1. Group Project on Government Schemes 2. Develop a comprehensive business plan integrating one or more government schemes and do presentations. 3. Case study on Success Stories	5	3, 4, 5

		<p>4. Each student presents an analysis of a chosen success story related to government support schemes.</p> <p>5. Propose an entrepreneurial idea based on plant and plant products, conduct preliminary research and submit a report.</p> <p>6. Make an audio-visual document of an interview with an entrepreneur.</p>		
4	<b>Teacher specific course component</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b> Interactive lectures, Group discussions, Problem-based learning, Flipped classroom, Discussion-based Learning, Case-based Learning, Experiential Learning, Inquiry-Based Learning, Game-Based Learning, Socratic Method, Peer Teaching, Simulations, Online Learning, Blended Learning, and other innovative approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar</p>
	<p><b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>


## References

1. Akther, F. (2023). Role of Skill India Programs in Fostering Entrepreneurship among Rural Youth in India. *Formosa Journal of Science and Technology*, 2(10), 2891-2902.
2. Altan, M. Z. (2019). Education for Creating an Entrepreneurship and Innovation Ecosystem. *Educația Plus*, 24(SI ISAT), 195-200.
3. Carayannis, E. G., Samara, E. T., & Bakouros, Y. L. (2015). Innovation and entrepreneurship. *Innovation, Technology, and Knowledge Management*. [Online]. Cham, Springer International Publishing. Available from: doi, 10, 978-3.
4. Chawla, S. L., Patil, S., Ahlawat, T. R., & Agnihotri, R. (2016). Present status, constraints and future potential of floriculture in India. *Commercial Horticulture*, 29-38.
5. Deeja, D. (2021). A Study on the problems of working capital of FPIs in Kerala. *Elementary Education Online*, 20(5), 5612-5612.
6. Emerging Lessons on Women's Entrepreneurship in Asia and the Pacific, ADB and The Asia Foundation, 2018.

7. Harisha, B. N. (2017). An economic analysis of floriculture in India. In *Proceedings of the Sixth Middle East Conference on Global Business, Economics, Finance and Banking (ME17 Dubai Conference)* (pp. 6-8).
8. Kumar<sup>1</sup>, R. S., & Manimegalai, G. (2004). Fruit and vegetable processing industries and environment. *Industrial Pollution & Management*, 97.
9. Lokare, P., & Patra, K. K. (2022). *Botanical entrepreneurship*. Book Saga Publications.
10. Milutinovic, Olivera & Anđelić, Slavica & Vukosavljević, Danijela & Pušara, Aleksandra. (2023). Role of innovation in Entrepreneurship development. *International Journal of Management Trends: Key Concepts and Research*. 2. 64-70. 10.58898/ijmt.v2i1.64-70.
11. Nakku, V. B., Agbola, F. W., Miles, M. P., & Mahmood, A. (2020). The interrelationship between SME government support programs, entrepreneurial orientation, and performance: A developing economy perspective. *Journal of Small Business Management*, 58(1), 2-31.
12. Radhakrishnan, A., Balan, S., Indulekha, V. P., Simi, S., & Krishnan, S. (2021). Potential, economics and constraints of mushroom cultivation in Wayanad, Kerala. *Journal of Krishi Vigyan*, 9(2), 171-176.
13. Rajeevan, P. K., Geetha, C. K., & Rajendran, P. (2016). Orchid-centric floriculture development in Kerala, India. In *International Symposium on Succulents and Other Ornamentals 1165* (pp. 15-26).
14. Sharma, V. P., Annepu, S. K., Gautam, Y., Singh, M., & Kamal, S. (2017). Status of mushroom production in India. *Mushroom Res*, 26(2), 111-120.
15. Vinitha, K. An Analysis of Performance of Agro-Based Industries in Kerala with Special Reference to Coconut. *KEIZAI: FOR WHOM THE BELL TOLLS?*, 91.
16. Wasnik, Anurag & Jain, Abhinav. (2023). Government Support for Startups: A Comprehensive Analysis of Funding Initiatives and the Role of the Indian Government in Nurturing the Startup Ecosystem. 10.31014/aior.1992.06.03.523.
17. Wickens, G. E. (2012). *Economic botany: principles and practices*. Springer Science & Business Media.
18. Youssef, A. B., Boubaker, S., & Omri, A. (2018). Entrepreneurship and sustainability: The need for innovative and institutional solutions. *Technological Forecasting and Social Change*, 129, 232-241.

## SUGGESTED READINGS

1. Kerala startup mission handbook 2021

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Environmental Science and Human Rights</b>					
<b>Type of Course</b>	<b>VAC</b>					
<b>Course Code</b>	<b>M24BO6VAC300</b>					
<b>Course Level</b>	<b>300</b>					
<b>Course Summary</b>	This course provides a comprehensive study of environmental sciences, covering pollution sources, their impacts on health and ecosystems, and mitigation techniques. It explores conservation biology, emphasizing the importance of biodiversity and threats like habitat destruction and climate change. Students learn about conservation strategies, environmental policies, and the intersection of environmental sciences with human rights, fostering critical thinking and advocacy skills for a sustainable and equitable world.					
<b>Semester</b>	6	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	-	-	
<b>Pre-requisites, if any</b>	No pre-requisites for this course.					

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains*</b>	<b>PSO No</b>
1	Describe the principles of ecology, ecosystem structure and function, and the importance of biodiversity.	U	2
2	Illustrate sustainable practices for the utilization of natural resources	An	2,5
3	Prioritize the control measures for air, water, and soil pollution by examining the environmental laws in India	An	2,5
4	Evaluate strategies and solutions aimed at biodiversity	An	2,5

	conservation from a global perspective.		
5	Develop the relevance of human rights in real-world scenarios to make responsible citizens.	A	2,5
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT

Module	Units	Course description	Hrs	CO No.
<b>Module 1 (15 hours)</b>				
<b>1</b>	1.1	Introduction to Environmental Science: a) Definition, scope & significance, multidisciplinary nature of environmental studies b) Principles of ecology, ecosystem structure and function, biodiversity and its importance	3	1, 2
	1.2	Natural Resources: a) Concept of resource b) Classification of natural resources (renewable and non-renewable) c) Sustainable practices for resource utilization	4	2
	1.3	Overview of Environmental Pollution: Definition and types of pollution. Overview of air, water, soil, noise, and light pollution.	1	3
		Air pollution: Air pollutants, types, sources, effect on plants and humans, control measures	2	3
	1.4	Water pollution: Common pollutants, sources, impact, control measures; water quality standards - DO and BOD; eutrophication.	2	3
	1.5	Soil Pollution: Causes, sources, solid waste, biodegradable, non-biodegradable, management of solid waste, composting, e-waste, waste management and recycling.	3	3
<b>Module 2 (15 hours)</b>				
<b>2</b>	2.1	Environmental issues: a) Global warming, greenhouse effect, causes and consequences of climate change, ozone layer depletion. b) Carbon sequestration. c) Carbon foot prints-Indian carbon footprint	3	4

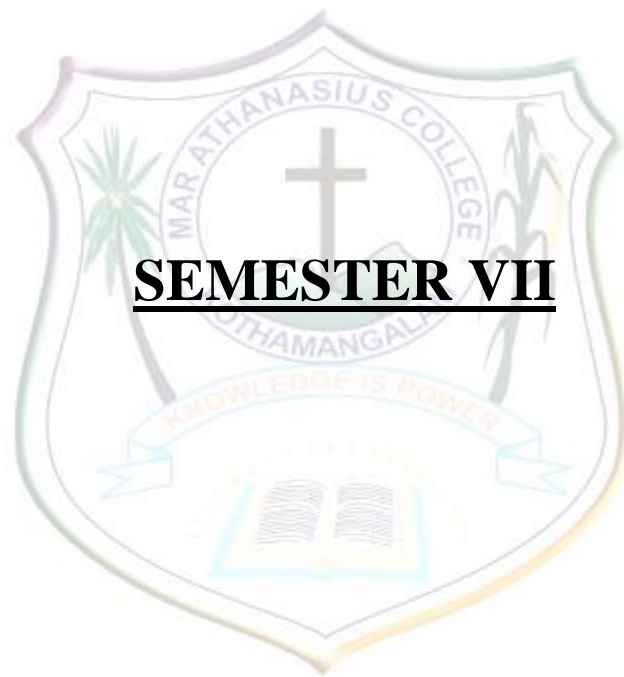
	2.2	<p>Global Conservation:</p> <p>a) Definition, importance, overview of threats to biodiversity</p> <p>b) International Conservation Organizations: Role of NGOs in Conservation (eg. WWF, Conservation International), United Nations Environment Programme (UNEP), International Union for Conservation of Nature (IUCN)-categories</p> <p>c) Overview of Key International Treaties (e.g., Kyoto Protocol, Paris Agreement)</p>	7	4
	2.3	<p>Overview of Environmental Legislation in India: Acts and Amendments</p> <p>a) Environment (Protection) Act 1986 and Environment (Protection) Amendment Rules, (2023)</p> <p>b) Wildlife (Protection) Act, 1972, amended in 2022,</p> <p>c) Forest (Conservation) Act, 1980, Forest (Conservation) Amendment Bill 2023 Biological Diversity (Amendment) Act, 2023 [brief account only].</p> <p>d) Corporate Environmental Responsibility [brief account only]</p>	5	4
	<b>Module 3 (15 hours)</b>			
	3.1	An introductory overview of human rights, history of Human Rights, Generations of Human Rights, Universality of Human Rights, Basic International Human Rights Documents - UDHR, ICCPR, ICESCR. - Value dimensions of Human Rights.	5	5
3	3.2	Human Rights and United Nations: Human Rights coordination within the UN system, Role of UN secretariat, Economic and Social Council, Commission of Human Rights, Security Council and Human Rights, Committee on the Elimination of Racial Discrimination, Committee on the Elimination of Discrimination Against Women, Committee on Economic, Social and Cultural Rights, The Human Rights Committee, Critical Appraisal of UN Human Rights Regime.	5	5
	3.3	Human Rights National Perspective: Human Rights in Indian Constitution, Fundamental Rights, Directive Principles of State Policy and Human Rights- Human Rights of Women-Children -Minorities-Prisoners, Science Technology and Human Rights- National Human Rights Commission- State Human Rights Commission- Human rights awareness in education.	5	5
4	<b>Teacher-specific course components</b>			



<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b> Interactive lectures, Group discussions, Problem-based learning, Flipped classroom, Discussion-based Learning, Case-based Learning, Experiential Learning, Inquiry-Based Learning, Game-Based Learning, Socratic Method, Peer Teaching, Simulations, Online Learning, Blended Learning, and other innovative approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar</p>
	<p><b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>

## References

1. Kumar, H. D. (2000). *Modern Concepts of Ecology*. Vikas Publishing House, New Delhi.
2. Kumar, U., Asija, M. (2006). *Biodiversity: Principles and conservation*. Agrobios India.
3. Misra, D. D., (2008). *Fundamental concepts in Environmental Studies*. S. Chand & Co. Ltd., New Delhi.
4. Nayar, M. P. (1997). *Biodiversity challenges in Kerala and science of conservation biology*. In: P. Pushpangadan, K S S Nair (Eds), *Biodiversity of tropical forests, the Kerala scenario*. STEC, Kerala.
5. Odum, E. P. (1971). *Fundamentals of Ecology*. WB Saunders.
6. Law Relating to Human Rights, Asia Law House, (2001).
7. Singh, S. P. (2011). *Human Rights Education in 21st Century*. Discovery Publishing House Pvt. Ltd. New Delhi.
8. Khanna, S. K. (1998). *Children and the Human Rights*. Commonwealth publishers.
9. Kapoor, S. (2001). *Human Rights in the 21st Century*. Mangal Deep Publications, Jaipur.
10. United Nations Development Programme, Human Development Report (2004). *Cultural liberty in today's diverse world*. Oxford University Press, New Delhi.
11. Santhra, S. C. (2004). *Environmental Science*. New Central Book Agency.
12. Panday, S. N., Misra, S. P. (2011). *Environment and Ecology*. Ane Books Pvt.Ltd. New Delhi
13. Sharma, P. D. (1999). *Ecology and Environment*. Rastogy Pub.
14. Varma, P. S., Agarwal, V. K. (1983). *Principles of Ecology*. S Chand and Co.



**SEMESTER VII**



**Mar Athanasius College (Autonomous), Kothamangalam**

**FYUGP SYLLABUS**

<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Modern Trends in Thallophytes</b>					
<b>Type of Course</b>	<b>DCC</b>					
<b>Course Code</b>	<b>M24BO7DCC400</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	This course will enable the students to identify, and compare the characteristics of the major groups of thallophytes and to classify them within a phylogenetic framework. Students will be able to use the evidence of comparative biology to correlate the evolutionary trends to the diversity of plant life on earth. Knowledge about the interactions and associations of lower plants will provide better insights on the adaptive strategies of plants. Awareness in the thrust areas of research will generate interest in students to pursue the same.					
<b>Semester</b>	7	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Knowledge in morphology and reproduction of thallophytes					

**COURSE OUTCOMES (CO)**

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Explore the course of evolution of algae	U	1
2	Analyze the diversity of thallus forms in algae and its adaptive strategies to diverse environments.	An	1,4
3	Review the affinities of fungi with other groups and differentiate morphological forms within the group.	E	1,4
4	Generate interest in recent research trends in Thallophyta.	I	5,6
5	Provide deep insights on the significance of thallophytes in the ecosystem, and the need for considering and conserving them	Ap, E	2

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

**COURSE CONTENT**

Module	Units	Course description	Hours	CO No.
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1	<b>Module 1 (15 hours)</b>			
	1.1	The range of thallus diversity in the algae. Polyphyletic origin of algae and its evolution, with emphasis on endosymbiosis. Evolution and structural variations of chloroplast in algae. Algal pigments involved in photosynthesis. Algal responses to light- phototaxis, photophobia, and gliding.	6	1
	1.2	Adaptation strategies of algae to different environmental conditions-Resting spores, Allelopathy in algae.	4	2
	1.3	Algae and the fossil record; Gene sequencing (18SrRNA, HTS) in algal systematics.	3	4
	1.4	Algal symbiosis-extracellular (lichens, association of cyanobacteria with <i>Azolla</i> , Coralloid roots) and intracellular associations. Nitrogen fixation by blue-green algae.	2	2
2	<b>Module 2 (15 hours)</b>			
	2.1	General features of fungi. Affinities with plants and animals. Morphological diversity of fungi- an overview (Slime molds, Mycelial and non-mycelial fungi).	3	3
	2.2	Modern trends in fungal classification; Molecular phylogeny of fungi with emphasis on 18SrRNA sequencing.	2	3,4
	2.3	Types of Fungal spores and its dispersal mechanisms (Ballistic dispersal, Dispersal by gravity, wind, water, insects and animals)	2	3
	2.4	Fungal Parasites - Common fungal parasites of plants, humans, insects and nematodes (Brief account only). Saprophytes - Fungal decomposition of organic matter, coprophilous fungi, cellulolytic fungi, lignolytic fungi.	3	3,5
	2.5	Lichens- Ecological role, Nature of associations of algal and fungal partners with emphasis on its nutritional relation, Establishment of a lichen thallus - the process. Mycorrhiza-Ectomycorrhiza, Endomycorrhiza and their significance. Phosphate solubilisation. Fungus-insect mutualism- Fungal farming by ants.	5	5
3	<b>Module 3 (15 hours)</b>			
	3.1	Applications of algae. Algae as the source of food and fodder. Algal polysaccharides-its commercial utilization. Algae as the source of diatomaceous earth, pigments, fatty acids and pharmaceuticals.	3	4

		Production of biofuel, biogas and bioplastics from algae. Algae as pollution indicator, algae-based wastewater treatment for biodiesel production, phycoremediation and biodegradation of plastics. Algae in soil fertility: Soil algae and cyanobacteria.		
	3.2	Algal blooms: Beneficial, harmful and toxic bloom. Common cultivated algal species in India. Algal research stations in India.	2	4,5
	3.3	Algal culture: scope and a brief account on isolation and culturing techniques (Axenic, Clonal, Unialgal, Enrichment, Maintenance, Batch, Continuous and Immobilized Culture) Molecular genetic techniques for algal bioengineering (Brief Account only), phylogenomics in algal research (Brief Account only) - current trends. Gene sequencing in algal systematics.	3	4
	3.4	Fungi in the food industry-Flavour & texture, Fermentation, Baking. Application of fungi in agriculture-Mycoherbicides, Mycoinsecticides, Myconematicides. Fungi as a biofertilizer Fungi as the source of Mycotoxins-Aflatoxins, Amatoxin, Ergot, Fusarin	3	4
	3.5	Commercial production of Organic acids, Enzymes, Plant hormones Mycoproteins, and alcohol from fungi. Antibiotics from fungi- penicillin, cephalosporin, Griseofulvin, Volatile organic compounds production by fungi.	2	4
	3.6	Fungi as plant and animal pathogen. Fungi as a model organism in genetic experiments-Neurospora, Saccharomyces. Recent research trends in fungi-Zombie ant fungi, Adaptive cognitive behavior and learning in slime molds.	2	4,5
	<b>Module 4 Practical (30 hours)</b>			
4	4.1	Study of the thallus morphology of the following algal genera; Cyanophyceae: <i>Gleocapsa</i> , <i>Microcystis</i> , <i>Lyngbya</i> , <i>Oscillatoria</i> , <i>Scytonema</i> , <i>Nostoc</i> , <i>Anabaena</i> .  Chlorophyceae: <i>Pandorina</i> , <i>Oedogonium</i> , <i>Zygnema</i> , <i>Mougeotia</i> , <i>Pithophora</i> , <i>Bulbochaete</i> , <i>Zygnema</i> , <i>Cosmarium</i> , <i>Chaetophora</i> , <i>Nitella</i> , <i>Caulerpa</i> , <i>Ulva</i> . Bacillariophyceae: <i>Cyclotella</i> , <i>Navicula</i> Phaeophyceae: <i>Ectocarpus</i> , <i>Turbinaria</i> , <i>Padina</i> , <i>Dictyota</i>	10	2,4

		Rhodophyceae: <i>Batrachospermum</i> , <i>Gracilaria</i> , <i>Gelidium</i> , <i>Kappaphycus</i>		
	4.2	Morphological study of the following types by preparing suitable micro preparations of the following fungi <i>Albugo</i> , <i>Rhizopus</i> , <i>Mucor</i> , <i>Aspergillus</i> , <i>Pilobolous</i> , <i>Xylaria</i> , <i>Peziza</i> , <i>Pleurotus</i> , <i>Auricularia</i> , <i>Lycoperdon</i> , <i>Fusarium</i> . Lichen- <i>Usnea</i> Isolation of fungi from rotten vegetables and culturing the same on PDA; Staining and observing VAM Fungal spore staining using lactophenol cotton blue. Lichen identification- morphological and chemical methods.	10	3,4
	4.3	<b>Activity:</b> Conduct a field visit to familiarize various habitats of algae and fungi and submit a report with geo-tagged photos related to their diversity.	10	2,3,4,5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.

	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>
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## References

1. Alexopoulos, C.J., Mims, C.W., & Blackwell, M. (1996). *Introductory Mycology*. John Wiley & Sons (Asia) Singapore. 4th edition.
2. Bold, H.C., & Wynne M.J. (1978). *Introduction to Algae: Structure and reproduction*. Prentice Hall, New Jersey.
3. Deacon, J. W. (2005). *Fungal biology*. John Wiley & Sons.
4. Fritsch, F.E. (1935). *The Structure and Reproduction of the Algae*. Volume I. Cambridge University Press, Cambridge.
5. Iyengar, M. O. P., & Desikachary, T. V. (1981). *Volvocales*. I.C.A.R., New Delhi. pp- 532.
6. Jacob-Lopes, E., Maroneze, M. M., Queiroz, M. I., & Zepka, L. Q. (Eds.). (2020). *Handbook of microalgae-based processes and products: fundamentals and advances in energy, food, feed, fertilizer, and bioactive compounds*, Academic Press, New York.
7. Kim, S.K. (2011). *Marine medicinal foods: Implications and Applications of micro and macroalgae*. Academic Press, New York.
8. Lee, R. (2008). *Phycology* (4th ed.). Cambridge: Cambridge University Press. doi:10.1017/CBO9780511812897
9. Linda, E.G & Lee, W.W., (2000). *Algae*. Prentice Hall, New Jersey.
10. Mamatha, Rao. (2009), *Microbes and Non flowering plants- impact and application*. Ane Books Pvt Ltd.
11. Moheimani, N. R., McHenry, M. P., De Boer, K., & Bahri, P. A. (2015). Biomass and biofuels from microalgae. *Biofuel and biorefinery technologies*. Springer-Verlag GmbH.
12. Pringsheim E G. (2016). *Pure culture of Algae*. Cambridge University Press
13. Sahho, D., & Seckback, J. (2015). *The algae world*. Springer Dordrecht Heidelberg, New York.
14. Sethi, L.K., & Walia, S.K. (2011). *Text book of Fungi and Their Allies*. Macmillan Publishers India Ltd.
15. Sharma, O.P (2011). *Algae*. Tata McGraw- Hill, New Delhi, p.419.
16. Smith, G.M. (1938). *Cryptogamic Botany (Vol. 1): Algae and Fungi*. Tata McGraw Hill Edition.
17. Webster, J., & Weber, R. (2007). *Introduction to Fungi*. Cambridge University Press, Cambridge. 3rd edition.

## Websites

<https://www.algaebase.org/>

[https://www.cell.com/current-biology/pdf/S0960-9822\(19\)31164-9.pdf](https://www.cell.com/current-biology/pdf/S0960-9822(19)31164-9.pdf)


<https://www.mycobank.org/>

<https://www.nature.com/articles/nature.2012.11811>

<https://www.routledge.com/Algal-Biotechnology-Current-Trends-Challenges-and-Future-Prospects-for/Sahu-Sridhar/p/book/9781032112688>





	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Plant Growth and Metabolism</b>					
<b>Type of Course</b>	<b>DCC</b>					
<b>Course Code</b>	<b>M24BO7DCC401</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	The course is designed to make students aware of advances and applications in Plant Metabolism. After completion of the course, the students would be able to; recall and articulate key concepts related to plant metabolism, including the pathways involved in energy production, biosynthesis of essential compounds, and regulatory mechanisms governing metabolic processes in plants.					
<b>Semester</b>	7				Credits	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	Total Hours
		4	-	-	-	
<b>Pre-requisites, if any</b>	Knowledge about plant cells, cell interaction, cytoskeleton, nucleic acids, plant physiology					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Describe the concepts of and fundamental principles of plant metabolism	U	3
2	Examine molecular and cellular processes in growth and development of plants	A	5
3	Analyse plant responses to environmental variables and regulatory mechanisms	An	3
4	Evaluate energy conversion and metabolic processes in plants	E	5
5	Illustrate the molecular mechanisms associated with flowering in plants	An	4

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Growth and Development in Plants- Patterns of growth and differentiation- Gene expression and mutations regulating meristem function, embryogenesis, seedling, root and leaf.	3	1,2
	1.2	Assimilation of inorganic nutrients: Nitrate assimilation, ammonium assimilation, biological nitrogen fixation, sulfur assimilation. The energetics of nutrient assimilation.	6	1,2
	1.3	Plant growth regulators: Biosynthesis (brief study), transport, physiological functions and commercial uses of the following plant growth regulators. Auxins, Gibberellin, Cytokinin, Ethylene, Abscisic acid, Brassinosteroid.	6	1,2,
2	<b>Module 2 (15 Hours)</b>			
	2.1	Photosynthesis: Chloroplast encoded genes and nuclear-encoded genes of photosynthesis. Genes encoding components of PET, Calvin cycle and other regulatory mechanism. Light absorption and energy conversion, electron transfer system in chloroplast membranes and ATP synthesis in chloroplast. Photosynthetic carbon reduction (C4 and CAM metabolism) photorespiration.	4	1,4
	2.2	Starch and sucrose synthesis, distribution of photo assimilates- Phloem loading and unloading. Concept of osmotically generated pressure flow. Importance of plasmodesmata in symplastic transport. Physiological and environmental consideration of photosynthesis.	4	1,4
	2.3	Respiration: Three stages of respiratory metabolism (brief study only). Plant mitochondrial electron transport and ATP synthesis – structure of electron transfer complexes (complex I – IV). ATPase – detailed structure of F1 and Fo subunits, binding change mechanism of ATP synthesis. Comparison of mitochondrial and chloroplast ATP synthesis. Cyanide resistant pathway - alternative oxidase, its regulation and significance. Lipid metabolism in oilseeds – glyoxylate cycle, gluconeogenesis	7	1,4


<b>Module 3 (15 Hours)</b>				
3	3.1	Flowering- Flowering-floral induction, evocation and morphogenesis. Floral organ identity genes (ABCD model in <i>Arabidopsis</i> ). Control of flowering-phytochrome, cryptochrome and biological clock. Factors affecting flowering: Photoperiodism and thermoperiodism.	6	1,3,5
	3.2	Phytochromes- structure, photochemical and biochemical properties, phytochrome induced plant responses. Cryptochromes: blue light hormones, photo physiology, cryptochrome induced plant responses.	6	1,3,5
	3.3	Phototropins- structure, regulation in chloroplast movement. UV light receptor and responses.	3	1,3,5
<b>Module 4 (15 Hours)</b>				
4	4.1	Senescence and programmed cell death: Apoptosis and necrosis. Programmed cell death in relation to reproductive development, and stress response. Genes associated with senescence, metabolism during senescence.	4	1,4
	4.2	Stress physiology: Different stress factors - Biotic stress, Water deficit and drought resistance, heat stress and heat shock, chilling and frost, salinity stress, high light stress, oxygen deficiency stress and heavy-metal pollution stress. Signaling pathways activated in response to abiotic stress. Salicylic acid and jasmonic acid mediated stress tolerance.	5	1,5
	4.3	Stress-associated changes in metabolites and metabolomics, homeostasis events under stress. Role of heat shock proteins (Hsp) in stress tolerance (Classification, role in protein refolding and resolubilizing, major functions). Transgenic plants for stress tolerance.	6	1,5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment</b>	<b>MODE OF ASSESSMENT</b>

<b>Types</b>	<b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester Examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## References

1. Buchanan, B. B., Gruissem, W., and Jones, R. L. (2015). *Biochemistry and molecular biology of plants*. 2nd ed. Wiley-Blackwell.
2. Dayananda, B. (1999). *Experiments in Plant Physiology*. Narosa Publishing House, New Delhi.
3. Heldt, H.-W., and Piechulla, B. (2021). *Plant Biochemistry*. 5th ed. Academic Press
4. Hopkins, W. G., Huner, N.P.A. (2008). *Introduction to plant physiology*. John Wiley and sons. New York.
5. Jain, J.L., Jain, S., Jain, N. (2005). *Fundamentals of Biochemistry*. S Chand, New Delhi.
6. Lehninger, A. L. (1961). *Biochemistry*. Lalyan publishers, Ludhiana.
7. Nelson, D. L., Cox, M.M. (1993). *Principles of Biochemistry*. MacMillan Publications.
8. Pandey, S.N., Sinha, B. K. (2006). *Plant Physiology*. Vikas Publishing House Pvt. Ltd
9. Plummer, D.T. (1988). *An introduction to practical biochemistry*. Tata McGraw-Hill publishing Company, New Delhi.
10. Sadasivam, S., Manickan, A. (1996). *Biochemical Methods*. New Age International Ltd. New Delhi.
11. Salisbury, F.B., Ross, C.W. (1992). *Plant Physiology*. CBS Publishers and Distributers, Delhi.
12. Srivastava, H. S. (2005). *Plant Physiology*. Rastogi publications, Meerut.
13. Taiz, L., Moller, I. M., Murphy, A., and Zeiger, E. (2023). *Plant Physiology and Development*. 7th ed. USA: Sinauer Associates Inc. Publisher
14. Taiz, L., Zeiger, E. (2002). *Plant Physiology* (III Edn). Panima publishing Corporation, New Delhi.
15. Verma, V. (2007). *Textbook of Plant Physiology*. Ane Books India, New Delhi.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Advances in Molecular Cell Biology and Immunology</b>					
<b>Type of Course</b>	<b>DCC</b>					
<b>Course Code</b>	<b>M24BO7DCC402</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	The course is designed as a comprehensive exploration to the advances of molecular biology and immunology. The course explores cell signaling mechanisms, cytoskeleton dynamics, and cell cycle control. It addresses cancer biology, DNA repair, and genetic recombination. The course also covers gene expression and regulation processes. Additionally, it delves into the immune system, including innate and acquired immunity, antibody functions, antigen processing, and immune responses.					
<b>Semester</b>	7		Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		4	-		-	60
<b>Pre-requisites, if any</b>	General overview and key concepts of Biotechnology					

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Apply principles of cell signaling, communication and understanding of gene expression regulation to cellular function	E	3,4
2	Analyze signal transduction pathways	E	3,4
3	Evaluate mechanisms of cytoskeleton dynamics and cell cycle regulation:	A	3
4	Analyze the genetic basis and molecular	A	3,4

	mechanisms of cancer		
5	Understand the principles of innate and acquired immunity, including the roles of immune cells and its functions.	U	3
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hrs)</b>			
	1.1	<b>Cell signaling &amp; communications</b> -Types of signals and signalling molecules, Types of Cell surface receptors- ion-channel linked receptors, G-protein coupled receptors, and Tyrosine-kinase linked receptors (RTK), Steroid hormone receptors.	10	1
	1.2	Signal transduction pathways- ras/map kinase pathway, phosphoinositide pathway, receptor serine kinases, secondary signal molecules, second messengers, calcium signaling, two component regulatory systems, bacterial chemotaxis, quorum sensing and quorum sensing disruptors.	5	1
2	<b>Module 2 (15hrs)</b>			
	2.1	<b>Cytoskeleton and Eukaryotic cell cycle</b> - Structure, assembly, disassembly and regulation of filaments involved – actin filaments (microfilaments), microtubules, and intermediate filaments. Functions of cytoskeleton;  Molecular motors – kinesins, dyneins, myosins. overview of the cell cycle and its control, regulation of CDK activity, commitment to the cell cycle and DNA replication, processes and steps of mitosis and meiosis, surveillance mechanisms in cell cycle regulation.	10	2
	2.2	<b>Cancer biology</b> - Genetic basis of cancer. Proto-oncogenes, oncogenes, conversion of proto-oncogenes to oncogenes. Tumor suppressor genes – functions, role of p53. Viral oncogenes.	5	3

<b>Module 3 (15 hrs)</b>				
<b>3</b>	3.1	<p>DNA repair mechanisms: Direct repair, excision repair – base excision repair and nucleotide excision repair (NER), eucaryotic excision repair – GG-NER, TC-NER. Mismatch repair, Recombination repair – homologous recombination repair, nonhomologous end joining, SOS response – Translesion DNA polymerase.</p> <p>Recombination: Homologous and nonhomologous recombination, molecular mechanism of homologous recombination. Site-specific recombination, transposition- types of transposons</p>	5	1,4
	3.2	<p><b>Gene expression</b> Molecular mechanism of Gene Regulation in Eukaryotes- Controlled transcription of DNA, Alternate splicing of RNA, Cytoplasmic control of mRNA stability, Induction of transcriptional activity by environmental and biological factors- Temperature- Heat shock proteins, Genes that respond to hormones- Proteins involved in control of transcription, transcriptional factors, activator proteins, enhancers, silencers, eukaryotic transcription complex, chromatin remodelling during gene expression, alternative promoter-</p> <p>Post transcriptional regulation, RNA interference, siRNAs, miRNAs, untranslated regions (UTRs), nonsense mediated decay, chromatin remodelling, DNA methylation, Imprinting.</p>	10	1,4
<b>Module 4 (15 hrs)</b>				
<b>4</b>	4.1	<p><b>Immunology</b> Innate and acquired immunity. Cells and molecules involved in innate and acquired immunity, humoral and cellular immunity Antigens, Epitopes. Structure, function and types of antibody molecules. Antigen-antibody interactions. Antigen processing and presentation. Activation and differentiation of B cells – formation, role.</p> <p>T cells – types, roles, T cell receptor. Primary and secondary immune modulation, complement system, pattern recognition receptors</p>	10	5

		– toll-like receptors. MHC molecules.		
	4.2	Cell-mediated effector functions, inflammation, hypersensitivity (Type I & II) and autoimmunity, congenital and acquired immune deficiencies.	5	
<b>5</b>	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar</p>
	<p><b>B. End Semester Examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>

## References

- Wayne M Becker, Lewis J Kleinsmith, Jeff Hardin (2007). *The world of the cell* (VI Edn). Pearson.
- Geoffrey M Cooper, Robert E Hausman (2009). *The Cell: A molecular approach* (V Edn). Sinauer.
- Gerald Karp (2008). *Cell and Molecular biology: Concepts and experiments* (V Edn). John Wiley & Sons.
- Harvey Lodish, Arnold Berk, Lawrence Zipursky, Paul Matsudaira, David Baltimore, James Darnell (2000). *Molecular cell biology* (IV Edn). W H Freeman & Company.
- Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter (2002)
- Molecular biology of the cell* (IV Edn). Garland Science, Taylor and Francis group.
- Robert J Brooker (2009). *Genetics: analysis and principles* (III Edn). McGraw Hill
- Jocelyn E Krebs, Elliott S Goldstein, Stephen T Kilpatrick (2011). *Lewin's Genes X*. Jones and Bartlett Publishers.
- Bob B Buchanan, Wilhelm Gruissem, Russel L Jones (2000). *Biochemistry and Molecular biology of plants*. I K International Pvt. Ltd.
- Daniel L Hartl, Elizabeth W Jones (2012). *Genetics: Analysis of genes and*



- genomes* (VII Edn). Jones and Bartlett publishers.
11. James D Watson, Tania A Baker, Stephen P Bell, Alexander Gann, Michael Levine, Richard Losick (2009). *Molecular biology of the gene* (V Edn). Pearson.
  12. William S Klug, Michael R Cummings (2004). *Concepts of Genetics* (VII Edn). Pearson.
  13. Daniel J Fairbanks, W Ralph Anderson (1999). *Genetics: The continuity of life*. Brooks/Cole publishing company.
  14. Robert F Weaver (2002). *Molecular biology* (II Edn). McGraw Hill.
  15. Bruce Alberts, Dennis Bray, Karen Hopkin, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter (2010). *Essential Cell Biology*. Garland Science.
  16. Leland H Hartwell, Leroy Hood, Michael L Goldberg, Ann E Reynolds, Lee M Silver, Ruth C Veres (2004). *Genetics from genes to genomes* (II Edn). McGraw Hill.
  17. Harvey Lodish, Arnold Berk, Chris A. Kaiser, Monty Krieger, Matthew P. Scott, Anthony Bretscher, Hidde Ploegh, Paul Matsudaira (2007). *Molecular cell biology* (VI Edn). W H Freeman & Company.
  18. James D. Watson, Amy A. Caudy, Richard M. Myers, Jan A. Witkowski (2007). *Recombinant DNA* (III Edn). W H Freeman.
  19. William H Elliott, Daphne C Elliott (2001). *Biochemistry and molecular biology* (II Edn). Oxford.
  20. Jeremy M Berg, John L Tymoczko, Lubert Stryer, Gregory J Gatto Jr. (2007). *Biochemistry*. W H Freeman & company.
  21. David P Clark (2010). *Molecular biology*. Elsevier.
  22. David R Hyde (2010). *Genetics and molecular biology*. Tata McGraw Hill.
  23. D Peter Snustad, Michael J Simmons (2010). *Principles of genetics* (V Edn). John Wiley and Sons.
  24. David A Micklos, Greg A Freyer with David A Crotty (2003). *DNA Science: A first course* (II Edn). L K Inter.
  25. Benjamin A Pierce (2008). *Genetics: A conceptual approach* (IV Edn). W H Freeman and Company.
  26. Anthony J F Griffiths, Susan R Wesler, Sean B Carroll, John Doebley (2012). *Introduction to genetic analysis*. W H Freeman & Company.
  27. T A Brown (2002). *Genomes* (II Edn). Bios.
  28. Robert H Tamarin (2002). *Principles of genetics*. McGraw Hill.
  29. David E Sadava (2009). *Cell biology: Organelle structure and function*. CBS.
  30. Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter (2010). *Essential Cell Biology* (III Edn.). Garland Science.
  31. Pranav Kumar, Usha Mina (2011). *Biotechnology: A problem approach*. Pathfinder Academy.
  32. Burton E Tropp (2012). *Molecular biology: Genes to Proteins* (IV Edn). Jones and Bartlett Learning.
  33. Lynne Cassimeris, Viswanath R Lingappa, George Plopper (Eds) (2011). *Lewin's Cells* (II Edn). Jones and Bartlett Publishers.
  34. Richard A Goldsby, Thomas J Kindt, Barbara A Osborne, Janis Kuby (2003). *Immunology* (V Edn). W H Freeman and Company.
  35. Peter J Delver, Seamus J Martin, Dennis R Burton, Ivan M Roitt (2011). *Roitt's essential immunology* (XII Edn). Wiley Blackwell

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Ecology and Environmental Conservation</b>				
<b>Type of Course</b>	<b>DCE</b>				
<b>Course Code</b>	<b>M24BO7DCE400</b>				
<b>Course Level</b>	<b>400</b>				
<b>Course Summary</b>	This course elaborates the concepts of plant ecology and environmental conservation. By the end of the course, students should be familiar with principles of biodiversity assessment, conservation and sustainable use of natural resources at local, regional, and global scales.				
<b>Semester</b>	7	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		4	-	-	-
<b>Pre-requisites, if any</b>	Basics of ecology and environmental conservation				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Explain the concepts of plant ecology and environmental conservation	U	2
2	Outline the structure and functions of population, community and ecosystems	U	2
3	Illustrate strategies of conservation for environment and biodiversity	An	2, 5
4	Critically assess the sustainable uses of natural resources	E	2, 5
5	Analyse the causes and effects of global warming and climate change	An	2, 5

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
		<b>Module 1 (15 Hours)</b>		
1	1.1	Introduction, definitions, autecology and synecology Population ecology: Characteristics of populations; Population growth - factors affecting population growth; concept of metapopulation- demes and dispersal, interdemic extinctions. Population interactions; Positive and negative interaction, Ecological consequence of overpopulations. Genecology - ecological amplitude, ecads, ecotypes, ecospecies, coenospecies, k-selection and r-selection populations.	5	1,2
	1.2	Community Ecology: Definition and concept, community characteristics; Community interdependence, Ecotone and Edge effect. Succession: Definition and reasons of succession. Classification of succession: Changes - autogenic and allogenic, primary and secondary, autotrophic and heterotrophic, concept of climax or stable communities. Characters used in the study of community structure – analytical and synthetic characters. Methods of study of community: quadrat, transect, sampling plots. Diversity indices - Simpson's index, Shannon-Weiner's index, Sorenson's similarity index.	5	1,2
	1.3	Ecosystem ecology: Ecosystem structure and function, ecosystem services. Energy flow and nutrient cycling in ecosystem. Primary and secondary productivity, trophic level, food chains, food webs, ecological pyramids. Habitat and Niche: Concept of habitat and niche; niche width and overlap; fundamental and realized niche; resource partitioning; character displacement. Biogeography: Major terrestrial biomes; theory of island biogeography; biogeographical zones of India	5	1,2
		<b>Module 2 (15 Hours)</b>		
2	2.1	Natural resources: Concept of resource, classification of natural resources-renewable and non-renewable resources. Factors influencing resource availability, distribution and uses.	5	4


		Major non-renewable resources- Forest, Soil, Water and Energy (fossil fuels): Sources, uses, over-exploitation, environmental impact. Ecological footprint and Carbon footprint.		
	2.2	Natural resource management and sustainable development: Sustainable use of natural resources – Resource management- meaning and concept. Sustainable forest management; Watershed management: Rain water harvesting and storage, recharging of ground water; Reclamation & Management of waste lands, soil conservation; Renewable and Alternative Energy Sources Sustainable development goals of the UN (Brief account)	5	4
	2.3	Environmental monitoring: Remote sensing and GIS- introduction, principle, application of remote sensing. GIS in natural resource mapping, disaster mapping and biodiversity conservation (brief account). Environmental impact assessment (EIA): EIA guidelines 1994, EIA methods; Environmental audit, ISO-14000 (Brief account).	3	3,4
	2.4	Environmental biotechnology and waste management: Bioremediation, Phytoremediation, bioaugmentation, biofilms, biofilters, bioscrubbers and trickling filters. Use of bioreactors in waste management.	2	4
	<b>3</b>	<b>Module 3 (15 Hours)</b>		
	3.1	Global warming: Causes and impact, green-house gases and their sources, acid rain, ozone layer depletion, sea level rise, global climate change, desertification and habitat loss. Effects of increased CO <sub>2</sub> on plants, carbon sequestration.	4	5
	3.2	Climate change: Drivers of climate change, <i>El-Nino</i> and <i>La Nina</i> phenomenon and its consequences. Implications on climate, oceans, agriculture, natural vegetation, wildlife and humans.	3	5
	3.3	International efforts to mitigate climate change: IPCC, UNFCCC, CoP, Paris Protocol, Kyoto Protocol, Vienna agreement, Montreal protocol (Brief account)	4	5
	3.4	Ecosystem resilience: Introduction, why resilience, resilience and stability of ecological systems, resilience of terrestrial ecosystems, regime shift, resilience and biodiversity in ecosystem management.	4	5

	<b>4</b>	<b>Module 4 (15 Hours)</b>		
	<b>4.1</b>	<p>Definition, importance, levels of biodiversity.          Concept of endemism, rare, endangered and threatened species (RET), key stone species.          Biodiversity hotspots</p> <p>Biodiversity prospecting and indigenous knowledge systems, community biodiversity registers.          Biodiversity as bio resources – use and values (consumptive and productive use values) of biodiversity.</p> <p><u>Activity</u>: One day field visit to study biodiversity</p>	8	3
	<b>4.2</b>	<p>Loss of biodiversity: Causes and rate of biodiversity loss; Extinction, causes of extinction - deforestation, habitat loss, industrialization, hunting and bio invasions; invasive species. Extinction through geological time scale: mass extinctions. Current extinction trends</p>	3	3
	<b>4.3</b>	<p>Conservation strategies: <i>In-situ</i> and <i>ex-situ</i> conservation method; Protected areas network, Wild life conservation projects. People's participation in biodiversity conservation (JFM).          Role of biotechnology in conservation of species.</p>	2	3
	<b>4.4</b>	<p>Conservation efforts: UNESCO- biosphere reserves, world heritage sites; IUCN and conservation, Red Data Book and categories.</p>	2	3
<b>5</b>		<b>Teacher specific course components</b>		

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b>          Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b>  <b>A. Continuous Comprehensive Assessment (CCA)</b>          Theory Total = 30 marks          Quiz, Test Papers, seminar</p>
	<p><b>B. End Semester Examination</b>          Theory Total = 70 marks, Duration 2 hrs          Part A (Short answer) – 10 out of 12 x 2 = 20 marks          Part B (Short essay) – 6 out of 9 x 5 = 30 marks          Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>

**References:**

1. Chaturvedi, P. (2003). Energy, environment and sustainable development. Concept Publishing Company.
2. Dash, M.C. (1993). Fundamentals of Ecology. Tata McGraw Hill.
3. Kumar, H. D. (2000). Modern Concepts of Ecology. Vikas Publishing House, New Delhi.
4. Kumar, U., & M. Asija (2006). Biodiversity: Principles and conservation. Agrobios India.
5. Misra, D. D. (2008). Fundamental concepts in Environmental Studies. S. Chand & Co. Ltd., New Delhi.
6. Misra, K.C. Manual of plant ecology. Oxford and IBH Pub. Com. P. Ltd.
7. Nayar, M.P. (1997). Biodiversity challenges in Kerala and science of conservation biology. *In*: P. Pushpangadan, K.S.S. Nair (Eds), Biodiversity of tropical forests the Kerala scenario. STEC, Kerala.
8. Odum, E.P. (1971). Fundamentals of Ecology. WB Sunders.
9. Panday, S.N. & S.P. Misra (2011). Environment and Ecology. Ane Books Pvt.Ltd. New Delhi
10. Sachs, J., Kroll, C., Lafortune, G., Fuller, G., & Woelm, F. (2022). Sustainable development report 2022. Cambridge University Press.
11. Santhra, S.C. (2004). Environmental Science. New Central Book Agency.
12. Sharma, P. D., & Sharma, P. D. (2012). *Ecology and environment*. Rastogi Publications.
13. Sulekha, Chendel. Plant Ecology and Soil. S Chand & Co. Ltd. New Delhi.
14. Thangavel, P., & Sridevi, G. (2016). Environmental Sustainability. Springer, India, Private.
15. Verma, M. K. (2022). Environment and Sustainable Development. British Library Cataloguing-in Publication Data.
16. Verma, P. S., & Agarwal, V. K. (2000). *Environmental Biology: Principles of Ecology*. S. Chand.
17. Xavier Savarimuthu, S. J., Rao, U., & Reynolds, M. F. (Eds.). (2021). Go Green for Environmental Sustainability: An Interdisciplinary Exploration of Theory and Applications. CRC Press.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b>					
<b>FYUGP SYLLABUS</b>						
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Conventional and Modern approaches in Plant Breeding</b>					
<b>Type of Course</b>	<b>DCE</b>					
<b>Course Code</b>	<b>M24BO7DCE401</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	The course deals with plant and crop improvement techniques.					
<b>Semester</b>	7	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		4	-	-	-	60
<b>Pre-requisites, if any</b>	Basic knowledge in plant reproduction and propagation					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Summarize the origin and scope of plant breeding along with the major research centers involved in plant breeding	U	3
2	Choose a proper plant breeding method for a crop improvement programme	A	3, 4
3	Explain the nuances of heterosis and inbreeding depression	U	3
4	Explore the importance and applications of plant genetic resources for food security and agriculture	A	5
5	Develop strategies for conserving the regional plant genetic resources	A	2, 3, 4
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Introduction, objectives and significance of plant breeding.	3	1
	1.2	National and International Centres of Plant breeding- ICAR, NBRI (National Botanical Research Institute), IRRI Philippines, IPGRI (International plant genetic resource institute, Rome). Plant breeding Stations in Kerala and their achievements – CPCRI, CTCRI, RRII.	6	1,2
	1.3	Germplasm: different types of germplasm, gene pool concept, genetic erosion, germplasm collections-requisites for a gene bank, genetic erosion in gene banks, constraints of gene bank. Germplasm conservation: - <i>in situ</i> and <i>ex-situ</i> conservation- seed banks, plant gene banks, shoot tip gene banks, cell and organ gene banks, DNA banks, germplasm evaluation- cataloguing-multiplication and distribution, germplasm utilization.	6	5
2	<b>Module 2 (15 Hours)</b>			
	2.1	Classes of seed- Basic nucleus seed, breeder seed, foundation seed, certified seed. Seed processing- Drying, grading, testing, treating, bagging and labeling. Seed certification – genetic purity, physical purity, germination, moisture content, freedom from weeds and diseases. National Seed Corporation, State Seed Certification Agencies, Activities of seed industry, Seed multiplication.	7	5
	2.2	Plant introduction: procedure of plant introduction - quarantine regulations, acclimatization, agencies of plant introduction in India, major achievements.	4	2,5
	2.3	Selection: History of selection, pureline selection, mass selection, pedigree selection, bulk method of selection, backcross method of selection procedure, applications, merits, demerits and achievements of each type.	4	1,2
3	<b>Module 3 (15 Hours)</b>			



	3.1	Hybridization: types, procedure, important achievements. Emasculation – different methods, consequences of hybridization.	4	5
	3.2	Heterosis and Inbreeding: Definitions. Inbreeding depression – effect of inbreeding – degree of inbreeding depression – homozygous and heterozygous balance. Heterosis and luxuriance – manifestation of heterosis. Genetic basis – dominance hypothesis-over dominance hypothesis – similarities and differences. Physiological basis of heterosis-mitochondrial complementation – fixation of heterosis. Recurrent selection types- simple-recurrent selection for general combining ability – for specific combining ability – reciprocal recurrent selection.	7	2,3
	3.3	Hybrids and synthetic varieties: Procedure – development of inbreds – methods for evaluation of inbreds–production of hybrid seeds- double cross and polycross hybrids. Role of cytoplasmic genetic male sterility and self-incompatibility in hybrid seed production	4	5
	<b>Module 4 (15 Hours)</b>			
	4.1	Mutation breeding: methods, applications and important achievements. Polyploidy breeding: methods and applications.	4	1
4	4.2	Ideotype breeding: Ideotype concepts, types, development of ideotypes, Steps in development–identification of traits for analysis determination value of traits, choice of traits for evaluation- ideotype breeding method limitations.	5	2
	4.3	Transgenic approaches for plant breeding Breeding for stress resistance - drought, mineral stresses, salinity, water stress - breeding approach- Problems.	6	4,5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment</b>	<b>MODE OF ASSESSMENT</b>


<b>Types</b>	<b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester Examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## REFERENCES

- Allard, R. W. 1999. *Principles of plant breeding*. John Wiley and Sons.
- Acquaah, G. 2009. *Principles of plant genetics and breeding*. John Wiley & Sons.
- Ahlawat, S. P., Bhatt, K. C., Semwal, D. P., Pradheep, K. and Dhariwal, O. P. 2022. Exploration and Collection of Plant Genetic Resources in India: Status and Priorities. *Indian Journal of Plant Genetic Resources*, 35(03), 117-123.
- Arora RK and Nayar, E. R. 1984. Wild Relatives of Crop Plants in India. NBPGR Monograph no. 7, NBPGR, New Delhi.
- Arora, R. K. and Pandey, A. 1996. Wild edible plants of India diversity, conservation and use, National Bureau of plant genetic resources. *New Delhi*.
- Chahal, G.S. and Gosal, S. S. 2002. *Principles and procedures of plant Breeding*. Narosa Publishing House. New Delhi.
- Hayward, M. D., Bosemark, N. O., and Romagosa, T. (Eds.). 2012. *Plant breeding: principles and prospects*. Springer Science & Business Media.
- ICRISAT (International Crops Research Institute for the Semi-Arid Tropics) Western and Central Africa Region. 2009. Protecting Biodiversity, Providing Options. BP 12404, Niamey, Niger: ICRISAT. 64 pp.
- IWANAGA, M. 1994. Role of International Organizations in Global Genetic Resources. In *Plant Genetic Resource Management in the Tropics: Proceedings of the 27th International Symposium on Tropical Agriculture Research, Tsukuba, Japan, August 25-26, 1993* (No. 2, p. 1). Japan International Research Center for Agricultural Sciences, Ministry of Agriculture, Forestry and Fisheries.
- Kirtikar, K R and Basu, B. D. 1991. *Indian Medicinal Plants*. Dehra Dun.
- Kurian, A. and Peter, K. V. 2007. *Commercial crops technology* (Vol. 8). New India Publishing.
- Mahour, K. 2016. Role of women in environment conservation. *Journal of Advanced Laboratory Research in Biology*, 7(1), 17-26.
- Paroda, R. S. and Arora, R. K. 1991. *Plant Genetic Resources Conservation and Management*, NBPGR, New Delhi.
- Peter, K.V. and Abraham, Z. 2007. *Biodiversity in Horticultural Crops* Vol.1, Daya Publishing House. New Delhi.
- Ram, M. 2014. *Plant breeding methods*. PHI Learning Pvt. Ltd.

16. Salgotra, R. K., and Chauhan, B. S. 2023. Genetic diversity, conservation, and utilization of plant genetic resources. *Genes*, 14(1), 174.
17. Singh, B. B., Neeta Singh and Kalyani Srinivasan (eds). 1996. *Principles and Procedures* in Germplasm Conservation, NBPGR. New Delhi
18. Singh, B. P. and Umesh Srivastava. 2004. *Plant Genetic Resources in Indian Perspective- Theory and Practice*, Directorate of Information and Publications of Agriculture, ICAR. New Delhi.
19. Singh, B. D. 2015. *Plant Breeding, principles and methods* (X Edn.). Kalyani publishers, New Delhi.
20. Singh, D. P., Singh, A. K. and Singh, A. 2021. *Plant breeding and cultivar development*. Academic Press.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Seed Technology</b>					
<b>Type of Course</b>	<b>DCE</b>					
<b>Course Code</b>	<b>M24BO7DCE402</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	This course is a comprehensive study of principles and application of seed science and technology. The course provides an understanding of the vital role in seed plays in agriculture, plant biology and sustainable development.					
<b>Semester</b>	7	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
<b>Pre-requisites, if any</b>	Knowledge in plant physiology and reproduction					

#### **COURSE OUTCOMES (CO)**

<b>CO No</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Explain the basics of seed biology and seed quality	U	3
2	Evaluate the quality of seeds using seed testing method	E	4
3	Describe the steps in seed processing and seed certification	U	3
4	Apply the role of biotechnology in seed development	A	3,5
5	Analyze seed marketing and trade	An	5
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 Hours)</b>			
	1.1	Definition of seed science and technology, scope; Heritage of seed technology and contribution of seed technologists towards the holistic development of modern science (interactive sessions)-	1	1
	1.2	Morphology and seed development: Seed Biology- Study of floral biology of monocots and dicots external and internal structures of monocot and dicot seeds; seed coat structure, different types of embryos, endosperm and cotyledons Seed development Physiology-Physiology of seed development and maturation; chemical composition, synthesis and accumulation of seed reserves, induction of desiccation tolerance, hormonal regulation of seed development Dormancy- definition, types, mechanisms, advantage, disadvantage, endogenous and exogenous factors regulating dormancy, role of phytochrome and PGR, genetic control of dormancy Seed deterioration- causes and factors affecting seed deterioration, Physiological, cytological and biochemical changes during seed storage and its implication in seed quality, methods to reduce seed deterioration <b>Activity:</b> Preparation of seed albums and identification	9	1
	1.3	Seed ripening and maturation process, Factors affecting seed setting. Seed germination -Seed germination; factors affecting germination; role of embryonic axis; growth hormones and enzyme activities, effect of age, size and position of seed on germination. Physiological processes during seed germination; seed respiration, breakdown of stored reserves in seeds, mobilization and interconversion pathways.	5	1
2	<b>Module 2 (15 Hours)</b>			
	2.1	Seed viability and longevity, pre and post-harvest factors affecting seed viability; seed aging; physiology of seed deterioration; lipid peroxidation and other viability theories; means to prolong seed viability; mechanism of desiccation sensitivity and recalcitrance with respect to seed. Varietal Seed vigour and its concept, vigour test methods, factors affecting seed vigour, physiological basis of seed vigour in relation to crop performance and	7	2

		<p>yield. Seed invigoration and its physiological and molecular control</p> <p>Methods to prolong seed viability, Procedures involved in seed testing, (Sampling, physical purity, germination, seed moisture, viability, health, vigour and determination of genuineness), Devices and tools used in seed testing. ISTA, AOSA and its role in seed testing.</p> <p><b>Activity:</b></p> <ul style="list-style-type: none"> <li>● Seed viability testing method (Tetrazolium),</li> <li>● Seed germination test (Between paper/Top of paper method)</li> <li>● Visit to seed production Unit</li> </ul>		
	2.2	<p>Seed storage: general principles, Seed drying and storage; drying methods-importance and factors affecting it, changes during storage, concepts and significance of moisture equilibrium, methods of maintaining safe seed moisture content. Methods to minimize the loss of seed vigour and viability; factors influencing storage losses. Methods of seed storage – modified atmospheric storage – ultra dry storage – vacuum storage – cryopreservation – germplasm storage – gene banks – NBPGR, IPGRI and National seed storage laboratory, Measures for pest and disease control during storage, Seed Bank</p> <p>Seed treatments-methods of seed treatment, seed treating formulations and equipments, Biological seed treatments, seed disinfestations, identification of treated seeds; Packaging: principles, practices and materials; bagging and labeling</p>	8	3
	<b>Module 3 (15 Hours)</b>			
3	3.1	<p>Seed production through crop improvement and breeding, hybrid seeds (Maize, Sunflower), Causes of varietal deterioration and maintenance of genetic purity during seed production</p> <p>Seed quality control – Definition of seed and its quality-concept and objectives; regulatory mechanisms – Seed Act (1966) – Seed Rules (1968), statutory bodies– Central Seed Committee – Central Seed Certification Board, DUS test. Detection of genetically modified seeds. Identification through Grow Out Test and Electrophoresis.</p> <p>Seed certification –objectives; general and specific crop standards, field and seed standards; seed certification agency – role of certification agencies, phases of seed certification; Brief account on role and working of CSTL. Seed processing technologies( seed cleaning and equipment in seed processing)</p>	8	3
	3.2	Seed quality enhancement	7	3,5

		Seed priming: types of priming technology, biochemical and molecular changes associated, pre-germination, film coating and pelleting, seed tapes, seed mats, seed colouring, biopriming		
4	<b>Module 4 (15 Hours)</b>			
	4.1	Impact of genetic engineering , Genetic purity analysis of seeds, Use of Molecular markers, GMOs in seed production, Detection of genetically modified crops,; Transgene contamination in non-GM crops; GM crops and organic seed production.; Application of tissue culture in genetic conservation-Embryo culture, Embryo rescue, pollen and anther culture	8	4
	4.2	Seed marketing: structure and organization, sales generation activities, promotional media.; Factors affecting seed marketing.	4	3,5
	4.3	Seed trade regulations, IPR in seed technology	3	3,5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar
	<b>B. End Semester Examination</b> Theory Total = 70 marks, Duration 2 hrs Part A (Short answer) – 10 out of 12 x 2 = 20 marks Part B (Short essay) – 6 out of 9 x 5 = 30 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks

## References


1. Agrawal RL. 2019. Seed Technology. Oxford & IBH Publishing Company Pvt. Ltd., New Delhi.

2. Joshi AK and Singh BD. 2004. Seed Science and Technology. Kalyani Publishers, New Delhi.
3. Kulkarni GN. 2011. Principles of Seed Technology. Kalyani Publishers, New Delhi
4. Baskin C and Baskin JM. 2014. Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination. Academic Press, Cambridge, UK.
5. Bewley J and Black M. 1994. Physiology of Development and Germination. Springer, New York.
6. Bewley JD, Bradford KJ, Hilhorst HWM and Nanogaki H. 2013. Seeds: Physiology of Development, Germination and Dormancy. Springer, New York.
7. Adkins SW, Ashmore SE and Navi SC. 2007. Seeds: Biology, Development and Ecology. CABInternational, Oxford shire, UK.
8. Anon. 2016. Manual of Seed Certification Procedures. Directorate of Seed Certification, Coimbatore, Tamil Nadu.
9. Chakrabarthy SK. 2010. Seed Production and Quality Control. Kalyani Publishers, New Delhi.

#### **Suggested Readings**

1. Mishra DK, Khare D, Bhale MS and Koutu GK. 2011. Handbook of Seed Certification. Agrobios, Jodhpur, Rajasthan
2. Sharma P. 2008. Seed Legislation. Gene-tech Book Publishers, New Delhi.
3. Chakrabarthy SK. 2010. Seed Production and Quality Control. Kalyani Publishers. New Delhi.
4. Chalam GV Singh A and Douglas JE. 1967. Seed Testing Manual. ICAR and United States Agency for International Development, New Delhi



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Agroecology</b>				
<b>Type of Course</b>	<b>DCE</b>				
<b>Course Code</b>	<b>M24BO7DCE403</b>				
<b>Course Level</b>	<b>400</b>				
<b>Course Summary</b>	This course provides a comprehensive exploration of the principles and applications of agroecology, offering undergraduate botany students a foundational understanding of how ecological processes can be strategically applied to agricultural systems. As the global agricultural landscape evolves, agroecology emerges as a transformative approach that integrates ecological principles with sustainable farming practices.				
<b>Semester</b>	7	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		4	-	-	60
<b>Pre-requisites, if any</b>	Knowledge in plant sciences and ecology				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Recognize the foundations of Agroecology	U	2
2	Apply Agroecological principles to Agriculture	A	2
3	Employ sustainable soil and crop management practices	A	2,5
4	Apply both cognitive understanding and practical skills in integrated livestock and pest management for sustainable agriculture	A	2,4
5	Analyze and promote sustainable agricultural practices	An	2,5
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Introduction to Agroecology- Definition and scope, historical development and evolution of agroecology,	2	1,5
		Distinctive features of agroecology as a discipline, ecological, Social and economic benefits, Sustainability in agriculture	4	2
	1.2	Basic principles and concepts- Agroecological Principles and elements and their implications in Agricultural systems.	4	2
	1.3	Soil Management for Sustainable Agriculture- Soil health and sustainability, importance of soil as a living ecosystem. Soil structure, texture and composition. Erosion control methods, cover cropping and mulching, contour plowing and terracing.	5	3
2	<b>Module 2 (15 hours)</b>			
	2.1	Crop Diversity and Rotation- Types and benefits of cover crops, incorporating cover crops in rotation, improving soil health and structure, Benefits of crop rotation	5	2,3,5
	2.2	Agroforestry- Introduction to Agroforestry, Principles of agroforestry, Alley cropping, wind breaks and integrating trees and crops for mutual benefits, Biodiversity enhancement, carbon sequestration and climate resilient farming, Economic and social benefits	10	2,5
	<b>Module 3 (15 Hours)</b>			
3	3.1	Water Management in Agriculture- Importance of water in agriculture, Role of water in plant growth and development. Efficient Irrigation techniques- Drip irrigation, sprinkler and furrow irrigation, Water conservation practices in irrigation. Rain water harvesting techniques, sustainable	7	2,3,5

		use of water resources		
	3.2	Livestock Integration in Agroecosystems- Silvio pasture and agroforestry systems with livestock, Grazing and mixed farming practices, grazing management for optimal land use	5	4,5
	3.3	Balancing crop and livestock systems, Interdependence between crops and livestock, Nutrient cycling and Waste utilization	3	4,5
	<b>Module 4 (15 Hours)</b>			
<b>4</b>	4.1	Environmental impact assessment of agricultural practices, mitigation strategies for minimizing negative effects	4	5
	4.2	Ensuring food security- understanding the ecological footprints of different farming systems	4	1,5
	4.3	Social and economic aspects of sustainable agriculture- Socioeconomic impact of agricultural practices, community engagement and involvement of communities in sustainable agriculture.	5	5
	4.4	Ethical values and practices involved in agriculture	2	5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, group discussions, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 30 marks Quiz, Test Papers, seminar

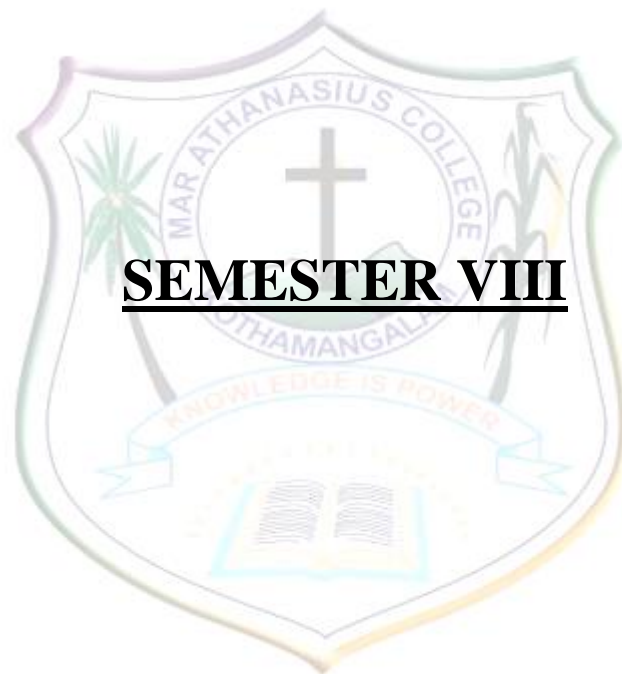
	<p><b>B. End Semester Examination</b>  Theory Total = 70 marks, Duration 2 hrs  Part A (Short answer) – 10 out of 12 x 2 = 20 marks  Part B (Short essay) – 6 out of 9 x 5 = 30 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks</p>
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## References


1. Agroecology: The ecology of sustainable food systems, Stephen R Gliessman
2. Agroecology: A transdisciplinary participatory and action oriented approach edited by Ernesto Mendez, Christopher M Bacon, Roseann Cohen.
3. Agroecology in Action: extending alternative agriculture through social
4. Temegne Nono, Carine & Ngome, Ajebesone & Paul Agendia, Atabong & Youmbi, Emmanuel. (2021). Agroecology for Agricultural Soil Management. 10.1007/978-981-16-3207-5\_9.
5. Jose, S. Agroforestry for ecosystem services and environmental benefits: an overview. *Agroforest Syst* **76**, 1–10 (2009). <https://doi.org/10.1007/s10457-009-9229-7>
6. Bullock, D. G. (1992). Crop rotation. *Critical reviews in plant sciences*, 11(4), 309-326.
7. Naik, S. K. (2019). Conservation Agriculture: A potential approach for carbon sequestration and climate resilient farming. *Conservation Agriculture for Climate Resilient Farming & Doubling Farmers' Income*, 246p. ICAR Research Complex for Eastern Region, Patna Training Manual No.
8. Reddy, P. P. (2015). *Climate resilient agriculture for ensuring Food security* (Vol. 373). New Delhi: Springer India.
9. Torquebiau, E. (1990). Introduction to the concepts of agroforestry. *Introduction to the concepts of agroforestry.*, (59).
10. Nair, PK Ramachandran, B. Mohan Kumar, Vimala D. Nair, PK Ramachandran Nair, B. Mohan Kumar, and Vimala D. Nair. "Definition and concepts of agroforestry." *An introduction to agroforestry: Four decades of scientific developments* (2021): 21-28., B. Mohan Kumar, Vimala D. Nair, PK Ramachandran Nair, B. Mohan Kumar, and Vimala D. Nair. "Definition and concepts of agroforestry." *An introduction to agroforestry: Four decades of scientific developments* (2021): 21-28.
11. Nair, PK Ramachandran, B. Mohan Kumar, Vimala D. Nair, PK Ramachandran Nair, B. Mohan Kumar, and Vimala D. Nair. "Definition and concepts of agroforestry." *An introduction to agroforestry: Four decades of scientific developments* (2021): 21-28.
12. Majumdar, D. K. (2001). *Irrigation water management: principles and practice*. PHI Learning Pvt. Ltd..
13. Bonaudo, T., Bendahan, A. B., Sabatier, R., Ryschawy, J., Bellon, S., Leger, F., ... & Tichit, M. (2014). Agroecological principles for the redesign of integrated crop–livestock systems. *European Journal of Agronomy*, 57, 43-51.

14. Soussana, J. F., Tichit, M., Lecomte, P., & Dumont, B. (2015). Agroecology: integration with livestock. FAO.
15. Martin, G., Moraine, M., Ryschawy, J., Magne, M. A., Asai, M., Sarthou, J. P., ... & Therond, O. (2016). Crop–livestock integration beyond the farm level: a review. *Agronomy for Sustainable Development*, 36(3), 53.
16. Altieri, M. A. (2002). Agroecology: the science of natural resource management for poor farmers in marginal environments. *Agriculture, ecosystems & environment*, 93(1-3), 1-24.
17. Mondal, S., & Palit, D. (2021). Agroecology for sustainable food system and footprint mitigation. *Agroecological Footprints Management for Sustainable Food System*, 69-114.
18. Gliessman, S., & Tittonell, P. (2015). Agroecology for food security and nutrition. *Agroecology and Sustainable Food Systems*, 39(2), 131-133.





## **SEMESTER VIII**

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b>				
<b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Modern Trends in Archegoniates</b>				
<b>Type of Course</b>	<b>DCC</b>				
<b>Course Code</b>	<b>M24BO8DCC400</b>				
<b>Course Level</b>	<b>400</b>				
<b>Course Summary</b>	This course enables students to identify and compare archegoniates, emphasizing their practical applications. It covers habitat variation, morphological diversity, reproductive behavior, and economic significance of archegoniates. Upon completion of the course, students will be able to classify archegoniates based on morphological traits, compare evolutionary trends and ecological significance, investigate diversity, and construct artificial ecosystems for conservation. Additionally, the course exposes students to current research areas, fostering interest in the field.				
<b>Semester</b>	8	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		3	-	1	-
<b>Pre-requisites, if any</b>	Basic knowledge about morphology and reproduction of archegoniates				

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PO No
1	Recognize the habitat variation, morphological diversity, and reproductive behaviour of archegoniates.	U, S	1,2,3
2	Identify archegoniates and classify them based on morphological and evolutionary characters.	An	1
3	Describe the economic and ecological significance of archegoniates and recognize and the need for conserving them	K, U	2
4	Summarize the diversity and distributions of prehistoric archegoniate flora	E	2
5	Appraise the applications of archegoniates across different fields and stay updated on the latest developments.	I, Ap, S	4,5

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Introduction- Salient features, classification by Goffinet <i>et al.</i> 2008 Comparative account of gametophyte, sporophyte, their inter relationship and spore dispersal mechanisms of the following phyla; Marchantiophyta, Bryophyta and Anthocerotophyta.	6	1,2
	1.2	Water relations in Bryophytes, Absorption, Conduction, xerophytic adaptation, drought tolerance, desiccation and rehydration, ectohydric, endohydric and myxohydric bryophytes. Ecology of bryophytes- habit, habitat, associated vegetation; Aquatic ( <i>Riccia fluitans</i> ), Terrestrial ( <i>Hyophila</i> , <i>Bryum</i> ), Epiphytic ( <i>Porella</i> ) Epiphyllous ( <i>Lejeunea</i> ). Role of bryophytes in ecosystem dynamics.	4	3
	1.3	Importance of bryophytes- medicinal, horticultural, antimicrobial, antifungal, active constituents, pollution monitoring (IAP), peat moss. Conservation of bryophytes- the need for conservation.	3	3
	1.4	Recent advances in the field of bryology- molecular studies, chemical constituents and physiological ecology, Bryological Research Centres of the world; major Herbaria.	2	5
2	<b>Module 2 (15 hours)</b>			
	2.1	Introduction, general characters, Trends, and concepts in classification of pteridophytes with emphasis on PPG 1. (brief study)	2	1,2
	2.2	Life Cycle of Pteridophytes-alternation of generations; apogamy, apospory, agamospermy and parthenogenesis. Stelar and soral evolution in pteridophytes.	2	1
	2.3	Structural organization of sporophyte and gametophyte (development of sex organs not necessary) of the following type with special reference to stelar structure, heterospory and seed habit. Psilopsida, Psilotopsida, Lycopsidea, Sphenopsida, Pteropsida (Eusporangiatae and Leptosporangiatae)	5	1,2



	2.4	Habitat ecology-Hydrophytes, Halophytes, Epiphytes, Xerophytes, Sciophytes, Climbers, Filmy Ferns, Tree Ferns, Saprophytes – Adaptation of Pteridophytes-RET Pteridophytes, Conservation.	3	3
	2.5	Cytology of pteridophytes-polyploidy and high chromosome number-origin of polyploids cytology and reproduction. Brief account of recent developments in molecular phylogenetics and DNA barcoding in pteridophytes. Applied Pteridology – horticulture – Food – Medicinal – Biofertilizer – Weeds (aquatic and terrestrial) – Ecological Indicators – Pollution amelioration.	3	2,3
	<b>Module 3 (15 hours)</b>			
	3.1	Introduction, general characters, evolutionary significance. Origin of seed plants: -Evolution of pollen and seed -the key reproductive evolutionary mechanisms for Life on Land. Relationships among gymnosperms - molecular phylogeny Distribution of living gymnosperms in India.	3	1,2
<b>3</b>	3.2	Study the Morphological and Applied Aspects of gymnosperms Cycadales - Ginkgoalesclade (general account on morphology) Coniferales -Pinaceae, Cupressaceae, Taxaceae, Podocarpaceae, Araucariaceae (general account on morphology) Gnetales: Gnetum (general account on morphology).	4	1,2,5
	3.3	Brief study of habit, morphology and reproductive characters of <i>Welwitschia mirabilis</i> Pollination strategies in gymnosperms Vascular system of gymnosperms (give emphasis to wood architecture)	2	1
	3.4	The ecological and economic importance of gymnosperms. Conservation of gymnosperms	2	3

	3.5	Introduction, fossil types & technique of study. Indian contribution to paleobotany Fossil plants Study of the following types; Fossil bryophytes: <i>Naiadita lanceolata</i> Fossil pteridophyte: <i>Rhynia</i> Fossil gymnosperms: <i>Williamsonia</i>	4	4
<b>Module 4 Practical (30 hours)</b>				
<b>4</b>	4.1	Morphological and Anatomical studies on Bryophytes: <i>Targionia, Cyathodium, Marchantia, Lunularia, Dumortiera, Reboulia, Pallavicinia, Fossombronia, Porella, Notothylas, Bryum.</i> Morphological and Anatomical studies on Pteridophytes: <i>Psilotum, Lycopodium, Isoetes</i> (no collection), <i>Equisetum, Angiopteris, Ophioglossum, Osmunda, Marsilea, Salvinia, Azolla, Lygodium, Acrostichum, Gleichenia, Adiantum, Polypodium and Asplenium.</i>	15	1,2
	4.2	Study of the morphology and anatomy of vegetative and reproductive parts of <i>Zamia, Cupressus, Podocarpus, Agathis, Araucaria</i> and <i>Gnetum</i> .  Study of the fossil plants mentioned in the syllabus with the help of specimens or permanent slides or photographs.	5	1,2
	4.3	<b>Activity:</b> Conduct a field survey to familiarize various archegoniate flora (bryophytes, pteridophytes and gymnosperms) and submit a report with geo-tagged photos related to their diversity.	10	1,2,3
<b>5</b>	<b>Teacher specific course content</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
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<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b></p> <p><b>A. Continuous Comprehensive Assessment (CCA)</b>  Theory Total = 25 marks  Quiz, Test Papers, seminar  Practical Total = 15 marks  Lab performance, record, field report etc.</p>
	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>


## References

1. Alam, A. (2019). *Textbook of Bryophyta*. Dreamtech Press.
2. Arnold, H.N. (1967). *Introduction to Paleobotany*. Tata Mc Graw- Hill, New Delhi.
3. Beck, C.B. (1988). *Origin and Evolution of Gymnosperms*, Columbia University Press, New York.
4. Biswas & John, B.M. (2004). *Gymnosperms*, Naresa Publishing house.
5. Bower, F.O. (1935). *Primitive Land Plants*, Cambridge, London.
6. Chopra, R.N. and Kumar, P. K. (1988). *Biology of Bryophytes*, Wiley Eastern Ltd, New Delhi.
7. Coutler, J.M., & Chamberlain, C. J. (1958). *Morphology of Gymnosperms*. Central Book Depot, Allahabad.
8. Dutta, S.C. (1991). *An Introduction to Gymnosperms*. Kalyan Publishing Co. New Delhi.
9. Kashyap, S. R. (1932). *Liverworts of Western Himalayas and the Punjab plains (Vol. I & II)*. Research.
10. Maarten, J. M. Christenhusz, Mark, & Chase, W. (2014). *Trends and concepts in fern classification, Annals of Botany*, Volume 113, Issue 4, Pages 571–594, <https://doi.org/10.1093/aob/mct299>
11. Mamatha, Rao. (2009), *Microbes and Non flowering plants- impact and application*. Ane Books Pvt Ltd.
12. PPG- I, (2016). *A community-derived classification for extant lycophytes and ferns. Journal of Systematics and Evolution*, vol. 54, no. 6, pp. 563-603. <http://dx.doi.org/10.1111/jse.12229>
13. Rasheed, A. (1999). *An Introduction to Pteridophyta*. Vikas Publishing House, New Delhi.
14. Rasheed, A. (2000). *An Introduction to Bryophyta*. Vikas Publishing House, New Delhi.
15. Sharma O.P. (2016). *Series on Diversity of Microbes and Cryptogams – Pteridophyta*. Tata McGraw Hill Education Private Limited, New Delhi.
16. Sharma, O.P. (2014). *Bryophyta*. McGraw Hill education (India) Pvt Ltd
17. Singh, V., Pande, P. C., & Jain, D. K. (2010). *Diversity of microbes and cryptogams*. Rastogi Publication, India.
18. Taylor, T.N. Taylor, E.L., & Krings, M. (2009). *Paleobotany—The Biology and Evolution of Fossil Plants*. Burlington: Academic Press.

19. Timell, T.L. (1986). *Compression Wood in Gymnosperms*:Springer-Verlag Berlin Heidelberg New York Tokyo.
20. Vanderpoorten, A., & Goffinet, B. (2009) *Introduction to Bryophytes*. Cambridge University Press.
21. Vashishta, B. R., Sinha, A.K., & Kumar, A. (2003). *Bryophyta*. S Chand & Co. Ltd.
22. Vashista, B. R, (1993). *Bryophyta*. S Chand & Co., New Delhi.
23. Vashista, B. R, (1993). *Gymnosperms*. S Chand & Co., New Delhi.
24. Vashista, B. R, (1993). *Pteridophyta*. S Chand & Co., New Delhi.

#### **Websites**

<http://www.artdata.slu.se/guest/SSCBryo/SSCBryo.html>  
<http://www.northernontarioflora.ca/links.cfm?val=bryophytes>  
<http://bryophytes.plant.siu.edu/>  
<http://worldofmosses.com/>  
<http://www.unomaha.edu/~abls/>  
<http://www.anbg.gov.au/bryophyte/index.html>  
<http://www.bryoecol.mtu.edu/>  
<http://www.mobot.org/MOBOT/tropicos/most/Glossary/glosefr.html>  
[http://www.fairhavenbryology.com/Master\\_Page.html](http://www.fairhavenbryology.com/Master_Page.html)  
<http://www.mygarden.ws/fernlinks.htm>  
<http://www.anbg.gov.au/fern/index.html>  
<http://www.bioimages.org.uk/HTML/T77.HTM>  
[http://botany.csd.tamu.edu/FLORA/gallery/gallery\\_query.htm](http://botany.csd.tamu.edu/FLORA/gallery/gallery_query.htm)  
<http://homepages.caverock.net.nz/~bj/fern/>  
<http://www.home.aone.net.au/~byzantium/ferns/>  
<http://www.northernontarioflora.ca/links.cfm?val=pteridophytes>  
[http://www.fiu.edu/~chusb001/giant\\_equisetum.html](http://www.fiu.edu/~chusb001/giant_equisetum.html)  
<http://www.mygarden.ws/fernlinks.htm>

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Modern Trends in Plant Systematics</b>					
<b>Type of Course</b>	<b>DCC</b>					
<b>Course Code</b>	<b>M24BO8DCC401</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	The morphological characters alone should not be considered in systematic classification of plants. Modern trends help plant taxonomists to look for more precise techniques in order to understand the relation between the genera and families. Complete knowledge of taxonomy is possible with the principles of various disciplines like cytology, palynology, phenology, biochemistry and numerical taxonomy. These have been found to be useful in solving some of the taxonomical problems by providing additional characters.					
<b>Semester</b>	8	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Knowledge in fundamentals of plant morphology and taxonomy					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PO No
1	Explain the conceptual basis of plant classification and the concept of family, genus and species and the taxonomic diversity within species	U	1,2
2	Develop working skills in modern techniques in plant systematics	A	2,3
3	Choose appropriate tools of modern systematics for plant identification	A	3
4	Determine evolutionary relationship between a group of species using molecular taxonomic tools and techniques	A	3
5	Construct phylogenetic trees based on molecular systematic data	C	3, 5
*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15)</b>			
	1.1	Definition, Concepts and theories of classification and biosystematics. History and theories of classification – Theophrastus, Linnaean and post Linnaean era- Phylogenetic classification - Angiosperm Phylogeny Group (APG)- Detailed Account. <b>Learning Activity:</b> Students should submit a review on plant classification- past to present.	7	1
	1.2	Hierarchy in classification. Concept of Family, Genera, Species, Subspecies and other infra-specific categories. Species concepts: Typological, Nominalistic and Biological species concepts (in plant perceptive).	6	1
	1.3	The new global taxonomy initiatives: Systematic Agenda-2020- Missions.	3	3
2	<b>Module 2 (15)</b>			
	2.1	Chemotaxonomy- Classification based on phytochemicals- phenolics, alkaloids, terpenoids and nonprotein amino acids. Serology and Taxonomy. Scope and limitations.	5	3
	2.2	Cytotaxonomy – chromosome number, chromosome size, chromosome banding and behaviour of chromosomes during division.	5	3
	2.3	Palynotaxonomy- Pollen morphological characters and their significance in taxonomy and evolution- Polarity, symmetry, NPC of pollen, exine stratification, excrescences, L/O pattern. PollenAtlas .	5	3
3	<b>Module 3 (15 hours)</b>			
	3.1	Stereo Microscopes, Scanning Electron Microscopy, Transmission Electron Microscopy, Microphotography (Image analyser software) for micromorphological studies - Trichomes and seed morphology	5	2,3
	3.2	Numerical Taxonomy (Phenetics): Theory and principles- Operational Taxonomic Unit (OTU) Cluster analysis; UPGMA Methods; NTSYS, Applications, Merits and Demerits, Cluster analysis, Dendrogram.	4	2,3
	3.3	Molecular taxonomy - concepts, scope and limitations, Plant DNA barcoding- Molecular markers- isozymes, AFLP, Internal Transcribed	6	3,4

		Spacer (ITS), rbcL, matK. NCBI, Similarity search tools- BLAST, FASTA, Cladistics (Monophyletic, polyphyletic and paraphyletic groups), Phylogenetic tree construction, methods and tools- MEGA, PHYLIP. Interpreting data. Detailed study.		
<b>4</b>	<b>Module 4 Practical (30 hours)</b>			
	4.1	Prepare and submit a review on plant classification past to present	5	1
	4.2	Students should familiarise themselves with the application of chemical data from TLC/ HPTLC/HPLC/GC for taxonomy	5	3
	4.3	Semipermanent pollen preparations by acetolysis method /any other alternative methods and study of different pollen morphotypes	5	3
	4.4	Study of plant surface attributes (trichomes /spines etc) / pollen characters with the help of stereo microscopes / SEM	5	3
	4.5	Practical based on numerical taxonomy- Construct OTU tables examining morphological characters of selected plants	5	4,5
	4.6	Construct phylogenetic trees using MEGA/PHYLIP or Sequence similarity searching through NCBI BLAST	5	4,5
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs

	Record 10 marks, Examination 25 marks
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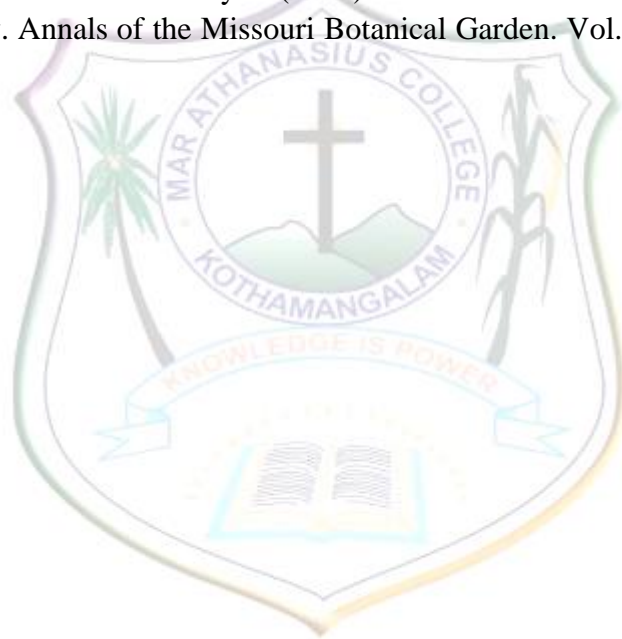
## References


1. Ash, A et al (1999). Manual of Leaf Architecture' by leaf Architecture Working Group - morphological description and categorization of dicotyledonous and net-veined monocotyledonous angiosperms by Leaf Architecture Working Group. 65p.
2. Bell, A.D (1991). Plant form- An illustrated guide to Flowering plant morphology. Oxford university press, NewYork, Tokyo
3. Blackmore S & Cutler D. (1996). Systematics Agenda 2000: the challenge for Europe. London: Linnean Society.
4. Carvalho & Maria, M & Lapenta, A & P.S, Machado. (2003). Functional classification of esterases from leaves of *Aspidosperma Polyneuron* M. Arg. (Apocynaceae). Genetics and Molecular Biology. 26.
5. Daly, M., Herendeen, P. S., Guralnick, R.P, Westneat, M.W & L. McDade (2012). Systematics Agenda 2020: The Mission Evolves. Syst Biol. 2012 Jul; 61(4): 549–552
6. Endress, P.K et al. (2000). Systematic plant morphology and anatomy: 50 years of progress
7. Felsenstein, J. (Ed.) (1983). Numerical Taxonomy. NATO ASI Subseries G
8. Guerra, M (2008). Chromosome numbers in plant cytotaxonomy: concepts and implications. Cytogenet Genome Res 120 (3-4): 339–350.
9. Hewitt, G.M, Johnston, A.W.B & J. P.W. Young (Eds..) (1991). Molecular Techniques in Taxonomy. NATO ASI Subseries H, Vol57
10. Hickey, L.J. & D.W. Taylor (1991). The leaf architecture of *Ticodendron* and the application of foliar characters in discerning its relationships. Annals of the Missouri Botanical Garden 105–130.
11. Hickey, L.J. (1973). Classification of the architecture of dicotyledonous leaves. American Journal of Botany 60(1): 17–33.
12. James Rohlf, F. (2009). NTSYSpc Numerical Taxonomy and Multivariate Analysis System Version 2.2, Applied Biostatistics Inc., 10 Inwood Road, Port Jefferson, New York. ISBN: 0-925031-31-3.
13. Taxon, Vol. 49, No. 3, Golden Jubilee Part 1 , pp. 401-434 (34 pages)

## SUGGESTED READINGS



1. Mohammad, Q *et al* (2022). Pollen characters and their evolutionary and taxonomic significance: Using light and confocal laser scanning microscope to study diverse plant pollen taxa from central India. *Palynology*.
2. Prance GT. (1995). Systematics, conservation and sustainable development. *Biodiv. Conserv*4:490–500.
3. Ramawat, K.G. (Eds) (2019). *Biodiversity and Chemotaxonomy (Sustainable Development and Biodiversity Book 24)*.Springer
4. Santanu, D & Bandyopadhyay, S (2009). *Molecular Taxonomy: An Approach Based on Molecular Markers*. *Science and culture*. 74. 397-496.
5. Simpson, M. G. (2019). *Plant systematics*. Academic press, London. pp.566–568
6. Sneath, P.H.A & Sokal, R.R (1973). *Numerical Taxonomy: The Principles and Practice of Numerical Classification*. W H Freeman & Co; First Edition
7. *Systematics Agenda* (2000). *Systematics agenda 2000: charting the biosphere*. Technical Report. New York: Systematics Agenda; 1994. pp. 1–34
8. Walker, J.W & J. A. Doyle (1975). The Bases of Angiosperm Phylogeny: *Palynology*. *Annals of the Missouri Botanical Garden*. Vol. 62, No. 3, pp. 664-723 (60 pages).



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Computational Biology and Omics in Plant Sciences</b>					
<b>Type of Course</b>	<b>DCE</b>					
<b>Course Code</b>	<b>M24BO8DCE400</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	The "Computational Biology and Omics in Plant sciences" course explores drug design using computational methods, covering drug discovery processes, molecular targets, and predictive modeling. It includes genomics, focusing on sequencing, mapping, and key genome projects. The course also examines gene expression, protein analysis, and metabolomics, emphasizing techniques like microarrays, mass spectrometry, and NMR spectroscopy for biological research.					
<b>Semester</b>	8	Credits				Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others	
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Basic understanding of molecular biology tools used in Bioinformatics					

### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Critically evaluate genomic, proteomic, and metabolomic data to draw conclusions about biological phenomena and drug targets	E	3,4
2	Evaluate the effectiveness of computational methods in drug design for lead compound identification and optimization.	E	2,3,4

3	Apply computational approaches in drug design, including target identification, lead compound identification, molecular docking, and virtual screening.	A	4,5,6
4	Analyze genomic data, including genetic mapping, molecular markers, and transcriptome expression profiling, to understand biological processes.	A	4,6
5	Utilize bioinformatics tools for binding site prediction, lead optimization, QSAR modeling, and ADME predictions.	A	2,3,4,6
<b>*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)</b>			

### COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hrs)</b>			
	1.1	<p><b>COMPUTER AIDED DRUG DESIGNING</b> Diseases and their causes-molecular basis of diseases. Immunology- cells and molecules in immune system, antigens &amp; antibodies, immune response, vaccines.</p> <p>Molecular targets, Characteristics of a drug compound, mechanism of drug action, small molecular drugs, peptide drugs. Traditional approaches in drug discovery, serendipity, high throughput screening, and drug discovery in post-genomic era.</p> <p>Drug discovery pipeline, pre-clinical &amp; clinical studies, IP issues in Drug Design, drug licensing in India.</p>	8	2,3,5
	1.2	<p>Computational approaches in Drug Design: Applications of bioinformatics in target identification &amp; validation, binding site prediction.</p> <p>Lead compound identification: Structure-based &amp; ligand based approaches;</p> <p>Molecular docking- algorithms and scoring functions; Virtual screening- combinatorial chemistry and ligand databases; Design of ligands for known target sites- de novo techniques.</p>	7	2,3

		Lead optimization. Pharmacophore ligand based & target based. QSAR - molecular descriptors, bio-activity predictions. ADME Predictions. Introduction to Software: Autodock, Discovery studio etc.		
2	<b>Module 2 (15 hrs)</b>			
	2.1	<b>Genomics</b> <b>Structural genomics</b> Basic steps in genome sequencing . Shot gun sequencing of small genomes . Genome mapping: Genetic mapping and physical mapping Cytogenetic and linkage map (brief study only) Molecular markers – RFLP, RAPD, AFLP, SSLP, SNP Construction of linkage maps using molecular markers – E.g., RFLP maps Genome projects:HGP,TAIR	10	1,4
	2.2	<b>Functional genomics</b> Transcriptome expression profiling (mRNA profiling) = Gene expression analysis using dot blotting and microarrays'. Fabrication of microarrays – spotted arrays, <i>in situ</i> synthesis Chromatin immunoprecipitation (ChIP) and its applications	5	1,4
3	<b>Module 3 (15 Hours)</b>			
	3.1	<b>Proteomics</b> Proteome, proteomics. Separation and identification of cellular proteins by 2D gel electrophoresis and mass spectrometry. Protein expression analysis using Protein microarray, protein localization using GFP · other applications of GFP.	5	4
	3.2	<b>Metabolomics</b> Overview of metabolomics: Definition, scope, and significance in biological research Principles of metabolite identification and quantification Techniques in metabolomics: Nuclear Magnetic Resonance (NMR) spectroscopy, Mass Spectrometry (MS), and chromatography	5	4
4	<b>Module 4 (Practical:30 hrs)</b>			

	4.1	Visit to Bioinformatics lab Molecular Docking Using PyRX and Discovery studio	20	2,5
	4.2	Hands-on experience in utilizing genomic tools and methods to address biological questions.	10	2,5
<b>5</b>	<b>Teacher specific course components</b>			


<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.
	<b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks

## References

1. C W Sensen (2002). *Genomics and Bioinformatics*. Wiley – VCH.
2. Charifson, P. S. (1997). *Practical application of computer-aided drug design*. Marcel Dekker, Inc.
3. Claverie, J. M., & Notredame, C. (2011). *Bioinformatics for dummies*. John Wiley & Sons.
4. D Peter Snustad, Michael J Simmons (2010). *Principles of genetics* (V Edn). John Wiley and Sons.
5. Eidhammer, I., Jonassen, I., & Taylor, W. R. (2004). *Protein Bioinformatics: An algorithmic approach to sequence and structure analysis* (pp. 3-23). J. Wiley & Sons
6. Higgins, D., & Taylor, W. (2000). *Bioinformatics: sequence, structure, and databanks: a practical approach*. Oxford University Press, Inc.

7. Jeremy M Berg, John L Tymoczko, Lubert Stryer, Gregory J Gatto Jr. (2007). *Biochemistry*. W H Freeman and company.
8. Jiang, T., Xu, Y., & Zhang, M. Q. (Eds.). (2002). *Current topics in computational molecular bi-ology*. MIT Press
9. Krane, D. E. (2003). *Fundamental concepts of bioinformatics*. Pearson Education India.
10. Lam, Y. W. F., & Cavallari, L. H. (Eds.). (2013). *Pharmacogenomics: Challenges and Opportu-nities in Therapeutic Implementation*. Academic Press.
11. Propst, C. L., & Perun, T. (1989). *Computer-aided drug design: methods and applications*. Marcel Dekker, Inc.
12. Reddy, M. R., & Erion, M. D. (Eds.). (2001). Free energy calculations in rational drug design.
13. Robert J Brooker (2009). *Genetics: analysis & principles* (III Edn). McGraw Hill.
14. S B Primrose, R M Twyman (2006). *Principles of gene manipulation and genomics* (VII Edn). Blackwell publishing.
15. T A Brown (2002). *Genomes* (II Edn). Bios.



	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>				
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Methods in Research, Biophysics and Biological Techniques</b>				
<b>Type of Course</b>	<b>DCE</b>				
<b>Course Code</b>	<b>M24BO8DCE401</b>				
<b>Course Level</b>	<b>400</b>				
<b>Course Summary</b>	This course provides the students the principles and applications of modern techniques used for research in biological sciences. The course prepares the students for roles in both research and professional settings.				
<b>Semester</b>	8	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		3	-	1	75
<b>Pre-requisites, if any</b>	The student must have completed courses in cell biology, biochemistry and plant physiology.				

#### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PSO No</b>
1	Discuss the principle and working of advanced microscopes used plant science research.	U	3
2	Illustrate the biological techniques employed in advanced research in plant science	A	3,5
3	Apply the expertise in experimental techniques and imaging methods	A,S	3,4,5
4	Develop skill in scientific research and communication	A,S	4, 5.6
5	Employ ethical considerations and safety measures in biological research	A	2,6

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hours	CO No.
1	<b>Module 1 (15 hours)</b>			
	1.1	Principles of microscopy- Types of microscopes: Optical, electron, and fluorescence microscopes. Image analysis software: ImageJ (brief account)	3	1,3
	1.2	Fluorescence Microscopy: Principles of fluorescence and fluorochromes. Applications in cell biology: Live cell imaging, immunofluorescence. Principles of Excitation emission and fluorophore selection. Commonly used fluorescent dyes. Confocal microscopy, FRET.	6	1
	1.3	Electron Microscopy: Transmission and scanning electron microscopy. Sample preparation techniques: Fixation, embedding, sectioning, Applications of Fluorescence Microscopy: Chromosome analysis: Banding techniques. Fluorescence in situ hybridization (FISH) Live cell imaging, super resolution microscopy	6	1
2	<b>Module 2 (15 hours)</b>			
	2.1	Centrifugation: Differential and density gradient centrifugation: Techniques for separating cellular components. Sucrose density gradient and CsCl <sub>2</sub> gradient centrifugation.	3	2
	2.2	Chromatography: Principles and applications. Types – GC, HPLC, HPTLC	3	2,3
	2.3	Spectroscopy: Principles and applications. UV-Visible spectrophotometer. Infra-red spectroscopy. FTIR, NMR and AAS (Brief account). Mass spectrometry: Principles and applications. GC-MS, LC-MS, MALDI-TOF (Brief account)	5	2,3
	2.4	Electrophoresis: Agarose gel electrophoresis, SDS-PAGE, Pulse field gel electrophoresis. Immunoassay systems, RIA, ELISA - ELISA reader.	4	2,3
3	<b>Module 3 (15 hours)</b>			
	3.1	Scientific research: Need for research, objectives of research, types of research. An outline of the steps in the research process. Constructing Research Questions, Methods: FINER criteria, PICOT criteria.	3	4
	3.2	Literature Survey: Sources of information- Journals, types of journals, journal abbreviations, Impact factor of journals; h-Index. Biological abstracts,	3	4



		Monographs. Web resources: E-journals, e-books, Google Scholar, Wiki- Databases, Science Direct, PubMed, Scopus. Finding and citing published information, Reference management software- Mendeley, EndNote, Zotero.		
	3.3	Science communication: Journal Article writing, Styles and formats, Title, Abstract, Key Words, Introduction, Materials and methods, Results, Figures & Tables, Discussion, List of References. Reference styles – APA, MLA, Chicago, Vancouver. Review articles, types of review. Oral Presentations. Poster Presentations. Writing project proposals to funding agencies.	4	4
	3.4	Ethics in research: Bioethics, Scientific misconduct, Ghost writing, Ethics in reporting research: data errors, omissions, redundancy, plagiarism. Checking documents for plagiarism, software. Conflict of Interest (COI) in academic publishing. Intellectual property rights (IPR): Copy right and patenting-Brief account.	3	4,5
	3.5	Safety in the laboratory: Biosafety level (BSL), Occupational safety and Health (OSH), General Safety and lab-safety procedures, Chemical, electrical and UV safety, safe handling of toxic and hazardous chemicals, storage and disposal of chemicals.	2	5
	<b>Module 4 Practical (30 hours)</b>			
	4.1	Collect and evaluate micrographs from different types of microscopes	3	1
	4.2	Estimate concentration biomolecules using spectrophotometer	4	2
	4.3	Separation of DNA samples using agarose gel electrophoresis	5	2
	4.4	Prepare a list of minimum 10 references in different citation styles	3	4
	4.5	Prepare an oral presentation based on a published research article	5	4
	4.6	Lab visit: Visit a well-established lab with advanced bioinstrumentation facility	10	1,2,3, 4,
<b>5</b>	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b>          Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b>  <b>A. Continuous Comprehensive Assessment (CCA)</b>          Theory Total = 25 marks          Quiz, Test Papers, seminar          Practical Total = 15 marks          Lab performance, record, field report etc.</p>
	<p><b>B. End Semester examination (ESE)</b>          Theory Total = 50 marks, Duration 1.5 hrs          Part A (Short answer) – 10 out of 12 x 1 = 10 marks          Part B (Short essay) – 4 out of 6 x 5 = 20 marks          Part C (Long essay) – 2 out of 4 x 10 = 20 marks          Practical Total = 35 marks; Duration- 2 hrs          Record 10 marks, Examination 25 marks</p>

## References


1. Alberts, B., et al. (2014). Molecular Biology of the Cell.
2. Berg, J. M., et al. (2015). Stryer's Biochemistry (8th ed.). W. H. Freeman.
3. Bowman, R. H., et al. (1970). Centrifugation: Practical Manual. American Elsevier Pub. Co.
4. Comas, I., & Schuenemann, V. J. (2018). A Brief Review of Molecular Archaeology.
5. Dawson Saunders, B., & Trapp, R. G. (1994). Basic & Clinical Biostatistics. Lange Medical Books/McGrawHill.
6. Drenth, J. (2007). Principles of Protein Xray Crystallography. Springer.
7. Ettore, L. S., & Snyder, L. R. (1976). Quantitative Paper Chromatography of Carbohydrates. Analytical Chemistry, 48(4), 586592.
8. Glantz, S. A. (2012). Primer of Biostatistics (7th ed.). McGrawHill.
9. Hayat, M. A. (2000). Principles and Techniques of Electron Microscopy: Biological Applications (4th ed.). Cambridge University Press.
10. Jürgen H. Gross (Ed.). (2005). Mass Spectrometry: A Textbook.
11. Miller, J. M. (2010). Chromatography: Concepts and Contrasts. John Wiley & Sons.
12. Murphy, D. B., & Davidson, M. W. (2012). Fundamentals of Light Microscopy and Electronic Imaging. Wiley.
13. Nelson, D. L., & Cox, M. M. (2008). Lehninger Principles of Biochemistry. W. H. Freeman.
14. Pawley, J. B. (2006). Handbook of Biological Confocal Microscopy (3rd ed.). Springer.
15. Plummer, D.T. (1996). An Introduction to Practical Biochemistry. Tata McGrawHill Publishing Co. Ltd. New Delhi. 3rd edition.

16. Richmond, R. C., & Sykes, G. (2004). *Isotopes in Biological Dinitrogen Fixation Research*. Springer.
17. Ruzin, S.E. (1999). *Plant Microtechnique and Microscopy*, Oxford University Press, New York. U.S.A.
18. Skoog, D. A., et al. (2017). *Fundamentals of Analytical Chemistry*. Cengage Learning.
19. Zar, J.H. (2012). *Biostatistical Analysis*. Pearson Publication. U.S.A. 4th edition.
20. Zubay, G., et al. (1995). *Principles of Biochemistry*. WCB/McGrawHill.

### **SUGGESTED READINGS**

1. Farago, M. E., & Mehra, A. (1994). 9 Analytical Techniques for Plant Analysis. *Plants and the Chemical Elements: Biochemistry, Uptake, Tolerance and Toxicity*, 253, 241.
2. Kalra, Y. P. (1998). *Methods for plant analysis*. CRC, USA, 85-88.
3. Garg, B. K. (2012). *Plant analysis: comprehensive methods and protocols*. Scientific Publishers.
4. Dhale, D. A. (2023). *Advanced Techniques in Plant Sciences*. Book Saga Publication



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<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>				
<b>Course Name</b>	<b>Phytochemistry and Pharmacognosy</b>				
<b>Type of Course</b>	<b>DCE</b>				
<b>Course Code</b>	<b>M24BO8DCE402</b>				
<b>Course Level</b>	<b>400</b>				
<b>Course Summary</b>	Phytochemistry is the study of the chemicals produced by plants, particularly the secondary metabolites which are synthesized as a measure for self-defense, and its medicinal, industrial, and commercial applications. Pharmacognosy is the study and science of medicine from natural sources. Natural medicines have been used for many thousands of years to enhance human health and treat diseases, and modern pharmaceutical medicine is largely dependent on drugs originally discovered in and isolated from natural sources.				
<b>Semester</b>	8	Credits			Total Hours
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical	Others
		3	-	1	-
<b>Pre-requisites, if any</b>	Knowledge about secondary metabolites present in plants				

### **COURSE OUTCOMES (CO)**

<b>CO No.</b>	<b>Expected Course Outcome</b>	<b>Learning Domains *</b>	<b>PO No</b>
1	Describe the importance of phytochemicals and pharmaceutical drugs.	U	2,5
2	Explain the principle involved in the extraction and isolation techniques.	U	4
3	Classify the different phytochemicals and pharmaceutical drugs.	A	3,4
4	Execute various phytochemical tests and procedures using different laboratory equipment.	An	3,4,5
5	Evaluate various drugs and estimate the presence of phytochemicals and adulterants present in pharmaceutical drugs.	E	4

**\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)**

## COURSE CONTENT

Module	Units	Course description	Hrs	CO No.
<b>1</b>	<b>Module 1 (15 hours)</b>			
	1.1	Definition, history and scope of Phytochemistry.	15	1,2,3
	1.2	Plant kingdom as source of drugs- plant secondary metabolites as drugs;		
	1.3	Brief study of basic metabolic pathways and formation of different secondary metabolites through these pathways- Shikimic acid pathway, Acetate pathways and Amino acid pathway.		
	1.4	Classification, structure and function of medicinally important plant products: glycosides, tannins, alkaloids, phenolic compounds, saponins, terpenoids, steroids, flavonoids, gums and mucilage.		
<b>Module 2 (15 hours)</b>				
<b>2</b>	2.1	Solvents- Petroleum ether, Chloroform, Ethanol, Acetone, Water;	15	2,3,4
	2.2	Extraction techniques- Cold extraction, Hot extraction, Soxhlet- Clavenger apparatus		
	2.3	Separation techniques- TLC, Column Chromatography, HPLC; Characterization techniques- GC-MS, LC-MS/MS, UV-VIS Spectrometry, IR Spectrometry		
	2.4	Quality control of drugs- Adulteration of drugs, tools for identification.		
	<b>Module 3 (15 hours)</b>			
<b>3</b>	3.1	Pharmacognosy: Definition, history, scope and development	15	1,4,5
	3.2	Plants in Medicine: Indigenous traditional drugs, traditional system of medicine, herbal medicine, folk medicine, unani, siddha, ayurveda, homoeopathy and Chinese medicine.		
	3.3	Ethnopharmacology		
	3.4	Techniques for production of drugs– purification, filtration, adsorption, solubilization, absorption, suspension and emulsification;		
	3.5	Therapeutic classification of crude drugs. Morphological, microscopical and organoleptic evaluation of crude drugs Drug preparation and storage. Collection and preparation of crude drugs for the market		

<b>Module 4 Practical (30 hours)</b>			
<b>4</b>	4.1	Extraction methods – Soxhlet / Clevenger	30 2,3,4,5
	4.2	TLC and column chromatography (Demonstration/ Activity).	
	4.3	Qualitative analysis of tannins, phenolics, flavonoids and alkaloids (Activity).	
	4.4	Histochemical localization of starch grains- rice, potato.	
	4.5	Histochemical analysis of plant components: Starch grains in rice and potato	
	4.6	Estimation of water content, dry matter and ash content	
	4.7	Interaction with subject expert in the field of Ayurvedic medicine / Ayurvedic or plant extract industry visit	
<b>5</b>	<b>Teacher specific course components</b>		


<b>Teaching and Learning Approach</b>	<p><b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.</p>
<b>Assessment Types</b>	<p><b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.</p>
	<p><b>B. End Semester examination (ESE)</b> Theory Total = 50 marks, Duration 1.5 hrs Part A (Short answer) – 10 out of 12 x 1 = 10 marks Part B (Short essay) – 4 out of 6 x 5 = 20 marks Part C (Long essay) – 2 out of 4 x 10 = 20 marks Practical Total = 35 marks; Duration- 2 hrs Record 10 marks, Examination 25 marks</p>

## References

1. Arumugam, K. R. & Murugesh (2005) Textbook of Pharmacognosy. Sathya Publishers, Madurai.
2. Biren, N. S. & Seth, A. K (2014) Textbook of Pharmacognosy and Phytochemistry. Elsevier Science Publishing Company. Inc
3. Daniel, M., & Daniel, M. (1991). *Methods in plant chemistry and Economic Botany*. Kalyani Publishers.
4. Dwivedi, J. N., Dwivedi, J. N., & Singh, R. B. (1990). *Essentials of plant techniques*. Scientific Publishers.
5. Gibbs, R. D. (1974). *Chemotaxonomy of Flowering Plants: Vol. 1 Constituents*. Queens University Press.
6. Jain, S. K. (1991). *Dictionary of Indian folk medicine and ethnobotany*.
7. Khandelwal, K. (2008). *Practical pharmacognosy*. Pragati Books Pvt. Ltd..
8. Miller, L. P (1973) *Phytochemistry Vol. I, II & III*. Van Nostrand Reinhold Co., New York.
9. Sabnis, S. D., & Daniel, M. (1990). A phytochemical approach to economic Botany. *Kalyani Pub. New Delhi*, 108-109.
10. Shirkhedkar, A., & Surana, D. S. (2008). *Pharmacognosy and Phytochemistry*.
11. Syed, A. I., & Khan, M. A. (2004) *Textbook of Phytochemistry*. Discovery Publishing. New Delhi.
12. Vasishta, P. C., & Gills, P. S. (1995) *Ethnobotany*. Pradeep Publications, Jalandhar.

## SUGGESTED READINGS

1. Harborne, J. B. (1973) *Phytochemical Methods*. Chapman and Hall Limited, London.
2. John, T., & Romeo (2006) *Recent Advances in Phytochemistry*. Elsevier Science Publishing Company Inc.
3. Trease, G. E. & Evans, W.C. (2002) *Pharmacognosy*. Collis Macmillan Publishers, Madras.

	<b>Mar Athanasius College (Autonomous), Kothamangalam</b> <b>FYUGP SYLLABUS</b>					
<b>Programme</b>	<b>B. Sc. (Honours) Botany</b>					
<b>Course Name</b>	<b>Aquatic Botany</b>					
<b>Type of Course</b>	<b>DCE</b>					
<b>Course Code</b>	<b>M24BO8DCE403</b>					
<b>Course Level</b>	<b>400</b>					
<b>Course Summary</b>	This syllabus aims to cover key aspects of aquatic botany, providing students with a comprehensive understanding of the diversity, ecology, and conservation of plants in aquatic environments.					
<b>Semester</b>	8	Credits			Total Hours	
<b>Course Details</b>	Learning Approach	Lecture	Tutorial	Practical		Others
		3	-	1	-	75
<b>Pre-requisites, if any</b>	Knowledge about aquatic ecosystems, plant morphology, physiology and anatomy.					

#### COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains *	PSO No
1	Acquire comprehensive understanding of aquatic ecosystems, including physiochemical properties, flora and biological productivity.	U,A,An	1,2
2	Acquire skills in identifying and classifying aquatic plants and their ecology	S, U, A	1,2
3	Analyse different types of water pollution, understand their sources and propose effective management and conservation strategies.	S, U, A	2
4	Acquire knowledge and develop and understanding of the physiology and adaptations in aquatic plants	U,A,An	2,3
5	Recognize threats to aquatic plant biodiversity and implement conservation strategies considering factors like climate change, aquaculture and habitat degradation.	U, A, E,	2,5

*\*Remember (K), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

#### COURSE CONTENT



Module	Units	Course description	Hours	CO No.
<b>Module 1 (15 hours)</b>				
<b>1</b>	1.1	Overview of Aquatic Ecosystems Fresh water- Lentic ecosystem and Lotic Ecosystem Rivers and Ponds: Physicochemical properties. Riparian flora, Biological productivity. Concept of watershed and watershed management Swamps and marshes: Types of swamps. Physicochemical conditions. Nutrient cycling. Lakes and reservoirs: Characteristics and stratification. Marine- definition, range of salinity, stratification Mangroves and Estuaries	5	1,3
	1.2	Identification and Classification of Aquatic Plants Classification based on growth form--freshwater higher vascular plants-floating (rooted and free floating), submerged and emerged, sea weeds, sea grasses and mangroves, invasive aquatic plants. Classification based on morphology-amphiphytes, Helophytes, nymphaeids, Isoetids, neuston etc. Micro and Macro algae: distribution and importance. Seaweeds and Seagrasses: structure, types and economic importance	6	2,3
	1.3	Functions of aquatic ecosystems. Importance in nutrient cycling, impact of soil chemistry and role in soil chemistry. Dynamics of plant aquatic community, common aquarium plants	4	1,4
<b>Module 2 (15 hours)</b>				
<b>2</b>	2.1	Water pollution: types- Thermal pollution, Oil pollution, Radioactive pollution, Detergent pollution, interaction of pollutants and factors affecting toxicity Sources: Characteristics of effluent sewage, industrial and agricultural discharges. Emerging pollutants, heavy metals, pesticides, plastic and microplastics, oil spills, etc.	5	3,4
	2.2	Biological concern: Eutrophication (change in the plant diversity in aquatic systems, change in DO levels), algal blooms, bioaccumulation and biomagnification, change in water quality ( BOD, COD, DO ), monitoring and control of pollutants, effect of waste disposal on marine ecosystem.	6	3,4
	2.3	Sustainable Management of Aquatic Environments. Conservation of freshwater ecosystems. Restoration of freshwater wetlands. Ramsar convention, Ramsar sites, Role of Pollution Control Board (PCB) Conservation of Mangroves: need, Impact of human, role of institutions and NGO's in India	4	3,5

	<b>Module 3 (15 hours)</b>			
3	3.1	Threats to Aquatic Plant Biodiversity: Climate change, Harmful aspects related to aquaculture activities, introduction of exotic species, destruction of mangroves, Expanding hydropower etc	5	5
	3.2	Conservation Strategies for Aquatic Plants: Conservation of freshwater ecosystems, habitat restoration ecology, Habitat protection, wetland conservation, riparian buffer zones, invasive species management.	5	5
	3.3	Physiology and Adaptations in Aquatic plants. Fine structure and properties of algal plastids. Morphological and anatomical modifications in aquatic plants. Physiological adaptations in mangroves.	5	4
	<b>Module 4 Practical ( 30 hours)</b>			
4	4.1	Collect common aquatic plants and identify	5	2
	4.2	Collect aquatic plants and plants from mangroves and conduct anatomical studies to understand anatomical adaptations	5	2
	4.3	Field visit to observe and identify aquatic ecosystems	10	1,5
	4.4	Conduct water quality analysis with samples collected from different aquatic ecosystems using titrimetric methods	3	3
	4.5	Visit mangroves to understand the ecological significance and the need for restoration activities	7	5
5	<b>Teacher specific course components</b>			

<b>Teaching and Learning Approach</b>	<b>Classroom Procedure (Mode of transaction)</b> Field based collection and interactions, Interactive lectures, flipped classroom, Lecture-based Learning, Project-Based Learning, Experiential Learning, Peer Teaching, invited lecture, Discussion-based Learning, Inquiry-Based Learning, Online Learning, Blended Learning, and other innovative learning approaches.
<b>Assessment Types</b>	<b>MODE OF ASSESSMENT</b> <b>A. Continuous Comprehensive Assessment (CCA)</b> Theory Total = 25 marks Quiz, Test Papers, seminar Practical Total = 15 marks Lab performance, record, field report etc.

	<p><b>B. End Semester examination (ESE)</b>  Theory Total = 50 marks, Duration 1.5 hrs  Part A (Short answer) – 10 out of 12 x 1 = 10 marks  Part B (Short essay) – 4 out of 6 x 5 = 20 marks  Part C (Long essay) – 2 out of 4 x 10 = 20 marks  Practical Total = 35 marks; Duration- 2 hrs  Record 10 marks, Examination 25 marks</p>
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### References

1. Bolton, J. J. (2016). What is aquatic botany?—And why algae are plants: The importance of non-taxonomic terms for groups of organisms. *Aquatic Botany*, 132, 1-4.
2. Barnes, R. S. K., & Mann, K. H. (Eds.). (2009). *Fundamentals of aquatic ecology*. John Wiley & Sons.
3. Cherry, J. A., & Pec, G. J. (2022). Advances, applications, and prospects in aquatic botany. *Applications in Plant Sciences*, 10(4).
4. Doležal, J., Kučerová, A., Jandová, V., Klimeš, A., Říha, P., Adamec, L., & Schweingruber, F. H. (2021). Anatomical adaptations in aquatic and wetland dicot plants: disentangling the environmental, morphological and evolutionary signals. *Environmental and Experimental Botany*, 187, 104495.
5. Jones, J. I., Li, W., & Maberly, S. C. (2003). Area, altitude and aquatic plant diversity. *Ecography*, 26(4), 411-420.
6. Goel, P.K. (2006). Water pollution, New age international publishers, New Delhi. · Kukul S.S. and Dhaliwal, G.S. (2005). Essential of environmental science, Kalyani Publishers, Ludhiyana